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(54) Title: **ROLES OF JAK/STAT FAMILY MEMBERS IN TOLERANCE INDUCTION**

(57) Abstract: The present invention relates to methods and compositions for reducing immune rejection, for example, transplant- or autoimmune disorder-related immune rejection. The present invention also relates to methods and compositions for monitoring transplant acceptance and for monitoring an autoimmune disorder in a subject mammal. The present invention still further relates to methods for identifying compounds that can reduce immune rejection. The present invention is based, in part, on the discovery, demonstrated herein, that immune rejection can be monitored by determining the amount of particular members of the Jak/Stat signal transduction pathway present within an affected tissue (that is, a transplant cell, tissue, organ, or organ system, or a cell, tissue, organ, or organ system that is, or is suspected of, being affected by an autoimmune disorder). The present invention is further based, in part, on the discovery, demonstrated herein, that immune rejection can be reduced and tolerance can be induced by modulating the amount of these particular members of the Jak/Stat signal transduction pathway present, expressed or active within an affected tissue. In particular, the results presented herein demonstrate that immune rejection can be monitored by determining the amount of Stat1 mRNA or protein, Stat2 mRNA or protein, Stat3 mRNA or protein, Stat4 mRNA or protein, Stat6 mRNA or protein, SOCS1 mRNA or protein, or SOCS3 mRNA or protein present, e.g., present in an affected tissue.

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Roles of Jak/Stat Family Members in Tolerance Induction

1. INTRODUCTION

5 The present invention relates to methods and compositions for reducing immune rejection, for example, transplant- or autoimmune disorder-related immune rejection. The present invention also relates to methods and compositions for monitoring transplant acceptance and for monitoring an autoimmune disorder in a subject mammal. The present invention still further relates to methods for identifying compounds that can reduce immune
10 rejection.

 The present invention is based, in part, on the discovery, demonstrated herein, that immune rejection can be monitored by determining the amount of particular members of the Jak/Stat signal transduction pathway present within an affected tissue (that is, a transplant cell, tissue, organ, or organ system, or a cell, tissue, organ, or organ system that is, or is
15 suspected of, being affected by an autoimmune disorder). The present invention is further based, in part, on the discovery, demonstrated herein, that immune rejection can be reduced and tolerance can be induced by modulating the amount of these particular members of the Jak/Stat signal transduction pathway present, expressed or active within an affected tissue. In particular, the results presented herein demonstrate that immune rejection can be
20 monitored by determining the amount of Stat1 mRNA or protein, Stat2 mRNA or protein, Stat3 mRNA, protein Stat4 mRNA or protein, Stat6 mRNA or protein, SOCS1 mRNA or protein, or SOCS3 mRNA or protein present, *e.g.*, present in an affected tissue.

2. BACKGROUND OF THE INVENTION

25 Ongoing advances in transplantation, including new immunosuppressive agents and improvements in histocompatibility matching, organ procurement, and surgical techniques, are gradually improving the outcome of clinical transplantation (Hariharan et al, 2000. N Engl J Med 342:605-12). However, chronic allograft rejection remains the prime determinant of long-term graft survival (Paul. L.C., 1999, Kidney International 56:783-
30 793).

 Tissue transplantation between genetically nonidentical individuals results in immunological rejection of the tissue through T cell-dependent mechanisms. To prevent allograft rejection, immunosuppressive agents such as calcineurin phosphatase inhibitors and glucocorticosteroids which directly or indirectly interfere with IL-2 signaling are
35 administered to transplant recipients (see, *e.g.*, Borel, J.F., 1989, Pharmacol. Rev. 42:260-372; Morris, P.J., 1991, Curr. Opin. Immunol. 3:748-751; Sigal et al., 1992, Ann. Rev. Immunol. 10:519-560; and L'Azou et al., 1999, Arch. Toxicol. 73:337-345). The most

commonly used immunosuppressive agents today are cyclosporin A, FK506, and rapamycin. These immunosuppressive agents act indiscriminately on all T cells by impairing T cell receptor ("TCR") signal transduction. Further, since the effect of the immunosuppressive agents is short-lasting, transplant recipients normally require life-long
5 treatment of immunosuppressive agents to prevent transplant rejection. As a result of the long-term nonspecific immunosuppression, these immunosuppressive agents have many serious adverse effects. For example, the administration of cyclosporin A or FK506 to a transplant recipient results in degenerative changes in renal tubules. Transplant recipients receiving long-term immunosuppressive treatment have a high risk of developing infections
10 and tumors. For example, patients receiving immunotherapy are at higher risk of developing lymphomas, skin tumors and brain tumors (see, *e.g.*, Fellstrom et al., 1993, Immunol. Rev. 134:83-98).

An alternative to immunosuppressive agents for the prevention of allograft rejection is the blockage of specific receptors involved in T cell costimulation. T cell activation
15 requires both TCR-mediated signal transduction and simultaneously delivered costimulatory signals. These costimulatory signals are contributed, in part, by the activation of the costimulatory molecule CD28, which is expressed on resting T cells, by CD80 (B7-1) or CD86 (B7-2) expressed on antigen presenting cells (APCs). The activation of the costimulatory molecule CD40, which is expression on antigen presenting cells (*i.e.*, B cells,
20 dendritic cells, and macrophages), by CD40 ligand ("CD40L"), which is expressed on activated T cells, contributes to the upregulation of T cell activation by inducing the expression of B7-1 and B7-2 on antigen presenting cells and the production of certain chemokines and cytokines such as IL-8, MIP-1 α , TNF- α , and IL-12 (Cella et al., 1996, J. Exp. Med. 184:747-752; and Caux et al., 1994, J. Exp. Med. 180:1263-1272). The
25 CD40/CD40L interaction also results in the differentiation of T cells to T helper ("TH") type 1 cells in part due to the expression of cytokines such as IL-12 by dendritic cells and macrophages.

CTLA-4 is normally expressed as a membrane-bound receptor on T cells and has been shown to downregulate T cell activation by competing with CD28 for B7-1 and B7-2.
30 The administration of soluble CTLA-4Ig is believed to prevent allograft rejection by competing with CD28 for B7-1 and B7-2. Soluble CTLA-4Ig has been administered to transplant recipients to disrupt the CD28/B7 interaction so that T cell costimulation is blocked and allograft rejection does not occur (Zheng et al., 1999, J. Immunol. 162:4983-4990; Lenschow et al., 1996, Ann. Rev. Immunol. 14:233-258). Unfortunately, CTLA-4Ig
35 has variable efficacy, and typically does not prevent development of chronic rejection.

Anti-CD40L (anti-CD154) monoclonal antibodies have also been administered to transplant recipients to prevent allograft rejection. These antibodies function by blocking the interaction of CD40 on antigen presenting cells (APC) and CD40L on activated T cells. It has recently been shown that graft survival achieved through the use of anti-CD40L monoclonal antibodies results in a significant inhibition of TH1 type cytokines (*i.e.*, IL-2, IL-12, TNF α , and IFN γ), and an increase in the levels of the TH2 type cytokines (*i.e.*, IL-4, and IL-10) in the graft sections (Hancock et al., 1996, Proc. Natl. Acad. Sci. USA 93:13967-13972). Although the administration of anti-CD40L monoclonal antibodies has been shown to result in permanent graft survival when given to mice in combination with donor-specific spleen cells, adverse side effects such as coagulation have also been shown to be associated with the administration of anti-CD40L monoclonal antibodies. Initial clinical trials in adult renal transplant recipients receiving anti-CD40L monoclonal antibody plus glucocorticoids were halted because of thromboembolic complications (Vincent, J., Biogen News, press release, November 2, 1999, www.prnewswire.com), though the extent to which thromboembolism was attributable to monoclonal antibodies versus non-specific factors in the antibody formulation is unclear (Kawai et al., 2000, Nature Med. 6:114; and Kirk et al., 2000, Nature Med. 6:114). Further, in the primate renal allograft study, concomitant use of mainstream immunosuppressive agents such as FK-506, methylprednisolone and mycophenolate mofetil diminished the efficacy of CD40L (CD154) mAb, though the exact contribution of each of the individual drugs to this reduction in efficacy was not determined (Kirk, A.D., 1999, Nature Medicine 5:686-693.). The results presented herein demonstrate that some, but not all, combinations of CD154 mAb and immunosuppressive agents are antagonistic, and that strategies for design of clinical trials based on use of CD154 mAb can be logically developed by taking into account the extent to which a given drug inhibits induction of CD154.

In addition, no satisfactory methods presently exist for monitoring whether a transplant graft is being accepted or rejected by a recipient. In general, signs of cellular damage within the transplant tissue can be assayed. Alternatively, for tissues such as kidney or liver, physiological function of the transplant tissue can be assayed. Often, however, by the time overt signs of either cellular damage or a decrease in physiological function are detected, the tissue graft is already beyond rescue. This is particularly true in the case of such organ transplants as heart transplants, with which the first overt signs of rejection are often complete failure of the heart's function.

Accordingly, there is a need for improved, safer immunomodulatory treatments that have long-lasting effects for the prevention of transplant rejection. In particular, there is a need for treatments that are more specific and less toxic than the currently available

factors. For example, Stat4 has been shown be activated in response to IL-12 induced signal transduction (Thierfelder et al., 1996, Nature 382:171-174; and Kaplan et al., 1996, Nature 382:174-177). Stat6 has been shown to be activated in response to IL-4 and IL-13 induced signal transduction (Takeda et al., 1996, Nature 380:627-630). Certain

transcription factors activated in response to a given cytokine have been shown to be important in TH1 and/or TH2 differentiation. Stat4 has been shown to be important in TH1 differentiation and Stat6 has been shown to be important in TH2 differentiation (see, *e.g.*, Romagnani, S., 1997, Immunology Today 18:263-266; Ray, A. and Cohn, L., 1999, J. Clin. Invest. 104(8):985-993).

With respect to TH1 and TH2, the majority of mature T lymphocytes can be divided into two distinct phenotypes: CD8⁺ cytotoxic T lymphocytes (CTLs), which display the CD8 marker on their cell surface, and CD4⁺ helper T lymphocytes (T helper or TH cells), which display the CD4 marker on their cell surface. TH cells are involved in both humoral (*i.e.*, antibody) and cell-mediated forms of immune response. TH cells have been further categorized into two distinct subpopulations, termed TH1 and TH2 cell subpopulations. These two subpopulations of TH cells have been categorized on the basis of their restricted cytokine profiles and different functions. For example, TH1 cells are known to produce IL-2, IL-12, tumor necrosis factor β ("TNF- β "), and interferon- α ("IFN- α "). TH2 cells are known to produce IL-4, IL-5, IL-10 and IL-13. Inappropriate immune responses have been shown to be associated with various diseases and disorders. For example, an inappropriate TH2-like response has shown to be associated with atopic conditions, such as asthma and allergy (see, *e.g.*, Holgate, S.T., 1997, Lancet 350(suppl. II):5-9; Ray, A. and Cohn, L, *supra*; Oettgen, H.C. and Geha, R.S., 1999, J. Clin. Invest. 104(7):829-835). Further, an inappropriate TH1-like response has been shown to associated with the pathogenesis of autoimmune diseases such multiple sclerosis, pancreases of insulin-dependent diabetes patients, thyroid glands of Hashimoto's thyroiditis, and gut of Crohn's disease patients.

2.2. NEGATIVE REGULATORS OF THE Jak/STAT SIGNALING PATHWAY

Three protein families have been discovered that negatively regulate cytokine-induced Jak/Stat signaling, tyrosine phosphatases SHP1 and SHP2, the suppressors of cytokine signaling ("SOCS"), and protein inhibitors of activated Stats (PIAS). SHP1 and SHP2 bind to phosphorylated tyrosine residues on receptors or Jaks, and inactivate signaling by dephosphorylating them (Haque et al., 1998, J. Biol. Chem. 273:33898-33896; and You et al., 1999, Mol. Cell. Biol. 19:2416-2424).

The SOCS family of proteins have been shown to inhibit the Jak/Stat pathway by inhibiting the activity of the Jaks (Hilton et al., Proc. Natl. Acad. Sci. USA 95:114-119; and

Hilton, 1999, Cell. and Mol. Life Sci. 55:1658-1577). The nature of the interaction between the different receptors, Jaks, and the SOCS is unclear (Hilton, D. J., 1999, Cell. Mol. Sci. 55:1568-1577). SOCS1 have been shown to directly interact with all the Jaks and Tyk2. CIS (Cytokine inducible SH2 containing protein), a member of the SOCS family, on the other hand, was shown to interact with the EPO receptor or the β chain of the IL-3 receptor in a phosphorylation dependent manner, indicating it may act by competing with Stat molecules for binding to receptors (Yoshimura et al., 1995, EMBO J. 14:2816-2826). SOCS1 expression inhibits IL-6, LIF, oncostatin M, IFN- γ , IFN- β , IFN- α , thrombopoietin, and growth hormone (GH) induced Jak/Stat signaling. SOCS3 expression inhibits IFN- γ , IFN- β , IFN- α , GH and leptin.

Four members of the PIAS family have been identified, PIAS1, PIAS3, PIASx α , and PIASx β . PIAS1 was found to bind only to activated Stat1, and PIAS3 to only activated Stat3 (Liu et al., 1998, Proc. Natl. Acad. Sci. USA 95:10626-10631; and Chung et al., 1997, Science 278:1803-1805). PIAS-mediated inhibition of the Jak/Stat signaling pathway, unlike SOCS-mediated inhibition of the Jak/Stat signaling pathway, is very specific. However, unlike some of the SOCS which are elevated rapidly in response to cytokines, the PIAS levels in the cells are more or less constant.

3. SUMMARY OF THE INVENTION

The present invention relates to methods and compositions for reducing immune rejection, for example, transplant- or autoimmune disorder-related immune injury or rejection. The present invention also relates to methods and compositions for monitoring transplant acceptance and for monitoring an autoimmune disorder in a subject mammal. The present invention still further relates to methods for identifying compounds that can reduce immune injury.

The present invention is based, in part, on the discovery, demonstrated herein, that immune rejection can be monitored by determining the amount of particular members of the Jak/Stat signal transduction pathway present within an affected tissue (that is, a transplant cell, tissue, organ, or organ system, or a cell, tissue, organ, or organ system that is, or is suspected of, being affected by an autoimmune disorder). In particular, the results presented herein demonstrate that immune rejection can be monitored by determining the amount of Stat4 mRNA or protein, Stat6 mRNA or protein, SOCS1 mRNA or protein, or SOCS3 mRNA or protein, present in an affected tissue. The results presented herein also demonstrate that immune rejection can be monitored by determining the amount of Stat1 mRNA or protein, Stat2 mRNA or protein, or Stat3 mRNA or protein present, *e.g.*, present in an affected tissue. The present invention is further based, in part, on the discovery,

demonstrated herein, that immune rejection can be reduced and tolerance can be induced by modulating the amount of these particular members of the Jak/Stat signal transduction pathway present, expressed or active within an affected tissue.

Thus, in one aspect, the invention relates to methods for monitoring acceptance of a
5 transplant in a subject mammal that has undergone a transplant, wherein said method comprises: determining the amount of at least one of the following: (i) Stat4 mRNA or Stat4 protein, (ii) Stat6 mRNA or Stat6 protein, (iii) SOCS1 mRNA or SOCS1 protein, or (iv) SOCS3 mRNA or SOCS3 protein, present in a transplant sample from the subject. In alternate embodiments, such methods comprise determining the amount of at least two, at
10 least three, or each of (i) to (iv) present in the transplant sample. In certain embodiments, the amount of mRNA is determined, and can, for example, be determined via use of nucleic acid microarrays. In other embodiments, the amount of protein is determined, while in still other embodiments, the amount of mRNA and protein is determined. With respect to Stat6, when the amount of Stat6 is being determined, it is preferable that the amount of Stat6
15 protein be determined. In any such embodiment wherein a Stat protein amount is determined, the amount determined can be the total amount of the Stat protein present in a sample or, alternatively, can be the amount of phosphorylated Stat protein present in the sample.

In a preferred embodiment, a method for monitoring acceptance of a transplant in a
20 subject mammal that has undergone a transplant comprises determining the amount of Stat4 and Stat6 mRNA or Stat4 and Stat6 protein present in a transplant sample from the subject. Such an embodiment can further comprise determining the ratio of Stat4 to Stat6 amounts.

The methods for monitoring acceptance of a transplant in a subject mammal that has undergone a transplant can further comprise assaying the transplant sample for evidence of
25 lymphocyte infiltration or tissue damage (cell injury) using standard techniques. For example, histological techniques well known to those of skill in the art can be utilized to evaluate internationally recognized and used diagnostic criteria for the evaluation of graft rejection, which include features specific for each organ involved. For example, immunohistologic evaluation of such tissues, via, *e.g.*, use of labeled antibody techniques
30 to localize and quantitate gene expression. The evaluation of such criteria can, therefore, be enhanced by, for example, localization of Stat4, Stat6, SOCS1 and/or SOCS3 proteins, and/or detection of corresponding mRNAs via, *e.g.*, *in situ* hybridization.

Such methods can also further comprise comparing the amount or ratio determined to that present in a control sample, for example, a corresponding pre-transplant subject
35 sample or a subject blood sample. In instances wherein the amount of Stat4, SOCS1, or SOCS3 mRNA or protein in the transplant sample is greater than, or the amount of Stat6

mRNA or protein in the transplant sample is less than, that of the control sample, such a result indicates that acceptance of the transplant has not occurred, has not been induced or is not being maintained. In instances wherein the amount of Stat4, SOCS1, or SOCS3 mRNA or protein in the transplant sample is less than, or the amount of Stat6 mRNA or protein in the transplant sample is equal to or greater than that of the control sample, such a result indicates that acceptance of the transplant has occurred, is being induced or is being maintained. In instances wherein the ratio of Stat4 to Stat6 in the transplant sample is greater than or equal to that in the control sample, such a result indicates that acceptance of the transplant has not occurred, has not been induced or is not being maintained. In instances wherein the ratio of Stat4 to Stat6 in the transplant sample is less than that in the control sample, such a result indicates that acceptance of the transplant has occurred, has been induced or is being maintained.

In another aspect, the invention relates to methods for monitoring acceptance of a transplant in a subject mammal that has undergone a transplant, wherein said method comprises: determining the amount of at least one of the following: (i) Stat1 mRNA or Stat1 protein, (ii) Stat2 mRNA or Stat2 protein, or (iii) Stat3 mRNA or Stat3 protein, present in a cell sample from the subject. In alternate embodiments, such methods comprise determining the amount of at least two or each of (i) to (iii) present in the sample. In certain embodiments, the amount of mRNA is determined, and can, for example, be determined via use of nucleic acid microarrays. In other embodiments, the amount of protein is determined, while in still other embodiments, the amount of mRNA and protein is determined. In any such embodiment wherein a Stat protein amount is determined, the amount determined can be the total amount of the Stat protein present in a sample or, alternatively, can be the amount of phosphorylated Stat protein present in the sample.

In a particular embodiment of such Stat 1-, Stat 2-, and/or Stat 3-related methods, the cell sample is a transplant sample obtained within 2 to 3 days post-transplantation. In an alternative embodiment of such Stat 1-, Stat 2-, and/or Stat 3-related methods, the cell sample is a subject blood sample.

Such Stat 1-, Stat-2, and/or Stat 3-related methods can also further comprise comparing the amount determined to that present in a control sample, for example, a corresponding pre-transplant subject sample or, in the case of embodiments wherein the cell sample is a transplant sample obtained within 2-3 days post-transplantation, a subject blood sample. In instances wherein the amount of Stat1, Stat2, or Stat3 mRNA or protein in the cell sample is greater than that of the control sample, such a result indicates that acceptance of the transplant has not occurred, has not been induced or is not being maintained. In instances wherein the amount of Stat1, Stat2, or Stat3 mRNA or protein in the transplant

sample is less than that of the control sample, such a result indicates that acceptance of the transplant has occurred, is being induced or is being maintained.

In another aspect, the invention relates to methods for monitoring an autoimmune disorder in a subject mammal, wherein said method comprises: determining the amount of
5 at least one of the following: (i) Stat4 mRNA or Stat4 protein, (ii) Stat6 mRNA or Stat6 protein, (iii) SOCS1 mRNA or SOCS1 protein, or (iv) SOCS3 mRNA or SOCS3 protein, present in a sample from a subject mammal being treated for or suspected of exhibiting the autoimmune disorder, wherein the sample is obtained from a tissue affected by the disorder. In alternate embodiments, such methods comprise determining the amount of at least two, at
10 least three, or each of (i) to (iv) present in the sample. In certain embodiments, the amount of mRNA is determined, and can, for example, be determined via use of nucleic acid microarrays. In other embodiments, the amount of protein is determined, while in still other embodiments, the amount of mRNA and protein is determined. With respect to Stat6, when the amount of Stat6 is being determined, it is preferable that the amount of Stat6 protein be
15 determined. In any such embodiment wherein a Stat protein amount is determined, the amount determined can be the total amount of the Stat protein present in a sample or, alternatively, can be the amount of phosphorylated Stat protein present in the sample.

In a preferred embodiment, a method for monitoring an autoimmune disorder in a subject mammal comprises determining the amount of Stat4 and Stat6 mRNA or Stat4 and
20 Stat6 protein present in a sample from the subject mammal being treated for or suspected of exhibiting the autoimmune disorder, wherein the sample is obtained from a tissue affected by the disorder. Such an embodiment can further comprise determining the ratio of Stat4 to Stat6 amounts.

The methods for monitoring an autoimmune disorder in a subject mammal can
25 further comprise assaying the sample for evidence of leukocyte infiltration or tissue damage (cell injury) using standard techniques. For example, histological techniques well known to those of skill in the art can be utilized. Alternatively, standard techniques can be utilized to assay (*e.g.*, in serum) for the presence of autoimmune antibodies associated with the particular autoimmune disorder of interest. There are internationally used diagnostic
30 criteria for evaluation of graft rejection, with features specific for each organ. The immunohistologic evaluation of such tissues, *i.e.*, use of unlabeled-antibody techniques to localize and quantitate gene expression, can be enhanced by localization of Stat4 and Stat6 proteins, or detection of corresponding mRNAs by *in situ* hybridization.

Such methods for monitoring an autoimmune disorder in a subject mammal can
35 further comprise comparing the amount or ratio determined to that present in a control sample, for example, a corresponding tissue not affected by the disorder or a subject blood

sample. In instances wherein the amount of Stat4, SOCS1, or SOCS3 mRNA or protein in the sample is greater than, or the amount of Stat6 mRNA or protein in the sample is less than, that of the control sample, such a result indicates that the subject mammal exhibits or continues to exhibit the disorder. In instances wherein the amount of Stat4, SOCS1, or SOCS3 mRNA or protein in the sample is less than, or the amount of Stat6 mRNA or protein in the sample is equal to or greater than that of the control sample, such a result indicates that the subject mammal does not exhibit the disorder or that treatment for the disorder is effective. In instances wherein the ratio of Stat4 to Stat6 in the sample is greater than or equal to that in the control sample, such a result indicates that the subject mammal exhibits or continues to exhibit the disorder. In instances wherein the ratio of Stat4 to Stat6 in the transplant sample is less than that in the sample, such a result indicates that the subject mammal does not exhibit the disorder or that treatment for the disorder is effective.

The methods for monitoring transplant acceptance or monitoring an autoimmune disorder can be performed with kits designed for carrying out such methods. As such, the present invention also relates to kits for monitoring transplant acceptance and autoimmune disorders.

In yet another aspect, the present invention relates to a method for identifying a compound to be tested for an ability to reduce immune rejection, said method comprises: (a) contacting an activated T cell sample with a test compound; (b) determining the amount of at least one of the following: (i) Stat1 mRNA or Stat1 protein, (ii) Stat2 mRNA or Stat2 protein, (iii) Stat3 mRNA or Stat3 protein, (iv) Stat4 mRNA or Stat4 protein, (v) Stat6 mRNA or Stat6 protein; (vi) SOCS1 mRNA or SOCS1 protein, or (vii) SOCS3 mRNA or SOCS3 protein, present in (a); and (c) comparing the amount(s) in (a) to that/those present in a corresponding control activated T cell sample that has not been contacted with the test compound, so that if the amount of (i), (ii), (iii), (iv), (vi), or (vii) is decreased, or the amount of (v) is increased, relative to the amount in the control sample, a compound to be tested for an ability to reduce immune rejection is identified. In alternate embodiments, such methods comprise determining the amount of at least two, at least three, at least four, at least five, at least six, or each of (i) to (vii) present in the activated T cell sample and comparing the amounts to those present in the control sample.

In certain embodiments, the amount of mRNA is determined, in other embodiments, the amount of protein is determined, while in still other embodiments, the amount of mRNA and protein is determined. With respect to Stat6, when the amount of Stat6 is being determined, it is preferable that the amount of Stat6 protein be determined. In any such embodiment wherein a Stat protein amount is determined, the amount determined

can be the total amount of the Stat protein present in a sample or, alternatively, can be the amount of phosphorylated Stat protein present in the sample.

In a preferred embodiment of a method for identifying a compound to be tested for an ability to reduce immune rejection, said method comprises: (a) contacting an activated T cell sample with a test compound; (b) determining the amount of Stat4 mRNA and Stat6 mRNA or Stat4 protein and Stat6 protein present in the sample; and (c) comparing the amounts in (b) to those present in a corresponding control activated T cell sample that has not been contacted with the test compound, so that if the amount of Stat4 is decreased or the amount of Stat6 is increased relative to the amount in the control sample, a compound to be tested for an ability to reduce immune rejection is identified.

In another preferred embodiment of a method for identifying a compound to be tested for an ability to reduce immune rejection, said method comprises: (a) contacting an activated T cell sample with a test compound; (b) determining the ratio of Stat4 mRNA to Stat6 mRNA or Stat4 protein to Stat6 protein present in the sample; and (c) comparing the ratio in (b) to that present in a corresponding control activated T cell sample that has not been contacted with the test compound, so that if the ratio in the sample is decreased relative to that in the control sample, a compound to be tested for an ability to reduce immune rejection is identified.

In another aspect, the present invention relates to a method for identifying a compound to be tested for an ability to reduce immune rejection, said method comprises: (a) contacting a resting T cell sample, a T cell activator and a test compound; (b) determining the amount of at least one of the following: (i) Stat1 mRNA or Stat1 protein, (ii) Stat2 mRNA or Stat2 protein, (iii) Stat3 mRNA or Stat3 protein, (iv) Stat4 mRNA or Stat4 protein, (v) Stat6 mRNA or Stat6 protein; (vi) SOCS1 mRNA or SOCS1 protein, or (vii) SOCS3 mRNA or SOCS3 protein, present in (a); and (c) comparing the amount(s) in (a) to that/those present in a corresponding resting T cell sample that has been contacted with the T cell activator, but has not been contacted with the test compound, so that if the amount of (i), (ii), (iii), (iv), (vi), or (vii) is decreased, or the amount of (v) is increased, relative to the amount in the control sample, a compound to be tested for an ability to reduce immune rejection is identified. In alternate embodiments, such methods comprise determining the amount of at least two, at least three, at least four, at least five, at least six, or each of (i) to (vii) present in the activated T cell sample and comparing the amounts to those present in the control sample.

In certain embodiments of such methods, the amount of mRNA is determined, in other embodiments, the amount of protein is determined, while in still other embodiments, the amount of mRNA and protein is determined. With respect to Stat6, when the amount of

Stat6 is being determined, it is preferable that the amount of Stat6 protein be determined. In any such embodiment wherein a Stat protein amount is determined, the amount determined can be the total amount of the Stat protein present in a sample or, alternatively, can be the amount of phosphorylated Stat protein present in the sample. Further, in certain

5 embodiments, the resting T cell is a primary T cell, and in other embodiments, the resting T cell is a T cell line.

 In a preferred embodiment of a method for identifying a compound to be tested for an ability to reduce immune rejection, said method comprises: (a) contacting a resting T cell sample, a T cell activator and a test compound; (b) determining the amount of Stat4
10 mRNA and Stat6 mRNA or Stat4 protein and Stat6 protein present in the sample; and (c) comparing the amounts in (b) to those present in a corresponding control resting T cell sample that has been contacted with the T cell activator, but has not been contacted with the test compound, so that if the amount of Stat4 is decreased or the amount of Stat6 is increased relative to the amount in the control sample, a compound to be tested for an
15 ability to reduce immune rejection is identified.

 In another preferred embodiment of a method for identifying a compound to be tested for an ability to reduce immune rejection, said method comprises: (a) contacting a resting T cell sample, a T cell activator and a test compound; (b) determining the ratio of Stat4 mRNA to Stat6 mRNA or Stat4 protein to Stat6 protein present in the sample; and (c)
20 comparing the ratio in (b) to that present in a corresponding control resting T cell sample that has been contacted with a T cell activator, but has not been contacted with the test compound, so that if the ratio in (a) is decreased relative to that in the control sample, a compound to be tested for an ability to reduce immune rejection is identified.

 In another aspect, the present invention relates to a method for identifying a
25 compound to be tested for an ability to reduce immune rejection, comprising: (a) contacting a T cell sample, a cytokine and a test compound, wherein the T cell sample is responsive to the cytokine; (b) determining the amount of at least one of the following: (i) Stat1 mRNA or Stat1 protein, (ii) Stat2 mRNA or Stat2 protein, (iii) Stat3 mRNA or Stat3 protein, (iv) Stat4 mRNA or Stat4 protein, (v) Stat6 mRNA or Stat6 protein; (vi) SOCS1
30 mRNA or SOCS1 protein, or (vii) SOCS3 mRNA or SOCS3 protein, present in (a); and (c) comparing the amount(s) in (a) to that/those present in a corresponding control T cell sample that has been contacted with the cytokine, but has not been contacted with the test compound, so that if the amount of (i), (ii), (iii), (iv), (vi), or (vii) is decreased, or the amount of (v) is increased, relative to the amount in the control sample, a compound to be
35 tested for an ability to reduce immune rejection is identified. In preferred embodiments, the cytokine is IL-2, IL-4, IL-12, or IL-13.

In certain embodiments of such methods, the amount of mRNA is determined, in other embodiments, the amount of protein is determined, while in still other embodiments, the amount of mRNA and protein is determined. With respect to Stat6, when the amount of Stat6 is being determined, it is preferable that the amount of Stat6 protein be determined. In
5 any such embodiment wherein a Stat protein amount is determined, the amount determined can be the total amount of the Stat protein present in a sample or, alternatively, can be the amount of phosphorylated Stat protein present in the sample.

In a preferred embodiment of such a method for identifying a compound to be tested for an ability to reduce immune rejection, said method comprises: (a) contacting a T cell
10 sample, a cytokine and a test compound, wherein the T cell sample is responsive to the cytokine; (b) determining the amount of Stat4 and Stat6 mRNA or Stat4 and Stat6 protein present in the sample; and (c) comparing the amounts in (a) to those present in a corresponding control T cell sample that has been contacted with the cytokine, but has not been contacted with the test compound, so that if the amount of Stat4 is decreased or the
15 amount of Stat6 is increased relative to the amounts in the control sample, a compound to be tested for an ability to reduce immune rejection is identified.

In another preferred embodiment of such a method for identifying a compound to be tested for an ability to reduce immune rejection, said method comprises: (a) contacting a T cell sample, a cytokine and a test compound, wherein the T cell sample is responsive to the
20 cytokine; (b) determining the ratio of Stat4 mRNA to Stat6 mRNA or Stat4 mRNA to Stat6 protein present in the sample; and (c) comparing the ratio to in (a) to that present in a corresponding control T cell sample that has been contacted with the cytokine, but has not been contacted with the test compound, so that if the ratio in the sample is decreased relative to that in the control sample, a compound to be tested for an ability to reduce
25 immune rejection is identified.

In yet another aspect, the present invention relates to methods for reducing immune rejection in a subject mammal, said methods comprising: administering to a subject mammal in need of such a reduction a concentration of a compound sufficient to decrease the level of Stat4 mRNA or protein in the subject relative to that observed in the subject in
30 the absence of the compound, wherein said compound does not induce platelet aggregation and does not affect NF- κ B activation in CD40L⁺ cells.

Alternatively, such methods for reducing immune rejection in a subject mammal can comprise: administering to a subject mammal in need of such a reduction a concentration of a compound sufficient to increase the level of Stat6 mRNA or protein in the subject
35 relative to that observed in the subject in the absence of the compound, wherein said

compound does not induce platelet aggregation and does not affect NF- κ B activation in CD40L⁺ cells.

Such methods for reducing immune rejection in a subject mammal can also comprise: administering to a subject mammal in need of such a reduction a concentration
5 of a compound sufficient to decrease the level of Stat4 mRNA or protein and maintain or increase the level of Stat6 mRNA or protein in the subject relative to that observed in the subject in the absence of the compound, wherein said compound does not induce platelet aggregation and does not affect NF- κ B activation in CD40L⁺ cells.

The methods of the present invention for reducing immune rejection can be utilized,
10 *e.g.*, for reducing immune rejection in a subject mammal that has undergone a transplant. For example, such methods can induce tolerance in a subject mammal that has undergone a transplant. The methods of the present invention for reducing immune rejection can also be utilized, *e.g.*, for reducing immune rejection in a subject mammal exhibiting an autoimmune disorder.

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3.1. DEFINITIONS

As used herein, the term “transplant” includes any cell, organ, organ system or tissue which can elicit an immune response in a recipient subject mammal. In general, therefore, a transplant includes an allograft or a xenograft cell, organ, organ system or tissue. An
20 allograft refers to a graft (cell, organ, organ system or tissue) obtained from a member of the same species as the recipient. A xenograft refers to a graft (cell, organ, organ system or tissue) obtained from a member of a different species as the recipient.

The term “immune rejection,” as used herein, is intended to refer to immune responses involved in transplant rejection, as well as to the concomitant physiological result
25 of such immune responses, such as for example, interstitial fibrosis, chronic graft arteriosclerosis, or vasculitis. The term “immune rejection,” as used herein, is also intended to refer to immune responses involved in autoimmune disorders, and the concomitant physiological result of such immune responses, including T cell-dependent infiltration and direct tissue injury; T cell-dependent recruitment and activation of
30 macrophages and other effector cells; and T cell-dependent B cell responses leading to autoantibody production.

The term “transplant rejection,” as used herein, refers to T cell-mediated rejection of transplant cells, organs, organ systems or tissue. In general, such transplant rejection generally includes accelerated, acute and chronic rejection. It is intended that the term, as
35 used herein, also refer to graft versus host disease, and the physiological results of such a disorder.

The term "reducing immune rejection," is meant to encompass prevention or inhibition of immune rejection, as well as delaying the onset or the progression of immune rejection. The term is also meant to encompass prolonging survival of a transplant in a subject mammal, or reversing failure of a transplant in a subject. Further, the term is meant to encompass ameliorating a symptom of an immune rejection, including, for example, ameliorating an immunological complication associated with immune rejection, such as for example, interstitial fibrosis, chronic graft atherosclerosis, or vasculitis. The term is also meant to encompass induction of tolerance in a subject mammal that has undergone a transplant.

The term "tolerance," as used herein, refers to a state wherein the immune system of a transplant recipient subject mammal is non-responsive to the transplant. This state is considered donor transplant-specific, and, as such, is distinguished from nonspecific immunosuppression. Operatively, the term as used herein, refers to permanent acceptance of a graft without ongoing immunosuppression, wherein, for example, challenge with a second graft of donor origin (especially when the second graft is of the same tissue as the first graft) should be accepted, and challenge with a third party graft should be rejected.

The term "autoimmune rejection," as used herein, refers to immune responses involved in autoimmune disorders, and the concomitant physiological result of such immune responses.

The term "activated T cell," as used herein, refers to a T cell that expresses antigens indicative of T-cell activation (that is, T cell activation markers). Examples of T cell activation markers include, but are not limited to, CD25, CD26, CD30, CD38, CD69, CD70, CD71, ICOS, OX-40 and 4-1BB. The expression of activation markers can be measured by techniques known to those of skill in the art, including, for example, western blot analysis, northern blot analysis, RT-PCR, immunofluorescence assays, and fluorescence activated cell sorter (FACS) analysis.

The term "resting T cell," as used herein, refers to a T cell that does not express T-cell activation markers. Resting T cells include, but are not limited to, T cells which are CD25⁻, CD69⁻, ICOS⁻, SLAMF⁻, and 4-1BB⁻. The expression of these markers can be measured by techniques known to those of skill in the art, including, for example, western blot analysis, northern blot analysis, RT-PCR, immunofluorescence assays, and fluorescence activated cell sorter (FACS) analysis.

The term "T cell activator," as used herein, refers to any compound or factor that is a T cell receptor stimulatory factor, that is, induces T cell receptor signalling. Preferably, the compound or factor also induces co-stimulatory pathways. Non-limiting examples of T cell activators include, but are not limited to, anti-CD3 antibodies (preferably monoclonal

antibodies) either alone or in conjunction with anti-CD28 antibodies (preferably monoclonal antibodies), or mitogens such as, for example, phorbol 12-myristate 13-acetate (PMA), phytohemagglutinin (PHA) or concanavalin-A (Con-A).

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4. BRIEF DESCRIPTION OF THE DRAWINGS

Figure 1. Human Stat1 nucleic acid and amino acid sequences (SEQ ID NOs:1, 2, respectively).

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Figure 2. Human Stat2 nucleic acid and amino acid sequences (SEQ ID NOs:3, 4, respectively).

Figure 3. Human Stat3 nucleic acid and amino acid sequences (SEQ ID NOs:5,6, respectively).

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Figure 4. Human Stat4 nucleic acid and amino acid sequences (SEQ ID NOs:7, 8, respectively).

Figure 5. Human Stat6 nucleic acid and amino acid sequences (SEQ ID NOs:9, 10, respectively).

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Figure 6. Human SOCS1 nucleic acid and amino acid sequences (SEQ ID NOs:11, 12, respectively).

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Figure 7. Human SOCS3 nucleic acid and amino acid sequences (SEQ ID NOs:13, 14, respectively).

Figure 8. Human Jak2 nucleic acid and amino acid sequences (SEQ ID NOs:15, 16 respectively).

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Figure 9. Human Tyk2 nucleic acid and amino acid sequences (SEQ ID NOs:17, 18 respectively).

Figure 10. Effects of immunosuppressants on CD154 mAb-induced cardiac allograft survival. Murine recipients were followed for up to 100 days post-transplant, and mean (\pm SD) cardiac allograft survival are shown (n = 6/group). Statistical analysis (Mann-

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Whitney U test) showed that CD154 mAb or combined CD154 and rapamycin (CD154RPM) induced highly significant prolongation of allograft survival ($p < 0.001$) compared to recipients treated with IgG, combined CD154 and cyclosporin A (CD154/CsA), or CD154 plus methylprednisolone (CD154/MP).

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Figure 11A-11B. Contrasting effects of immunosuppressive agents on activation-induced CD154 expression by CD4⁺ T cells, as determined by dual color flow cytometry after 7 hr of culture. Except for the control sample depicted in the uppermost section of Panel A, all culture plates were pre-coated with CD3 mAb.

10 Representative individual samples are shown in panel A, in which the histograms depict CD154 expression by gated CD4⁺ T cells. The percentage of CD154⁺ cells and the mean channel fluorescence of CD154 staining are indicated. Panel B shows the average activation-induced CD154 expression determined from 3 independent experiments. Percent CD154 expression (mean \pm SD) was calculated relative to the percentage of CD4⁺ CD154⁺ cells observed in the absence of immunosuppressive drugs. Cyclosporin A (CsA) and methylprednisolone (MP) significantly inhibited activation-induced CD154 expression ($p < 0.01$).

Figure 12A-12B. Critical role for NF- κ B in activation-induced CD154 expression by CD4⁺ T cells in vitro. The experimental design and data presentation are identical to that shown in Figure 10: splenic cells were derived from either NF κ B/p50 KO or control wild-type (WT) mice: Genetic deletion of NF κ B/p50 inhibited activation-induced CD154 expression ($p < 0.02$). Inhibition of NF- κ B p50 in WT mice by the proteasome antagonists lactacystin (LC) and MG-273 (MG) also significantly blocked activation-induced CD154 expression ($p < 0.04$).

Figure 13. Permanent cardiac allograft survival using CD154 mAb is NF- κ B-dependent. Mice were followed for up to 100 days post-transplant and mean (\pm SD) cardiac allograft survival are shown ($n = 6$ /group). Statistical analysis (Mann-Whitney U test) showed that use of NF- κ B/p50 KO mice as recipients, or administration of a lactacystin-derived (LC) proteasome inhibitor to wild-type mice, significantly impaired the efficacy of CD154 mAb therapy ($p < 0.001$).

Figure 14A. Kinetics of Stat RNA expression in heart allografts. Post-transplantation expression of Stats at day 1, day 2, day 3, day 5 were studied using RNA obtained from transplanted (Balb/c), native (B6/129), and control (B6/129) hearts of 8-10

weeks old female mice. Equal amounts of heart RNA (25 µg) were loaded onto each lane of three 1.2% agarose-formaldehyde gels. Hybridizations were done with probes specific to the transcriptional activation domains and 3'-untranslated regions of the Stats. The Stat probes described in the Materials and Methods Section of Section 8, below, were used for all the hybridizations. The locations of probes are shown underneath the cDNA line drawings next to the hybridization patterns. Murine GAPDH cDNA fragment was used as a control. The Stats and SOCS listed on the right of the GAPDH hybridization patterns indicate the probes used with the particular membranes. Designations: C, control heart (B6/129); N, native heart; T, transplanted heart. These data demonstrate that in the allografts Stats 1 and 2 increase progressively, peaking at day 5, whereas Stat3 rises to a plateau level by day 1. All 3 Stats increase within native control hearts by day 5.

Figure 14B. Kinetics of Stat RNA expression in heart allografts. The Stat levels were normalized against the GAPDH values. Normalized Stat levels are graphed as relative optical density (relative to other Stats). Closed, open, and gray bars indicate Stat RNA levels in transplant, native, and control hearts, respectively. Designations: D1, day 1; D2, day2; D3, day3; D5, day5; NAT, native; TRA, transplant; CONT, control.

Figure 15. Kinetics of SOCS/CIS RNA expression in heart allografts. Post-transplantation expression of SOCS/CIS RNA at day 1, day 2, day 3, and day 5 post-transplantation were studied by deprobing and rehybridizing the same membranes initially used for studying the Stat RNA expression studies described above in Figure 14A. Hybridizations were done with probes specific to the 3'-untranslated regions of the SOCS. The SOCS and CIS probes described in the Materials and Methods Section of Section 8, below, were used for all the hybridizations. The locations of probes are shown underneath the cDNA line drawings next to the hybridization patterns. Murine GAPDH cDNA fragment was used as a control. The SOCS/CIS list on the right of the GAPDH hybridization patterns indicate the probes used with the individual membranes. Designations: N, native hearts; T, transplanted hearts; C, control hearts (B6/129). These data demonstrate that SOCS1 is densely expressed in allografts at day 5, with only minor expression in native heart samples. SOCS3 is well-expressed from day 1 in allografts, with a further increase at day 5; no expression was seen in native hearts. SOCS5 is expressed predominantly as a 4.4 kb species, with a minor species at 3.8 kb, in control and native heart samples. However, in allografts, the 2 SOCS5 species are expressed in approximately equal amounts. By contrast, CIS expression remained unchanged in native hearts but decreased in allografts from day 1 onwards.

Figure 16. The similarity in the expression patterns of Stat4 and SOCS3 RNA in heart allografts. Stat4 and SOCS3 RNA expression in the days that follow the surgery show a high degree of similarity between the patterns of Stat4 and SOCS3 RNAs, albeit differences in their abundance (using Kodak Biomax MR film at -80°C , with intensifying screen, the exposure time for the Stat4 blot was approximately 4 days, and for SOCS3 only 15 hrs).

Figure 17. Stat protein levels following cardiac transplantation. Total proteins from the native and transplant hearts of untreated mice were extracted following transplantation, electrophoresed, transferred onto Immobilon-P membranes and Stat levels analyzed by Western blots as described in Materials and Methods section of Section 8, below. To identify Stats correctly, cell extracts recommended as positive controls by the antibody suppliers were used in the Western blots, together with Precision Protein standards. Designations: N, native hearts; T, transplant hearts; C, control hearts (Balb/c or B6/129), HSC 70, constitutive heat-shock protein. Molecular weights of the Stats are shown on the right. The list on the right of the HSC70 patterns indicates which Stat antibody was used with the individual membranes. These data demonstrate a general agreement with the mRNA data, and show that all of the Stats except Stat5A increase in allografts just prior to rejection (i.e. at day 5), with the greatest and most allograft-specific expression being found for Stat4.

Figure 18. Phosphorylated Stat levels following cardiac transplantation. Proteins were prepared and Stat levels analyzed by Western blotting as described in Materials and Methods section of Section 8, below. Designations: N, native hearts; T, transplant hearts; C, control hearts (Balb/c or B6/129), HSC 70, constitutive heat-shock protein. Molecular weights of the Stats are shown on the right. These data demonstrate an increase in pStat1 which parallels the rise in Stat1 levels.

Figure 19. Stat RNA expression in day 5 post-transplant heart allografts and the effect of anti-CD40L antibody MR-1 on Stat expression. Expression of Stat1, Stat2, Stat3, Stat4, Stat5A and Stat6 RNA was studied using RNA from transplanted (Balb/c) native (B6/129), and control (B6/129) hearts of 8-10 wks old female mice. Equal amounts of heart RNA (25 μg) were loaded onto each lane of four 1.2% agarose-formaldehyde gels, blotted and analyzed as explained in the Materials and Methods section presented in Section 8, below. The Stat probes described in the Materials and Methods section presented in

Section 8 were used for all the hybridizations. Murine GAPDH cDNA fragment was used as a control. The size of the Stat RNAs are shown on the right. The Stat and SOCS list on the right of the GAPDH hybridization patterns indicate the probes used with the particular membranes. Designations: C, control heart (B6/129); N, native heart; T, transplanted heart.

5 These data demonstrate that CD40L mAb suppresses expression of each of the Stat mRNA.

Figure 20. SOCS/CIS RNA expression in day 5 post-transplant heart allografts and the effect of anti-CD40L antibody MR-1 on SOCS/CIS expression. Expression of SOCS1, SOCS2, SOCS3, and CIS RNA was studied by deprobing and rehybridizing the same membranes initially used for studying the Stat RNA expression. Hybridizations were done with probes specific 3'-untranslated regions of the SOCS. The locations of probes are shown on the right. Designations: C, control heart (B6/129); N, native heart; T, transplanted heart. These data demonstrate that SOCS1 and SOCS3 levels increased during allograft rejection, but CD40L mAb prevented this induction. SOCS2 levels remained constant. CIS levels were decreased in transplants, regardless of CD40L mAb therapy.

Figure 21. The effect of treatment with the anti-CD40L monoclonal antibody MR-1 on the levels of Stats. Mice were given a single dose injection (DST plus IgG or DST plus MR-1) the day of the transplant surgery. Proteins were extracted from the native and transplanted hearts of both the IgG and MR-1 treated groups (2 mice/group) 5 days and 7 days following transplant surgery. Designations: C, control heart (B6/129); N, native heart; T, transplanted heart. Molecular weights of the Stats are shown on the right. These data demonstrate that CD40L mAb suppresses intragraft levels of each of the Stat proteins except that of Stat6, which at day 7 in this series was increased post-CD40L mAb therapy.

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Figure 22. Stat levels in isografts versus allografts following cardiac transplantation. Stat levels were studied in protein extracts from isografts (both donor and recipient mice were Balb/c) and allografts (donor mice were Balb/c and recipient mice were B6/129). Proteins were prepared and Stat, Bax, and HSC70 levels were analyzed by Western blotting as described in Materials and Methods section of Section 8, below. Designations: N, native hearts; T, transplant hearts; C, control hearts (Balb/c or B6/129), HSC 70, constitutive heat-shock protein. Molecular weights of the Stats are shown on the right. The list on the right of the HSC70 patterns indicates which Stat antibody was used with the individual membranes. These data demonstrate that isografts showed only low levels of Stats, whereas allografts had progressive increases in Stat proteins, peaking at day

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5. This pattern of induction was allo-specific and differed from that of an unrelated protein, Bax, which increased in both isografts and allografts.

5 5. DETAILED DESCRIPTION OF THE INVENTION

5.1. Methods and Compositions for Monitoring Transplant Acceptance and Autoimmune Disorders

As demonstrated below, immune rejection can be monitored by determining the amount of particular members of the Jak/Stat signal transduction pathway present within an affected tissue. Specifically, the results presented herein demonstrate that immune rejection can be monitored by determining the amount of Stat4 mRNA or protein, Stat6 mRNA or protein, SOCS1 mRNA or protein, or SOCS3 mRNA or protein present in an affected tissue. In particular, the results presented herein demonstrate that immune rejection can be monitored by determining the amount of Stat4 mRNA or protein, Stat6 mRNA or protein, SOCS1 mRNA or protein, or SOCS3 mRNA or protein, present in an affected tissue. The results presented herein also demonstrate that immune rejection can be monitored by determining the amount of Stat1 mRNA or protein, Stat2 mRNA or protein, or Stat3 mRNA or protein present, *e.g.*, present in an affected tissue.

The term "affected tissue," as used herein, refers to a transplant cell, tissue, organ, or organ system, or a cell, tissue, organ, or organ system. For example, such an affected tissue can include, but is not limited to, heart, liver, kidney, lung, bone marrow, skin, muscle, pancreatic islet, or intestine transplant cells, tissues, organs or organ system. The term "affected tissue," as used herein, also refers to a cell, tissue, organ or organ system that is, or is suspected of, being affected by an autoimmune disorder. For example, such an affected tissue can include, but is not limited to, a cell, tissue, organ, or organ system involved in systemic lupus erythematosus, glomerulonephritis, rheumatoid arthritis, Wegener's granulomatosis, chronic active hepatitis, or vasculitis.

Thus, aspects of the present invention relate to methods and compositions for monitoring such immune rejection. In particular, such methods and compositions can relate, for example, to methods for monitoring acceptance of a transplant in a subject mammal that has undergone a transplant, and can also relate, for example, to methods for monitoring an autoimmune disorder in a subject mammal being treated for or suspected of exhibiting an autoimmune disorder. Such methods and compositions are discussed in detail herein.

In one aspect, the invention relates to methods for monitoring acceptance of a transplant in a subject mammal that has undergone a transplant, wherein said method comprises: determining the amount of at least one of the following: (i) Stat4 mRNA or Stat4 protein, (ii) Stat6 mRNA or Stat6 protein, (iii) SOCS1 mRNA or SOCS1 protein, or
5 (iv) SOCS3 mRNA or SOCS3 protein, present in a transplant sample from the subject. In alternate embodiments, such methods comprise determining the amount of at least two, at least three, or each of (i) to (iv) present in the transplant sample. In certain embodiments, the amount of mRNA is determined, and can, for example, be determined via use of nucleic acid microarrays. In other embodiments, the amount of protein is determined, while in still
10 other embodiments, the amount of mRNA and protein is determined. With respect to Stat6, when the amount of Stat6 is being determined, it is preferable that the amount of Stat6 protein be determined. In any such embodiment wherein a Stat protein amount is determined, the amount determined can be the total amount of the Stat protein present in a sample or, alternatively, can be the amount of phosphorylated Stat protein present in the
15 sample.

In a preferred embodiment, a method for monitoring acceptance of a transplant in a subject mammal that has undergone a transplant comprises determining the amount of Stat4 and Stat6 mRNA or Stat4 and Stat6 protein present in a transplant sample from the subject. Such an embodiment can further comprise determining the ratio of Stat4 to Stat6 amounts.

20 Such methods can also further comprise comparing the amount or ratio determined to that present in a control sample, for example, a corresponding pre-transplant subject sample (*e.g.*, a sample from a corresponding pre-transplant cell, tissue, organ, or organ system) or a subject blood sample.

In instances wherein the amount of Stat4, SOCS1, or SOCS3 mRNA or protein in
25 the transplant sample is greater than, or the amount of Stat6 mRNA or protein in the transplant sample is less than, that of the control sample, such a result indicates that acceptance of the transplant has not been induced or is not being maintained. Likewise, in instances wherein the ratio of Stat4 to Stat6 in the transplant sample is greater than or equal to that in the control sample, such a result indicates that acceptance of the transplant has not
30 occurred, has not been induced or is not being maintained. Such results suggest a course of action that can include, for example, administration of a high dose of immunosuppressive drugs (*e.g.*, a high dose of corticosteroids, in, for example, the form of a single bolus intravenous injection) and /or administration of compounds to effectuate T cell depletion, including but not limited to administration of anti-CD3 antibodies.

35 In instances wherein the amount of Stat4, SOCS1, or SOCS3 mRNA or protein in the transplant sample is less than, or the amount of Stat6 mRNA or protein in the transplant

sample is equal to or greater than that of the control sample, such a result indicates that acceptance of the transplant has occurred, is being induced or is being maintained.

Likewise, in instances wherein the ratio of Stat4 to Stat6 in the transplant sample is less than that in the control sample, such a result indicates that acceptance of the transplant has occurred, has been induced or is being maintained.

The methods for monitoring acceptance of a transplant in a subject mammal that has undergone a transplant can further comprise assaying the transplant sample for evidence of lymphocyte infiltration or tissue damage (cell injury) using standard techniques. For example, histological techniques well known to those of skill in the art can be utilized to evaluate internationally recognized and used diagnostic criteria for the evaluation of graft rejection, which include features specific for each organ involved. For example, for evaluation of heart allograft transplants see, *e.g.*, Billingham, M.E., 1990, J. Heart Transplant. 9(3 Pt 2):272-6. For evaluation of renal allografts see, *e.g.*, Racusen et al., 1999, Kidney Int. 55(2):713-23. In one non-limiting embodiment, immunohistologic evaluation of transplant tissues (such as heart or kidney) can be performed via, *e.g.*, use of labeled antibody techniques to localize and quantitate gene expression. The evaluation of such criteria can, therefore, be enhanced by, for example, localization of Stat4, Stat6, SOCS1 and/or SOCS3 proteins, and/or detection of corresponding mRNAs via, *e.g.*, *in situ* hybridization.

Additionally, methods for monitoring acceptance of a transplant in a subject mammal that has undergone a transplant can comprise: determining the amount of at least one of the following: (i) Stat1 mRNA or Stat1 protein, (ii) Stat2 mRNA or Stat2 protein, or (iii) Stat3 mRNA or Stat3 protein, present in a cell sample from the subject. In alternate embodiments, such methods comprise determining the amount of at least two or each of (i) to (iii) present in the sample. In certain embodiments, the amount of mRNA is determined, and can, for example, be determined via use of nucleic acid microarrays. In other embodiments, the amount of protein is determined, while in still other embodiments, the amount of mRNA and protein is determined. In any such embodiment wherein a Stat protein amount is determined, the amount determined can be the total amount of the Stat protein present in a sample or, alternatively, can be the amount of phosphorylated Stat protein present in the sample.

In a particular embodiment of such Stat 1-, Stat 2-, and/or Stat 3-related methods, the cell sample is a transplant sample obtained within 2 to 3 days post-transplantation. In an alternative embodiment of such Stat 1-, Stat 2-, and/or Stat 3-related methods, the cell sample is a subject blood sample.

Such Stat 1-, Stat-2, and/or Stat 3-related methods can also further comprise comparing the amount determined to that present in a control sample, for example, a corresponding pre-transplant subject sample or, in the case of embodiments wherein the cell sample is a transplant sample obtained within 2-3 days post-transplantation, a subject blood
5 sample. In instances wherein the amount of Stat1, Stat2, or Stat3 mRNA or protein in the cell sample is greater than that of the control sample, such a result indicates that acceptance of the transplant has not occurred, has not been induced or is not being maintained. In instances wherein the amount of Stat1, Stat2, or Stat3 mRNA or protein in the transplant sample is less than that of the control sample, such a result indicates that acceptance of the
10 transplant has occurred, is being induced or is being maintained. Such results suggest a course of action that can include, for example, administration of a high dose of immunosuppressive drugs (*e.g.*, a high dose of corticosteroids, in, for example, the form of a single bolus intravenous injection) and /or administration of compounds to effectuate T cell depletion, including but not limited to administration of anti-CD3 antibodies.

15 Methods for monitoring acceptance of a transplant can be performed at any point post-transplantation. In a preferred embodiment, monitoring is performed daily during the first week post-transplant, followed by weekly monitoring until approximately one month post-transplant, followed by monthly monitoring until approximately one year post-transplant. It is understood, of course, that the frequency of monitoring can, at least in part,
20 depend upon the particular situation (*e.g.*, the nature of the graft, overall health of the recipient subject mammal, the particular immunotherapeutic, immunosuppressive, or immunomodulatory treatment being administered, etc.).

In another aspect, the invention relates to methods for monitoring an autoimmune disorder in a subject mammal, wherein said method comprises: determining the amount of
25 at least one of the following: (i) Stat4 mRNA or Stat4 protein, (ii) Stat6 mRNA or Stat6 protein, (iii) SOCS1 mRNA or SOCS1 protein, or (iv) SOCS3 mRNA or SOCS3 protein, present in a sample from a subject mammal being treated for or suspected of exhibiting the autoimmune disorder, wherein the sample is obtained from a tissue affected by the disorder. In alternate embodiments, such methods comprise determining the amount of at least two, at
30 least three, or each of (i) to (iv) present in the sample. In certain embodiments, the amount of mRNA is determined, in other embodiments, the amount of protein is determined, while in still other embodiments, the amount of mRNA and protein is determined.

In a preferred embodiment, a method for monitoring an autoimmune disorder in a subject mammal comprises determining the amount of Stat4 and Stat6 mRNA or Stat4 and
35 Stat6 protein present in a sample from the subject mammal being treated for or suspected of exhibiting the autoimmune disorder, wherein the sample is obtained from a tissue affected

by the disorder. Such an embodiment can further comprise determining the ratio of Stat4 to Stat6 amounts.

Such methods for monitoring an autoimmune disorder in a subject mammal can further comprise comparing the amount or ratio determined to that present in a control sample, for example, a corresponding tissue not affected by the disorder or a subject blood sample.

In instances wherein the amount of Stat4, SOCS1, or SOCS3 mRNA or protein in the sample is greater than, or the amount of Stat6 mRNA or protein in the sample is less than, that of the control sample, such a result indicates that the subject mammal exhibits or continues to exhibit the disorder. Likewise, in instances wherein the ratio of Stat4 to Stat6 in the sample is greater than or equal to that in the control sample, such a result indicates that the subject mammal exhibits or continues to exhibit the disorder. Such results suggest a course of action that can include, for example, the need to increase immunosuppression, as might be undertaken by bolus intravenous steroids (*e.g.*, methylprednisolone) or use of a CD3 mAb such as OKT3.

In instances wherein the amount of Stat4, SOCS1, or SOCS3 mRNA or protein in the sample is less than, or the amount of Stat6 mRNA or protein in the sample is equal to or greater than that of the control sample, such a result indicates that the subject mammal does not exhibit the disorder or that treatment for the disorder is effective. Likewise, in instances wherein the ratio of Stat4 to Stat6 in the transplant sample is less than that in the sample, such a result indicates that the subject mammal does not exhibit the disorder or that treatment for the disorder is effective.

The methods for monitoring an autoimmune disorder in a subject mammal can further comprise assaying the sample for evidence of leukocyte infiltration or tissue damage (cell injury) using standard techniques. For example, histological techniques well known to those of skill in the art can be utilized. Alternatively, standard techniques can be utilized to assay (*e.g.*, in serum) for the presence of autoimmune antibodies associated with the particular autoimmune disorder of interest. For example, there are standard immunohistology methods for detection of autoantibodies directed against a particular tissue (*e.g.*, anti-glomerular basement membrane, anti-parietal cell, anti-thyroid and anti-islet etc.), as well as assays for their detection in serum (*e.g.*, rheumatoid factor assay and anti-double-stranded DNA antibodies). See, *e.g.*, Manual of Clinical Laboratory Immunology (NR Rose, H Friedman, JL Fahey eds. 1986, Am Soc Microbiol, Washington, DC; Diagnostic Immunopathology (RB Colvin, et al., eds., 1995, Raven Press, New York).

The methods described herein can be performed using a sample from any subject mammal that has undergone a transplant or either exhibits or is suspected of exhibiting an

autoimmune disorder. Preferably, the mammal is a human, however, such subject mammals can also include, but are not limited to, pigs, dogs, cats, horses, cattle, sheep, mice, rats, and rabbits.

- It is noted that such methods for monitoring transplant acceptance and for
5 monitoring autoimmune disorders can be used to determine whether a subject can be effectively treated with a specific agent or class of agents intended to promote transplant acceptance or to treat the autoimmune disorder. Thus, in one embodiment, the present invention provides such methods for determining whether a subject can be effectively treated with an agent for an autoimmune disorder or for reducing immune rejection.
- 10 Monitoring the influence of agents (*e.g.*, drugs and compounds) on the expression or activity of Stat1, Stat2, Stat3, Stat4, Stat6, SOCS1 or SOCS3 mRNA or polypeptide can, therefore, be applied in basic drug screening, preclinical studies, clinical trials and during therapeutic treatment regimens designed to reduce immune rejection or to ameliorate a symptom of an autoimmune disorder.
- 15 The methods described herein comprise determining the amount of Stat1 mRNA or protein, Stat2 mRNA or protein, Stat3 mRNA or protein, Stat4 mRNA or protein, Stat6 mRNA or protein, SOCS1 mRNA or protein, and/or SOCS3 mRNA or protein present in a sample. Standard techniques, as described below, can routinely be utilized to determine these amounts. In general, such methods of the invention can routinely be performed using
20 standard techniques for detecting the presence or absence of a polypeptide or nucleic acid of the invention in a biological sample. This involves obtaining a biological sample from a test subject and contacting the biological sample with a compound or an agent capable of detecting the polypeptide or mRNA such that the presence of a polypeptide or nucleic acid of the invention is detected in the biological sample.
- 25 When comparing levels, such comparisons can be either quantitative or qualitative. Thus, in qualitative instances, for example, in instances wherein a control sample is determined to contain none of a given molecule (that is, Stat1, Stat2, Stat3, Stat4, Stat6, SOCS1, SOCS3 mRNA or protein) and the molecule is determined to be present in the test sample (that is, either a transplant or autoimmune sample), the amount of the molecule in
30 the test sample is greater than that present in the control sample. In quantitative instances wherein both the control and test samples are determined to contain a given molecule, using standard techniques, the amount in the test sample can routinely be determined to be greater than, equal to, or less than that of the control sample. Likewise, using standard techniques, the ratio of Stat4 to Stat6 mRNA or protein present in test and control samples can routinely
35 be determined. In general, the amount of a given molecule in test and control samples will

differ by at least 2-fold, and in certain instances, 2.5-fold, at least 3-fold, at least 4-fold, at least 5-fold, or at least 10-fold.

With respect to methods for monitoring acceptance of a transplant, such methods can be performed on any transplant from which a sample to be analyzed can be obtained.

5 Such transplants include, but are not limited to, heart, liver, kidney, lung, bone marrow, skin, muscle, pancreatic islet, or intestine transplants.

Likewise, with respect to methods for monitoring autoimmune disorders, such methods can be performed for any autoimmune disorder (or suspected autoimmune disorder) for which a sample of an affected tissue (or a tissue suspected of being affected) can be obtained. Such autoimmune disorders include, but are not limited to, systemic lupus erythematosus, glomerulonephritis, rheumatoid arthritis, Wegener's granulomatosis, chronic active hepatitis, and vasculitis

Methods for obtaining samples from a recipient transplant subject mammal or from a subject mammal exhibiting or suspected of exhibiting an autoimmune disorder are well known to those of skill in the art. Such methods can include biopsy methods, such as, for example, standard needle or punch biopsy methods. In certain embodiments, a particular subset of the sample can be isolated for the analysis. For example, a particular subset of a transplant or autoimmune disorder sample containing cell types of interest (*e.g.*, leukocyte cell types) can be isolated. Such isolation can be performed utilizing standard techniques such as, for example, laser microdissection (see, *e.g.*, Fend et al., 1999, *Am. J. Pathol.* 154(1):61-6; Schutze et al., 1998, *Nat. Biotechnol.* 16(8):737-42; and Simone et al., 1998, *Trends Genet.* 14(7):272-6).

Stat1, Stat2, Stat3, Stat4, Stat6, SOCS1 and SOCS3 nucleic acid and amino acid sequences are well known to those of skill in the art. For Stat1, see, for example, Schindler et al., 1992, *Proc. Natl. Acad. Sci. U.S.A.* 89:7836-7839 and Zhong et al., 1994, *Science* 264:95-98. For Stat2, see, for example, Yan et al., 1995, *Nucleic Acids Res.* 23(3):459-463, Bluyssen and Levy, 1997, *J. Biol. Chem.* 272(7):4600-4605, and Paulson et al., 1999, *J. Biol. Chem.* 274(36):25343-25349. For Stat3, see, for example, Ripberger, 1995, *J. Biol. Chem.* 270(50):29998-30006, Akira et al., 1994, *Cell* 77(1):63-71, Zhong et al., 1994, *Science* 264:95-98, Zhong et al., 1994, *Proc. Natl. Acad. Sci. U.S.A.* 91:4806-4810. For Stat4, see, for example, Quelle et al., 1995, *Mol. Cell. Biol.* 15:3336-3343; and Hou et al., 1994, *Science* 265:1701-1706. For Stat6, see, for example, Yamamoto et al., 1994, *Mol. Cell. Biol.* 17:4342-4349; Zhang et al., 1994, 91:4806-4810; and Yamamoto et al., 1997, *Cytogenet. Cell.* 77:207-210. For SOCS1 and 3 sequences, see, for example, Starr et al., 1997, *Nature*, 387:917-921; Minamoto et al., 1997, *Biochem., Biophys. Res. Commun.* 237:79-83; Masuhara et al., 1997, *Biochem., Biophys. Res. Commun.* 239:439-446; Naka et

al., 1997, Nature 387:924-929; and Endo et al., 1997, Nature 387:921-924. Representative examples of human Stat1, Stat2, Stat3, Stat4, Stat6, SOCS1 and SOCS3 sequences are shown in FIGS. 1-7 (SEQ ID NOs:1-14), respectively.

Further, additional forms, e.g., alleles or species homologs of such sequences can routinely be obtained and detected using the sequences described above in conjunction with standard cloning and hybridization techniques such as those find in Sambrook et al., eds., *Molecular Cloning: A Laboratory Manual*, 2nd. ed., Cold Spring Harbor Laboratory, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY, 1989.

The methods for monitoring transplant acceptance or monitoring an autoimmune disorder can be performed with kits designed for carrying out such methods. As such, the present invention also relates to kits for monitoring transplant acceptance and autoimmune disorders.

Such kits can be utilized for determining the amount of Stat1, Stat2, Stat3, Stat4, Stat6, SOCS1, and/or SOCS3 mRNA present within a sample (e.g., a transplant sample or a sample obtained from an autoimmune tissue or a tissue suspected of being effected by an autoimmune disorder). Alternatively, such kits can be utilized for determining the amount of Stat1, Stat2, Stat3, Stat4, Stat6, SOCS1, and/or SOCS3 polypeptide present within a sample (e.g., a transplant sample or a sample obtained from an autoimmune tissue or a tissue suspected of being effected by an autoimmune disorder). A kit can be capable of being used to determine the amount of any one, two, three, four, five, six, or seven of Stat1, Stat2, Stat3, Stat4, Stat6, SOCS1, and/or SOCS3 sequences (mRNA or polypeptide).

The kit, for example, can comprise a microarray for determining such amounts, wherein the microarray comprises one or more nucleic acid sequences immobilized onto a solid surface, said nucleic acid sequence or sequences exhibiting complementarity to at least one of Stat1, Stat2, Stat3, Stat4, Stat6, SOCS1, and/or SOCS3 mRNA. The kit can, in addition, comprise a labeled compound or agent capable of detecting the of Stat1, Stat2, Stat3, Stat4, Stat6, SOCS1, and/or SOCS3 polypeptide or mRNA encoding the polypeptide in a biological sample and means for determining the amount of the polypeptide or mRNA in the sample (e.g., an antibody which binds the polypeptide or an oligonucleotide probe which binds to DNA or mRNA encoding the polypeptide). Kits can also include instructions for use in determining the amount or amounts of mRNA or polypeptide, and can also include directions for monitoring and diagnosis.

For antibody-based kits, the kit can comprise, for example: (1) a first antibody (e.g., attached to a solid support) which binds to a of Stat1, Stat2, Stat3, Stat4, Stat6, SOCS1, and/or SOCS3 polypeptide; and, optionally, (2) a second, different antibody which binds to either the polypeptide or the first antibody and is conjugated to a detectable agent.

For oligonucleotide-based kits, the kit can comprise, for example: (1) an oligonucleotide, *e.g.*, a detectably labeled oligonucleotide, which hybridizes to a of Stat1, Stat2, Stat3, Stat4, Stat6, SOCS1, and/or SOCS3 nucleic acid sequence; or (2) a pair of primers useful for amplifying a of Stat1, Stat2, Stat3, Stat4, Stat6, SOCS1, and/or SOCS3 nucleic acid molecule. The kit can also comprise, *e.g.*, a buffering agent, a preservative, or a protein stabilizing agent. The kit can also comprise components necessary for detecting the detectable agent (*e.g.*, an enzyme or a substrate).

For microarray-based kits, such kits can comprise a nucleotide sequence, *e.g.*, an oligonucleotide sequence, immobilized onto the surface of a solid support (*e.g.*, a glass or porous solid support).

The kits can also contain a control sample or a series of control samples (postive control, negative control, or both) which can be assayed and compared to the test sample contained.

Each component of the kit is usually enclosed within an individual container and all of the various containers are within a single package along with instructions for observing and monitoring transplant acceptance or monitoring an autoimmune disorder.

NUCLEIC ACID DETECTION

Preferred agents for detecting an mRNA of interest (that is Stat1, Stat2, Stat3, Stat4, Stat6, SOCS1 or SOCS3 mRNA) are primers or labeled nucleic acid probes capable of hybridizing to the mRNA under stringent hybridization conditions. Nucleic acid probes can be, for example, full-length sequences, such as the nucleic acid sequences depicted in FIGS. 1-7 (SEQ ID NOs:1, 3, 5, 7, 9, 11 and 13), or complements thereof, or portions of such sequences (or complements thereof), such as oligonucleotides of at least about 12, 15, 25 30, 50, 100, 250 or 500 nucleotides in length and sufficient to specifically hybridize under stringent conditions to the mRNA.

In certain embodiments, determination of the amount of Stat1, Stat2, Stat3, Stat4, Stat6, SOCS1 and/or SOCS3 mRNA involves the use of a probe/primer in a polymerase chain reaction (PCR) (*see, e.g.*, U.S. Patent Nos. 4,683,195 and 4,683,202), such as, for example, anchor PCR, RACE PCR or RT-PCR. Such methods can include the steps of collecting a cell sample, isolating mRNA from the cells of the sample, reverse transcribing the mRNA, contacting the sample with one or more primers which specifically hybridize to the selected sequence under conditions such that hybridization and amplification of the sequence (if present) occurs, and determining the amount of product that is present.

Alternative amplification methods can also routinely be utilized. Such methods can include, for example, self sustained sequence replication (Guatelli et al., 1990, Proc. Natl.

Acad. Sci. USA 87:1874-1878), transcriptional amplification system (Kwoh et al., 1989, Proc. Natl. Acad. Sci. USA 86:1173-1177), Q-Beta Replicase (Lizardi et al., 1988, Bio/Technology 6:1197), or any other nucleic acid amplification method, followed by the detection/quantitation of the amplified molecules using techniques well known to those of skill in the art. These schemes are especially useful for the detection of nucleic acid molecules if such molecules are present in very low numbers.

As used herein, the term "hybridizes under stringent conditions" is intended to describe conditions for hybridization and washing under which nucleotide sequences at least 60% (preferably at least 75%, more preferably at least 85%, most preferably at least 95%) identical to each other typically remain hybridized to each other. Such stringent conditions are known to those skilled in the art and can be found in *Current Protocols in Molecular Biology*, John Wiley & Sons, N.Y. (1989), 6.3.1-6.3.6. A preferred, non-limiting example of stringent hybridization conditions are hybridization in 6X sodium chloride/sodium citrate (SSC) at about 45° C, followed by one or more washes in 0.2 X SSC, 0.1% SDS at 50-65° C (preferably 65° C).

Probes can comprise any readily detectable label moiety. For example, probes utilized herein comprise a radioisotope, a fluorescent compound, an enzyme, or an enzyme co-factor as a label moiety.

In alternate embodiments, the Stat1, Stat2, Stat3, Stat4, Stat6, SOCS1 and/or SOCS3 mRNA sequences can be detected "*in situ*" directly upon the sample, *e.g.*, the biopsy sample. Techniques for such procedures are well known to those of skill in the art. See, *e.g.*, Nuovo, G.J., 1992, "PCR *In Situ* Hybridization: Protocols and Applications," Raven Press, NY.

In other embodiments, the amount of Stat1, Stat2, Stat3, Stat4, Stat6, SOCS1 and/or SOCS3 mRNA can be determined by hybridizing nucleic acid arrays, *e.g.*, microarrays. In a specific embodiment of the invention, the expression of one or more of Stat1, Stat2, Stat3, Stat4, Stat6, SOCS1 and SOCS3 is measured or detected using a DNA microarray. A DNA microarray or chip is a microscopic array of DNA fragments or synthetic oligonucleotides, disposed in a defined pattern on a solid support, wherein they are amenable to analysis by standard hybridization methods (see, *e.g.*, Schena, 1996, *BioEssays* 18: 427).

Microarrays share certain preferred characteristics: The arrays are reproducible, allowing multiple copies of a given array to be produced and easily compared with each other. Preferably the microarrays are small, usually smaller than 5 cm², and they are made from materials that are stable under binding (*e.g.*, nucleic acid hybridization) conditions. Microarrays contain a surface to which sequences corresponding to gene products (*e.g.*, mRNA, cDNA, crRNA, or complements thereof), can be specifically hybridized or bound at

a known position. For practicing the methods of the present invention, the binding sites of the microarray are polynucleotides, preferably DNA polynucleotides, that specifically hybridize to at least a portion of Stat1, Stat2, Stat3, Stat4, Stat6, SOCS1, or SOCS3 mRNA or cDNA, or any combination of such mRNA or cDNA molecules, produced by a subject mammal. That is, a given binding site or unique set of binding sites in the microarray will specifically bind the product (*e.g.*, mRNA or cDNA) of a single gene, *e.g.*, Stat1, Stat2, Stat3, Stat4, Stat6, SOCS1 or SOCS3.

Preferably, the nucleotide sequence of each of the different polynucleotide bound to the surface is in the range of about 15 to about 100 nucleotides in length. Polynucleotides can be synthesized using conventional methods, such as phosphoramidite-based synthesis methods. Alternatively, the binding site polynucleotide sequences can be derived from cDNA or genomic clones.

DNA microarrays can be probed using mRNA, extracted and, optionally, reverse-transcribed and amplified from a sample (*e.g.*, a transplant, autoimmune or control sample). Nucleic acid hybridization and wash conditions are optimally chosen so that the probe "specifically binds" or "specifically hybridizes" to a specific array site, *i.e.*, the probe hybridizes, duplexes or binds to a sequence array site with a complementary nucleic acid sequence but does not hybridize to a site with a non-complementary nucleic acid sequence. As used herein, one polynucleotide sequence is considered complementary to another when, if the shorter of the polynucleotides is less than or equal to 25 bases, there are no mismatches using standard base-pairing rules or, if the shorter of the polynucleotides is longer than 25 bases, there is no more than a 5% mismatch. Preferably, the polynucleotides are perfectly complementary (no mismatches). It can easily be demonstrated that specific hybridization conditions result in specific hybridization by carrying out a hybridization assay including negative controls (*see, e.g.*, Shalon et al., 1996, *Genome Research* 6:639-645, and Chee et al., 1996, *Science* 274:610-614) or positive controls. Thus, in a preferred embodiment, a microarray of the invention further comprises a binding site designed to act as a negative control and/or a binding site designed to act as a positive control. For example, a positive control can relate to a constitutively expressed gene sequence, *e.g.*, a ubiquitin sequence, HSC70, or GADPH. A negative control can relate to a gene sequence not expressed in the test cell or tissue being assayed.

Exemplary, non-limiting examples of hybridization conditions that can be utilized with DNA microarrays are as follows: hybridization in 5 X SSC plus 0.2% SDS at 65° C for 4 hours followed by washes at 25° C in low stringency wash buffer (1 X SSC plus 0.2% SDS) followed by 10 minutes at 25° C in high stringency wash buffer (0.1 X SSC plus 0.2% SDS) (Shena et al., 1996, *Proc. Natl. Acad. Sci. USA*, 93:10614-19).

The use of a two-color fluorescence labeling and detection scheme to define alterations in gene expression has been described, *e.g.*, in Shena et al., 1995, Science 270:467-470. An advantage of using mRNA, cRNA, or cDNA labeled with two different fluorophores is that a direct and internally controlled comparison of the mRNA levels
5 corresponding to each arrayed gene in two cell states (*e.g.*, control and transplant) can be made, and variations due to minor differences in experimental conditions (*e.g.*, hybridization conditions) will not affect subsequent analyses. However, it will be recognized that it is also possible to use cDNA from a single cell, and compare, for example, the absolute amount of a particular mRNA in, *e.g.*, a transplant or autoimmune
10 sample cell.

To facilitate detection the mRNA or cDNA are typically labeled with fluorescent dyes that emit at different wavelengths. Examples of fluorescent dyes include, but are not limited to, rhodamine, fluorescein, isothiocyanate, rhodamine, phycoerythrin, phycocyanin, allophycocyanin, *o*-phthaldehyde and fluorescamine. The fluorescence emissions at each
15 site of a DNA array can be, preferably, detected by scanning confocal laser microscopy. In one embodiment, a separate scan, using the appropriate excitation line, is carried out for each of two fluorophores used. Alternatively, a laser can be used that allows simultaneous specimen illumination at wavelengths specific to the two fluorophores and emissions from the two fluorophores can be analyzed simultaneously (see, *e.g.*, Shalon et al., 1996, Genome
20 Research 6:639-645).

Signals are recorded and, in a preferred embodiment, analyzed by computer, *e.g.*, using a 12 bit analog to digital board. In one embodiment the scanned image is despeckled using a graphics program (*e.g.*, Hijaak Graphics Suite) and then analyzed using an image gridding program that creates a spreadsheet of the average hybridization at each wavelength
25 at each site.

It will be appreciated that when mRNA or cRNA is hybridized to a microarray under suitable hybridization conditions, the level of hybridization to the site in the array corresponding to any particular gene will reflect the prevalence in the cell of mRNA transcribed from that gene. For example, when detectably labeled (*e.g.*, with a fluorophore)
30 cRNA complementary to the total cellular mRNA is hybridized to a microarray, the site on the array corresponding to a gene (*i.e.*, capable of specifically binding the product of the gene) that is not transcribed in the cell will have little or no signal (*e.g.*, fluorescent signal), and a gene for which the encoded mRNA is prevalent will have a relatively strong signal.

Microarrays can be made in a number of ways well known to those of skill in the art.
35 With respect to the nucleic acids of the binding sites, the nucleic acid for the microarray can be generated by synthesis of synthetic polynucleotides or oligonucleotides, *e.g.*, using N-

phosphonate or phosphoramidite chemistries (*e.g.*, Froehler et al., 1986, *Nucleic Acid Res* 14:5399-5407). In some embodiments, synthetic nucleic acids include non-natural bases, *e.g.*, inosine. Additionally, it is possible to vary the charge on the phosphate backbone of the oligonucleotide, for example, by thiolation or methylation, or even to use a peptide
5 rather than a phosphate backbone. The making of such modifications is within the skill of one trained in the art. Further, nucleic acid analogues may be used as binding sites for hybridization. An example of a suitable nucleic acid analogue is peptide nucleic acid (*see, e.g.*, Egholm et al., 1993, *Nature* 365:566-568; *see also* U.S. Patent No. 5,539,083, Cook et al., entitled "Peptide nucleic acid combinatorial libraries and improved methods of
10 synthesis," issued July 23, 1996). In addition, binding (hybridization) sites can also be made from plasmid or phage clones of genes, cDNAs (*e.g.*, expressed sequence tags), or inserts therefrom (Nguyen et al., 1995, *Genomics* 29:207-209). In yet another embodiment, the polynucleotide of the binding sites is RNA.

The nucleic acid or analogue is attached to a solid support to produce the binding
15 site. Solid supports may be made from glass, silicon, plastic (*e.g.*, polypropylene, nylon, polyester), polyacrylamide, nitrocellulose, cellulose acetate or other materials. In general, non-porous supports, and glass in particular, are preferred. The solid support may also be treated in such a way as to enhance binding of oligonucleotides thereto, or to reduce non-specific binding of unwanted substances thereto. Preferably, the glass support is treated
20 with polylysine or silane to facilitate attachment of oligonucleotides to the slide.

Methods of immobilizing DNA on the solid support may include direct touch, micropipetting (Yershov et al., *Proc. Natl. Acad. Sci. USA*, 1996, 93:4913-4918), or the use of controlled electric fields to direct a given oligonucleotide to a specific spot in the array (U.S. Patent No. 5,605,662). In principal, any type of array, for example, dot blots on a
25 nylon hybridization membrane (*see* Sambrook et al., 1989, *Molecular Cloning - A Laboratory Manual (2nd Ed.)*, Vols. 1-3, Cold Spring Harbor Laboratory, Cold Spring Harbor, New York), can be used, although, as will be recognized by those of skill in the art, very small arrays are preferred because hybridization volumes will be smaller. DNA can typically be immobilized at a density of 50, 75, 100, up to 10,000 oligonucleotides per cm²
30 and preferably at a density of about 1000 oligonucleotides per cm².

In addition, nucleic acids can be attached to a surface by printing on glass plates (Schena et al., 1995, *Science* 270:467-470; DeRisi et al., 1996, *Nature Genetics* 14:457-460; Shalon et al., 1996, *Genome Res.* 6:639-645; and Schena et al., *Proc. Natl. Acad. Sci. USA*, 1996, 93(20):10614-19.) As an alternative to immobilizing pre-fabricated
35 oligonucleotides onto a solid support, it is possible to synthesize oligonucleotides directly on the support (Maskos et al., 1993, *Nucl. Acids Res.* 21: 2269-70; Fodor et al., 1991,

Science 251: 767-73 ; Lipshutz et al., 1999, Nat. Genet. 21(1 Suppl):20-4; McGall et al., Proc. Natl. Acad. Sci. USA 93: 13555-60, 1996). Other methods for making microarrays, *e.g.*, by masking (Maskos and Southern, 1992, Nuc. Acids Res. 20:1679-1684), may also be used.

5

PROTEIN DETECTION

Standard techniques can also be utilized for determining the amount of the protein or proteins of interest (that is, Stat1, Stat2, Stat3, Stat4, Stat6, SOCS1 and/or SOCS3 protein) present in a sample. It is to be understood, that such a determination of the amount of a
10 protein present includes determining the total amount of a protein present, and also includes, especially with respect to determining the amount of a Stat protein present, determining the amount of a phosphorylated form of the protein present.

For example, standard techniques can be employed using, *e.g.*, immunoassays such as, for example, Western blot, immunoprecipitation followed by sodium dodecyl sulfate
15 polyacrylamide gel electrophoresis (SDS-PAGE), immunocytochemistry, and the like to determine the amount of the protein or proteins of interest present in a sample. A preferred agent for detecting a protein of interest is an antibody capable of binding to a protein of interest, preferably an antibody with a detectable label.

With respect to determining the amount of a phosphorylated form of a protein of
20 interest that is present in a sample, such a determination can also be performed using standard techniques well known to those of skill in the art. For example, such a determination can include, first, immunoprecipitation with an antibody that is specific for a phosphorylated amino acid residue, *e.g.*, an anti-phosphotyrosine antibody, such that all exhibiting such a phosphorylated residue in a sample will be immunoprecipitated. Second,
25 the immunoprecipitated proteins can be contacted with a second antibody that is specific for the particular protein of interest, *e.g.*, Stat1, Stat2, Stat3, Stat4, or Stat6. Alternatively, a phosphorylated protein of interest can be identified and quantitated using an antibody specific for the phosphorylated form of the particular protein itself, *e.g.*, an antibody specific for phosphorylated Stat1 that does not recognize non-phosphorylated Stat1. Such
30 antibodies exist, and are well known to those of skill in the art.

For such detection methods, protein from the sample to be analyzed can easily be isolated using techniques which are well known to those of skill in the art. Protein isolation methods can, for example, be such as those described in Harlow and Lane (Harlow, E. and Lane, D., 1988, "Antibodies: A Laboratory Manual", Cold Spring Harbor Laboratory Press,
35 Cold Spring Harbor, New York).

Preferred methods for the detection of the protein or proteins of interest involve their detection via interaction with a protein-specific antibody. For example, antibodies directed a protein of interest can be utilized as described herein. Antibodies directed against Stat1, Stat2, Stat3, Stat4, Stat6, SOCS1 or SOCS3 protein are well known to those of skill in the art. For example, antibodies directed against Stat1, Stat2, Stat3, Stat4, Stat6, SOCS1 or SOCS3 can be obtained from such companies as Zymed Laboratories, Inc. (South San Francisco, CA), Santa Cruz Biotechnology, Inc. (Santa Cruz, CA), and Research Diagnostics, Inc., (Flanders, NJ). Alternatively, such antibodies can be generated utilizing standard techniques well known to those of skill in the art. See, *e.g.*, Section 5.3, below, for a more detailed discussion of such antibody generation techniques. Briefly, such antibodies can be polyclonal, or more preferably, monoclonal. An intact antibody, or a fragment thereof (*e.g.*, Fab or F(ab')₂) can, for example, be used.

For example, antibodies, or fragments of antibodies, specific for a protein of interest can be used to quantitatively or qualitatively detect the presence of the protein. This can be accomplished, for example, by immunofluorescence techniques. Antibodies (or fragments thereof) can, additionally, be employed histologically, as in immunofluorescence or immunoelectron microscopy, for *in situ* detection of a protein of interest. *In situ* detection can be accomplished by removing a histological specimen (*e.g.*, a biopsy specimen) from a patient, and applying thereto a labeled antibody thereto that is directed to a Stat1, Stat2, Stat3, Stat4, Stat6, SOCS1 or SOCS3 protein. The antibody (or fragment) is preferably applied by overlaying the labeled antibody (or fragment) onto a biological sample. Through the use of such a procedure, it is possible to determine not only the presence of the protein of interest, but also its distribution, its presence in lymphocytes within the sample. A wide variety of well-known histological methods (such as staining procedures) can be utilized in order to achieve such *in situ* detection.

Immunoassays for a protein of interest typically comprise incubating a biological sample, *e.g.*, a biopsy or subject blood sample, of a detectably labeled antibody capable of identifying a protein of interest, and detecting the bound antibody by any of a number of techniques well-known in the art. As discussed in more detail, below, the term "labeled" can refer to direct labeling of the antibody via, *e.g.*, coupling (*i.e.*, physically linking) a detectable substance to the antibody, and can also refer to indirect labeling of the antibody by reactivity with another reagent that is directly labeled. Examples of indirect labeling include detection of a primary antibody using a fluorescently labeled secondary antibody.

The biological sample can be brought in contact with and immobilized onto a solid phase support or carrier such as nitrocellulose, or other solid support which is capable of immobilizing cells, cell particles or soluble proteins. The support can then be washed with

suitable buffers followed by treatment with the detectably labeled fingerprint gene-specific antibody. The solid phase support can then be washed with the buffer a second time to remove unbound antibody. The amount of bound label on solid support can then be detected by conventional means.

- 5 By "solid phase support or carrier" is intended any support capable of binding an antigen or an antibody. Well-known supports or carriers include glass, polystyrene, polypropylene, polyethylene, dextran, nylon, amylases, natural and modified celluloses, polyacrylamides, gabbros, and magnetite. The nature of the carrier can be either soluble to some extent or insoluble for the purposes of the present invention. The support material can
10 have virtually any possible structural configuration so long as the coupled molecule is capable of binding to an antigen or antibody. Thus, the support configuration can be spherical, as in a bead, or cylindrical, as in the inside surface of a test tube, or the external surface of a rod. Alternatively, the surface can be flat such as a sheet, test strip, etc. Preferred supports include polystyrene beads. Those skilled in the art will know many other
15 suitable carriers for binding antibody or antigen, or will be able to ascertain the same by use of routine experimentation.

- One of the ways in which a Stat1-, Stat2-, Stat3-, Stat4-, Stat6-, SOCS1- or SOCS3-specific antibody can be detectably labeled is by linking the same to an enzyme and use in an enzyme immunoassay (EIA) (Voller, A., "The Enzyme Linked Immunosorbent Assay
20 (ELISA)", 1978, Diagnostic Horizons 2:1-7, Microbiological Associates Quarterly Publication, Walkersville, MD); Voller, A. et al., 1978, J. Clin. Pathol. 31:507-520; Butler, J.E., 1981, Meth. Enzymol. 73:482-523; Maggio, E. (ed.), 1980, ENZYME IMMUNOASSAY, CRC Press, Boca Raton, FL; Ishikawa, E. et al., (eds.), 1981, ENZYME IMMUNOASSAY, Kigaku Shoin, Tokyo). The enzyme which is bound to the antibody will
25 react with an appropriate substrate, preferably a chromogenic substrate, in such a manner as to produce a chemical moiety which can be detected, for example, by spectrophotometric, fluorimetric or by visual means. Enzymes which can be used to detectably label the antibody include, but are not limited to, malate dehydrogenase, staphylococcal nuclease, delta-5-steroid isomerase, yeast alcohol dehydrogenase, alpha-glycerophosphate,
30 dehydrogenase, triose phosphate isomerase, horseradish peroxidase, alkaline phosphatase, asparaginase, glucose oxidase, beta-galactosidase, ribonuclease, urease, catalase, glucose-6-phosphate dehydrogenase, glucoamylase and acetylcholinesterase. The detection can be accomplished by colorimetric methods which employ a chromogenic substrate for the enzyme. Detection can also be accomplished by visual comparison of the extent of
35 enzymatic reaction of a substrate in comparison with similarly prepared standards.

Detection can also be accomplished using any of a variety of other immunoassays. For example, by radioactively labeling the antibodies or antibody fragments, it is possible to detect a protein of interest through the use of a radioimmunoassay (RIA) (see, for example, Weintraub, B., Principles of Radioimmunoassays, Seventh Training Course on Radioligand
5 Assay Techniques, The Endocrine Society, March, 1986, which is incorporated by reference herein). The radioactive isotope (*e.g.*, ^{125}I , ^{131}I , ^{35}S or ^3H) can be detected by such means as the use of a gamma counter or a scintillation counter or by autoradiography.

It is also possible to label the antibody with a fluorescent compound. When the fluorescently labeled antibody is exposed to light of the proper wavelength, its presence can
10 then be detected due to fluorescence. Among the most commonly used fluorescent labeling compounds are fluorescein isothiocyanate, rhodamine, phycoerythrin, phycocyanin, allophycocyanin, α -phthaldehyde and fluorescamine.

The antibody can also be detectably labeled using fluorescence emitting metals such as ^{152}Eu , or others of the lanthanide series. These metals can be attached to the antibody
15 using such metal chelating groups as diethylenetriaminepentacetic acid (DTPA) or ethylenediaminetetraacetic acid (EDTA).

The antibody also can be detectably labeled by coupling it to a chemiluminescent compound. The presence of the chemiluminescent-tagged antibody is then determined by detecting the presence of luminescence that arises during the course of a chemical reaction.
20 Examples of particularly useful chemiluminescent labeling compounds are luminol, isoluminol, therrromatic acridinium ester, imidazole, acridinium salt and oxalate ester.

Likewise, a bioluminescent compound can be used to label the antibody of the present invention. Bioluminescence is a type of chemiluminescence found in biological systems in, which a catalytic protein increases the efficiency of the chemiluminescent
25 reaction. The presence of a bioluminescent protein is determined by detecting the presence of luminescence. Important bioluminescent compounds for purposes of labeling are luciferin, luciferase and aequorin.

30 5.2 . Methods and Compositions for Identifying Compounds That Reduce Immune Rejection

As demonstrated below, immune rejection can be reduced and tolerance can be induced by modulating the amount of particular members of the Jak/Stat pathway present, expressed or active within an affected tissue. Specifically, the results presented herein demonstrate that modulation of Stat1, Stat2, Stat3, Stat4, Stat6, SOCS1 and/or SOCS3
35 levels can reduce immune rejection.

The methods described herein identify compounds that modulate the expression and/or activity of Stat1, Stat2, Stat3, Stat4, Stat6, SOCS1 or SOCS3 sequences in a manner that can reduce immune rejection (either, for example, in a transplant situation or in an autoimmune situation). The compounds identified via such methods are, therefore, useful
5 as lead compounds in the development of therapeutic compositions for the reduction of immune rejection. Such methods are particularly useful in that the effort and great expense involved in testing potential therapeutics *in vivo* is efficiently focused on those compounds identified via the *in vitro* and *ex vivo* methods described herein.

Thus, the present invention relates to a method for identifying a compound to be
10 tested for an ability to reduce immune rejection, said method comprises: (a) contacting an activated T cell sample with a test compound; (b) determining the amount of at least one of the following: (i) Stat1 mRNA or Stat1 protein, (ii) Stat2 mRNA or Stat2 protein, (iii) Stat3 mRNA or Stat3 protein, (iv) Stat4 mRNA or Stat4 protein, (v) Stat6 mRNA or Stat6 protein; (vi) SOCS1 mRNA or SOCS1 protein, or (vii) SOCS3 mRNA or SOCS3 protein,
15 present in (a); and (c) comparing the amount(s) in (a) to that/those present in a corresponding control activated T cell sample that has not been contacted with the test compound, so that if the amount of (i), (ii), (iii), (iv), (vi), or (vii) is decreased, or the amount of (v) is increased, relative to the amount in the control sample, a compound to be tested for an ability to reduce immune rejection is identified. In alternate embodiments,
20 such methods comprise determining the amount of at least two, at least three, at least four, at least five, at least six, or each of (i) to (vii) present in the activated T cell sample and comparing the amounts to those present in the control sample.

In certain embodiments, the amount of mRNA is determined, in other embodiments, the amount of protein is determined, while in still other embodiments, the
25 amount of mRNA and protein is determined. With respect to Stat6, when the amount of Stat6 is being determined, it is preferable that the amount of Stat6 protein be determined. In any such embodiment wherein a Stat protein amount is determined, the amount determined can be the total amount of the Stat protein present in a sample or, alternatively, can be the amount of phosphorylated Stat protein present in the sample.

30 In a preferred embodiment of a method for identifying a compound to be tested for an ability to reduce immune rejection, said method comprises: (a) contacting an activated T cell sample with a test compound; (b) determining the amount of Stat4 mRNA and Stat6 mRNA or Stat4 protein and Stat6 protein present in the sample; and (c) comparing the amounts in (a) to those present in a corresponding control activated T cell sample that has
35 not been contacted with the test compound, so that if the amount of Stat4 is decreased or the

amount of Stat6 is increased relative to the amount in the control sample, a compound to be tested for an ability to reduce immune rejection is identified.

In another preferred embodiment of a method for identifying a compound to be tested for an ability to reduce immune rejection, said method comprises: (a) contacting an activated T cell sample with a test compound; (b) determining the ratio of Stat4 mRNA to Stat6 mRNA or Stat4 protein to Stat6 protein present in the sample; and (c) comparing the ratio in (a) to that present in a corresponding control activated T cell sample that has not been contacted with the test compound, so that if the ratio in (a) is decreased relative to that in the control sample, a compound to be tested for an ability to reduce immune rejection is identified.

In another aspect, the present invention relates to a method for identifying a compound to be tested for an ability to reduce immune rejection, said method comprises: (a) contacting a resting T cell sample, a T cell activator and a test compound; (b) determining the amount of at least one of the following: (i) Stat1 mRNA or Stat1 protein, (ii) Stat2 mRNA or Stat2 protein, (iii) Stat3 mRNA or Stat3 protein, (iv) Stat4 mRNA or Stat4 protein, (v) Stat6 mRNA or Stat6 protein; (vi) SOCS1 mRNA or SOCS1 protein, or (vii) SOCS3 mRNA or SOCS3 protein, present in (a); and (c) comparing the amount(s) in (a) to that/those present in a corresponding resting T cell sample that has been contacted with the T cell activator, but has not been contacted with the test compound, so that if the amount of (i), (ii), (iii), (iv), (vi), or (vii) is decreased, or the amount of (v) is increased, relative to the amount in the control sample, a compound to be tested for an ability to reduce immune rejection is identified. In alternate embodiments, such methods comprise determining the amount of at least two, at least three, at least four, at least five, at least six, or each of (i) to (vii) present in the activated T cell sample and comparing the amounts to those present in the control sample.

Optionally, such methods can further include comparing the amount or amounts in (a) to a control resting T cell sample that has not been contacted with the T cell activator or with the test compound and/or with a control resting T cell sample that has been contacted with the test compound, but has not been contacted with a T cell activator (and, therefore, remains in the resting state). Such controls provide evidence regarding the specificity and toxicity of the test compound.

In certain embodiments of such methods, the amount of mRNA is determined, in other embodiments, the amount of protein is determined, while in still other embodiments, the amount of mRNA and protein is determined. With respect to Stat6, when the amount of Stat6 is being determined, it is preferable that the amount of Stat6 protein be determined. In any such embodiment wherein a Stat protein amount is determined, the amount determined

can be the total amount of the Stat protein present in a sample or, alternatively, can be the amount of phosphorylated Stat protein present in the sample. Further, in certain embodiments, the resting T cell is a primary T cell, and in other embodiments, the resting T cell is a T cell line.

- 5 In a preferred embodiment of a method for identifying a compound to be tested for an ability to reduce immune rejection, said method comprises: (a) contacting a resting T cell sample, a T cell activator and a test compound; (b) determining the amount of Stat4 mRNA and Stat6 mRNA or Stat4 protein and Stat6 protein present in the sample; and (c) comparing the amounts in (a) to those present in a corresponding control resting T cell
- 10 sample that has been contacted with the T cell activator, but has not been contacted with the test compound, so that if the amount of Stat4 is decreased or the amount of Stat6 is increased relative to the amount in the control sample, a compound to be tested for an ability to reduce immune rejection is identified.

- In another preferred embodiment of a method for identifying a compound to be
- 15 tested for an ability to reduce immune rejection, said method comprises: (a) contacting a resting T cell sample, a T cell activator and a test compound; (b) determining the ratio of Stat4 mRNA to Stat6 mRNA or Stat4 protein to Stat6 protein present in the sample; and (c) comparing the ratio in (a) to that present in a corresponding control resting T cell sample that has been contacted with a T cell activator, but has not been contacted with the test
- 20 compound, so that if the ratio in the sample is decreased relative to that in the control samples, a compound to be tested for an ability to reduce immune rejection is identified.

- In another aspect, the present invention relates to a method for identifying a compound to be tested for an ability to reduce immune rejection, comprising: (a) contacting a T cell sample, a cytokine and a test compound, wherein the T cell sample is
- 25 responsive to the cytokine; (b) determining the amount of at least one of the following: (i) Stat1 mRNA or Stat1 protein, (ii) Stat2 mRNA or Stat2 protein, (iii) Stat3 mRNA or Stat3 protein, (iv) Stat4 mRNA or Stat4 protein, (v) Stat6 mRNA or Stat6 protein; (vi) SOCS1 mRNA or SOCS1 protein, or (vii) SOCS3 mRNA or SOCS3 protein, present in (a); and (c) comparing the amount(s) in (a) to that/those present in a corresponding control T cell
- 30 sample that has been contacted with the cytokine, but has not been contacted with the test compound, so that if the amount of (i), (ii), (iii), (iv), (vi), or (vii) is decreased, or the amount of (v) is increased, relative to the amount in the control sample, a compound to be tested for an ability to reduce immune rejection is identified.

- Cytokines that can be used as part of these methods include, but are not limited to,
- 35 IL-2, IL-4, IL-12, or IL-13.

In certain embodiments of such methods, the amount of mRNA is determined, in other embodiments, the amount of protein is determined, while in still other embodiments, the amount of mRNA and protein is determined. With respect to Stat6, when the amount of Stat6 is being determined, it is preferable that the amount of Stat6 protein be determined. In
5 any such embodiment wherein a Stat protein amount is determined, the amount determined can be the total amount of the Stat protein present in a sample or, alternatively, can be the amount of phosphorylated Stat protein present in the sample.

In a preferred embodiment of such a method for identifying a compound to be tested for an ability to reduce immune rejection, said method comprises: (a) contacting a T cell
10 sample, a cytokine and a test compound, wherein the T cell sample is responsive to the cytokine; (b) determining the amount of Stat4 and Stat6 mRNA or Stat4 and Stat6 protein present in the sample; and (c) comparing the amounts in (a) to those present in a corresponding control T cell sample that has been contacted with the cytokine, but has not been contacted with the test compound, so that if the amount of Stat4 is decreased or the
15 amount of Stat6 is increased relative to the amounts in the control sample, a compound to be tested for an ability to reduce immune rejection is identified.

In another preferred embodiment of such a method for identifying a compound to be tested for an ability to reduce immune rejection, said method comprises: (a) contacting a T cell sample, a cytokine and a test compound, wherein the T cell sample is responsive to the
20 cytokine; (b) determining the ratio of Stat4 mRNA to Stat6 mRNA or Stat4 mRNA to Stat6 protein present in the sample; and (c) comparing the ratio to in (a) to that present in a corresponding control T cell sample that has been contacted with the cytokine, but has not been contacted with the test compound, so that if the ratio in the sample is decreased relative to that in the control sample, a compound to be tested for an ability to reduce
25 immune rejection is identified.

Standard methods and compositions for determining the amount of Stat1 mRNA or protein, Stat2 mRNA or protein, Stat3 mRNA or protein, Stat4 mRNA or protein, Stat6 mRNA or protein, SOCS1 mRNA or protein, and SOCS3 mRNA or protein can be utilized. Such methods and compositions are described in detail, above, in Section 5.1.

30 In addition to the ability to modulate Stat and/or SOCS levels as described herein, it may be desirable, at least in certain instances, that compounds that reduce immune rejection also modulate the expression or activity of such molecules as IL-4, interferon- γ (IFN- γ), IL-12, or IL-13. Thus, the methods described herein for identifying compounds to be tested for an ability to reduce immune rejection can further comprise determining the level of IL-
35 4, IFN- γ or IL-13 in the T cell sample that has been contacted with the test compound, and comparing this level with that of the control T cell sample that has not been contacted with

the test compound. Preferred compounds are ones wherein: the level of IL-12 or IFN- γ in the test sample is decreased relative to the corresponding level in the control sample, or wherein the level of IL-4 or IL-13 in the test sample is equal to or greater than the corresponding level in the control sample.

5 The present methods of identifying compounds that to be tested for an ability to reduce immune rejection, can comprise methods for identifying compounds that modulate the activity of Stat1, Stat2, Stat3, Stat4, Stat6, SOCS1, or SOCS3. Thus, such methods can comprise: (a) contacting a T cell sample with a test compound; (b) determining the activity of at least one of Stat1, Stat2, Stat3, Stat4, Stat6, SOCS1, or SOCS3; and (c) comparing the
10 activity level or levels to that/those in a corresponding control T cell sample that has not been contacted with the test compound, so that if the level of Stat1, Stat2, Stat3, Stat4, SOCS1 or SOCS3 activity in (a) is decreased, or the level of Stat6 activity in (a) is increased, relative to the level of activity in the control sample, a compound to be tested for an ability to reduce immune rejection is identified.

15 In one preferred embodiment, the activity of Stat4 and Stat6 is determined. Such a preferred embodiment can further include determining the ratio of Stat4 activity to Stat6 activity so that if the ratio in the test sample is decreased relative to that in the control sample, a compound to be tested for an ability to reduce immune rejection is identified.

 The T cell sample utilized herein can be an activated T cell sample, a resting T cell
20 sample, or a cytokine-responsive T cell sample, as discussed above. In instances wherein the T cell sample is a resting T cell sample, the T cell sample is contacted with a T cell activator and the test compound. In instances wherein the T cell sample is a cytokine-responsive T cell sample, the T cell sample is contacted with the cytokine and the test compound.

25 Standard techniques can be utilized to determine the level of Stat1, Stat2, Stat3, Stat4, Stat6, SOCS1 or SOCS3 activity. For example, the activity of Stat1, Stat2, Stat3, Stat4, Stat6, SOCS1 or SOCS3 can be determined by detecting the binding of Stat1, Stat2, Stat3, Stat4, Stat6, SOCS1 or SOCS3 to its cognate DNA binding element, via, for example, an electromobility shift assay ("EMSA"), detecting the expression of a gene
30 whose expression is controlled by a promoter that is responsive to Stat1, Stat2, Stat3, Stat4, Stat6, SOCS1 or SOCS3, detecting the induction of a reporter gene that comprises a regulatory element that is responsive to Stat1, Stat2, Stat3, Stat4, Stat6, SOCS1 or SOCS3, wherein the element is operably linked to a nucleic acid encoding a detectable marker, *e.g.*, luciferase.

35 Genes whose expression is controlled by a Stat 1-responsive promoter are well known, and include, for example GBP-1, inducible NO synthase (iNOS), ICAM, IRF-1,

major histocompatibility complex (MHC) class II transactivator (CIITA). See, *e.g.*, Lew et al., 1991, Mol. Cell. Biol. 11:182-191, Gao et al., 1997, J. Biol. Chem. 272:1226-1230, Caldenhoven et al., 1994, J. Biol. Chem. 269:21146-21154, Sims et al., 1993, Mol. Cell. Biol. 13:690-702, Pine et al., 1994, EMBO J. 13:158-167, Harada et al., 1994, Mol. Cell. Biol. 14:1500-1509, and Piskurich et al., 1999, Mol. Cell. Biol. 19(1):431-40. Thus, expression of such genes in the T cell sample in the presence and absence of a test compound can routinely be determined using standard techniques. Likewise, the structure of Stat 1-responsive promoters are well known (see, *e.g.*, Sims et al., 1993, Mol. Cell. Biol. 13:690-702, Pine et al., 1994, EMBO J. 13:158-167, and Piskurich et al., 1999, Mol. Cell. Biol. 19(1):431-40), making the construction and assay of Stat 1-reporter genes routine.

Genes whose expression is controlled by a Stat 2-responsive promoter are well known, and include, for example IRF-1. See, *e.g.*, Li et al., 1996, J. Biol. Chem. 271(10):5790-5794. Thus, expression of such genes in the T cell sample in the presence and absence of a test compound can routinely be determined using standard techniques. Likewise, the structure of Stat 2-responsive promoters are well known (see, *e.g.*, Ghislain et al., 1996, J. Biol. Chem. 271(21):12408-12413 and Li et al., 1996, J. Biol. Chem. 271(10):5790-5794), making the construction and assay of Stat 2-reporter genes routine.

Genes whose expression is controlled by a Stat 3-responsive promoter are well known, and include, for example alpha-2-macroglobulin, fibrinogen, junB, haptoglobin, matrix metalloproteinase (MMP-1), TIMP-1, and p21^{WAF/CIP1}. See, *e.g.*, Wegenka et al., 1993, Mol. Cell. Biol. 13:276-288, Fujitani et al., 1994, Biochem. Bioph. Res. Co. 202:1181-1187, Coffey et al., 1995, Oncogene 10:985-994, Akira et al., 1994, Cell 77:63-71, and Chin et al., 1996, Science 272:719-722. Thus, expression of such genes in the T cell sample in the presence and absence of a test compound can routinely be determined using standard techniques. Likewise, the structure of Stat 3-responsive promoters are well known (see, *e.g.*, Wegenka et al., 1993, Mol. Cell. Biol. 13:276-288 and Chin et al., 1996, Science 272:719-722), making the construction and assay of Stat 3-reporter genes routine.

Genes whose expression is controlled by a Stat 4-responsive promoter are well known, and include, for example interferon- γ and IL-12. See, *e.g.*, Grigorieva et al., 2000, J Biol. Chem. 275(10):7343-7350 and Naeger, L.K. et al., 1999, J. Biol. Chem. 274:1875-1878). Thus, expression of such genes in the T cell sample in the presence and absence of a test compound can routinely be determined using standard techniques. Likewise, the structure of Stat 4-responsive promoters are well known (see, *e.g.*, Grigorieva et al., 2000, J Biol. Chem. 275(10):7343-7350 and Naeger, L.K. et al., 1999, J. Biol. Chem. 274:1875-1878), making the construction and assay of Stat 4-reporter genes routine.

Genes whose expression is controlled by a Stat 6-responsive promoter are well known, and include, for example IL-4, CD23, IL-4 receptor, MHC class II. See, *e.g.*, Tinnell et al., 1998, *Int. Immunol.* 10(10):1529-38, Linehan et al., 1998, *J. Immunol.* 161(1):302-10, and Kotanides et al., 1996, *J. Biol. Chem.* 271(41):25555-25561. Thus, expression of such genes in the T cell sample in the presence and absence of a test compound can routinely be determined using standard techniques. Likewise, the structure of Stat 6-responsive promoters are well known (see, *e.g.*, Curiel, R.E. et al., 1997, *Eur. J. Imm.* 27:1982-1987, Linehan et al., 1998, *J. Immunol.* 161(1):302-10, and Kotanides et al., 1996, *J. Biol. Chem.* 271(41):25555-25561), making the construction and assay of Stat 6-reporter genes routine.

EMSAs can also routinely be utilized to assess Stat1, Stat2, Stat3, Stat4 or Stat6 activity. Such techniques are well known to those of skill in the art. See, *e.g.*, Amici et al., 1995, *Cancer Research* 55: 14452-4457. Briefly, in a representative, non-limiting example, extracts of cells treated with a test compound are mixed with ³²P-Stat 4 (or Stat1, 2, 3, or 6) element or a control oligonucleotide and poly(dI-dC) (Pharmacia Biotech Inc.) in binding buffer (*e.g.*, Tris-Cl, pH 7.8, 50 mM NaCl, 1 mM EDTA, 0.5 mM dithiothreitol, 5% glycerol). After an approximately 20 minute incubation at room temperature, Stat4 (or Stat1, 2, 3, or 6)-DNA-complexes or control oligonucleotide complexes are analyzed by nondenaturing 4% polyacrylamide gel electrophoresis and autoradiography. The amount of shifted Stat4 (or Stat1, 2, 3, or 6) probe, an indicator Stat4 (or Stat1, 2, 3, or 6) activity, respectively, can be quantitated by Molecular Dynamics PhosphoImager (MDP) analysis.

The activity of SOCS1 or SOCS3 activity can be determined by, *e.g.*, detecting the expression of a gene whose expression is controlled by SOCS1 or SOCS3. For example, SOCS1 expression inhibits IL-6, LIF, oncostatin M, IFN- γ , IFN- β , IFN- α , thrombopoietin, and growth hormone (GH) induced Jak/Stat signaling. SOCS3 expression inhibits IFN- γ , IFN- β , IFN- α , GH and leptin. Thus, expression of such genes in the T cell sample in the presence and absence of a test compound can routinely be determined using standard techniques.

The activity of Stat1, Stat2, Stat3, Stat4, Stat6, SOCS1 or SOCS3 can also be assessed by detecting the proliferation of the T cell sample, detecting the effector function of the sample or detecting differentiation of the sample. Techniques known to those of skill in the art can be used for measuring these activities. For example, cellular proliferation can be assayed by ³H-thymidine incorporation assays and trypan blue cell counts. The effector function of T-cells can be measured, for example, by a ⁵¹Cr-release assay (see, *e.g.*, Palladino et al., 1987, *Cancer Res.* 47:5074--5079 and Blachere et al., 1993, *J. Immunotherapy* 14:352-356).

As set forth above, the methods described herein for identifying compounds to be tested for an ability to reduce immune rejection assay whether a test compound has an effect on the expression and/or activity of Stat1 mRNA or protein, Stat2 mRNA or protein, Stat3 mRNA or protein, Stat4 mRNA or protein, Stat6 mRNA or protein, SOCS1 mRNA or protein, and/or SOCS3 mRNA or protein produced by a T cell, in particular, an activated T cell, or at a minimum, a T cell that has the ability to respond to exogenous cytokines.

The T cell used as part of the methods can be one that is constitutively activated (e.g., a constitutively activated T cell line), one that has or has gained the ability to respond to cytokines, one that is activated prior to performing the method, or one that is activated concurrently with the method. A T cell to be used as part of the methods described herein can be activated either prior to or simultaneously with contacting the cell with a test compound. With respect to activated, including constitutively activated T cells, activation of such T cells can, in certain instances be further enhanced by addition and contact with a T cell activator.

An activated T cell is one that expresses antigens indicative of T-cell activation (that is, T cell activation markers). Examples of T cell activation markers include, but are not limited to, CD25, CD26, CD30, CD38, CD69, CD70, CD71, ICOS, OX-40 and 4-1BB. The expression of activation markers can be measured by techniques known to those of skill in the art, including, for example, western blot analysis, northern blot analysis, RT-PCR, immunofluorescence assays, and fluorescence activated cell sorter (FACS) analysis. The activated T cell used as part of the present methods can be an activated T cell line or can be a primary cell that has been activated:

Activated T cell lines are well known to those of skill in the art. Examples of activated T cell lines include TH1 cell lines such as AE7, PL17, and OF6, and TH2 cell lines such as D10 and CDC35.

T cell lines that do not express the T cell activation markers required to constitute activation, but nonetheless have the ability to respond to cytokines are also well known to those of skill in the art. Examples of such T cell lines include CTLL-2 and HT-2.

Alternatively, primary T cells can be isolated, the majority of which will be in a resting state, and activated using standard techniques. For example, immune cells can be collected or isolated from blood, or secondary lymphoid organs of the subject, such as but not limited to lymph nodes, tonsils, the spleen, Peyer's patch of the intestine, and bone marrow, by any of the methods known in the art. Immune cells obtained from such sources typically comprise predominantly recirculating lymphocytes and macrophages at various stages of differentiation and maturation. Optionally, standard techniques, such as morphological observation and immunochemical staining, can be used, if desired, to verify

the presence of the desired cells, that is, T cells. In a preferred aspect, the immune cells used in the methods of the invention described herein are human peripheral blood compositions lacking red blood cells, *e.g.*, whole blood leukocytes (whole peripheral blood from which the red blood cells and serum have been substantially removed), which can be collected from a human subject by standard techniques, such as by use of a syringe to withdraw the blood, followed by subjecting the blood to Ficoll-Hypaque (Pharmacia) gradient centrifugation. Blood, anticoagulated with preservative-free heparin, usually yields 0.5 to 1×10^6 lymphocytes/ml. Separated blood cells (*e.g.*, leukocytes) may be frozen by standard techniques prior to use in the present methods. In a specific embodiment, the immune cells used are purified white blood cells comprising lymphocytes and macrophages.

In one embodiment wherein further purification of T cells is desired, antibodies against specific surface markers can be directly labeled by conjugation of a detectable compound to such antibodies to facilitate detection and separation of T cells. Alternatively, in another embodiment, the primary antibody is detected by detecting binding of a secondary antibody or reagent to the primary antibody. In a further embodiment, the secondary antibody is labeled. Examples of detectable compounds include, but are not limited to, biotin, photobiotin, fluorescein isothiocyanate (FITC), or phycoerythrin (PE), or other compounds known in the art. Cells retaining labeled antibodies are then separated from cells that do not bind such antibodies by techniques known in the art such as, but not limited to, various cell sorting methods (*e.g.*, FACS), affinity chromatography, and panning.

In another embodiment wherein further purification of T cells is desired, T cells are sorted using a fluorescence activated cell sorter (FACS). Fluorescence activated cell sorting (FACS) is a well-known method for separating particles, including cells, based on the fluorescent properties of the particles (Kamarch, 1987, *Methods Enzymol*, 151:150-165). Laser excitation of fluorescent moieties in the individual particles results in a small electrical charge allowing electromagnetic separation of positive and negative particles from a mixture.

In another embodiment wherein further purification of T cells is desired, magnetic beads can be used to separate T cells. T cells may be sorted using a magnetic activated cell sorting (MACS) technique, a method for separating particles based on their ability to bind magnetic beads (0.5-100 μ diameter; Dynal, Inc., Lake Success, NY) as undertaken according to the manufacturer's instructions. A variety of useful modifications can be performed on the magnetic microspheres, including covalent addition of antibody which specifically recognizes a cell-solid phase surface molecule or hapten. A magnetic field is then applied, to physically manipulate the selected beads. The beads are then mixed with

the immune cells to allow binding. Cells are then passed through a magnetic field to separate out cells having T cell surface markers.

The isolated resting T cells can then be activated by contacting with a T cell activator. Any T cell activator can be utilized for this purpose. For example, any compound or factor that is a T cell receptor stimulatory factor, that is, induces T cell receptor signalling can be used. Preferably, the compound or factor also induces co-stimulatory pathways. Representative, non-limiting examples of T cell activators include, but are not limited to, anti-CD3 antibodies (preferably monoclonal antibodies) either alone or in conjunction with anti-CD28 antibodies (preferably monoclonal antibodies), or mitogens such as, for example, phorbol 12-myristate 13-acetate (PMA), phytohemagglutinin (PHA), or concanavalin-A (ConA).

Compounds that can be tested and identified methods described herein can include, but are not limited to, compounds obtained from any commercial source, including Aldrich (1001 West St. Paul Ave., Milwaukee, WI 53233), Sigma Chemical (P.O. Box 14508, St. Louis, MO 63178), Fluka Chemie AG (Industriestrasse 25, CH-9471 Buchs, Switzerland (Fluka Chemical Corp. 980 South 2nd Street, Ronkonkoma, NY 11779)), Eastman Chemical Company, Fine Chemicals (P.O. Box 431, Kingsport, TN 37662), Boehringer Mannheim GmbH (Sandhofer Strasse 116, D-68298 Mannheim), Takasago (4 Volvo Drive, Rockleigh, NJ 07647), SST Corporation (635 Brighton Road, Clifton, NJ 07012), Ferro (111 West Irene Road, Zachary, LA 70791), Riedel-deHaen Aktiengesellschaft (P.O. Box D-30918, Seelze, Germany), PPG Industries Inc., Fine Chemicals (One PPG Place, 34th Floor, Pittsburgh, PA 15272). Further any kind of natural products may be screened using the methods of the invention, including microbial, fungal, plant or animal extracts.

Furthermore, diversity libraries of test compounds, including small molecule test compounds, may be utilized. For example, libraries may be commercially obtained from Specs and BioSpecs B.V. (Rijswijk, The Netherlands), Chembridge Corporation (San Diego, CA), Contract Service Company (Dolgoprudny, Moscow Region, Russia), Comgenex USA Inc. (Princeton, NJ), Maybridge Chemicals Ltd. (Cornwall PL34 OHW, United Kingdom), and Asinex (Moscow, Russia).

Still further, combinatorial library methods known in the art, can be utilize, including, but not limited to: biological libraries; spatially addressable parallel solid phase or solution phase libraries; synthetic library methods requiring deconvolution; the "one-bead one-compound" library method; and synthetic library methods using affinity chromatography selection. The biological library approach is limited to peptide libraries, while the other four approaches are applicable to peptide, non-peptide oligomer or small molecule libraries of compounds (Lam, 1997, Anticancer Drug Des. 12:145). combinatorial

libraries of test compounds, including small molecule test compounds, can be utilized, and may, for example, be generated as disclosed in Eichler & Houghten, 1995, Mol. Med. Today 1:174-180; Dolle, 1997, Mol. Divers. 2:223-236; and Lam, 1997, Anticancer Drug Des. 12:145-167.

- 5 Examples of methods for the synthesis of molecular libraries can be found in the art, for example in: DeWitt et al., 1993, Proc. Natl. Acad. Sci. USA 90:6909; Erb et al., 1994, Proc. Natl. Acad. Sci. USA 91:11422; Zuckermann et al., 1994, J. Med. Chem. 37:2678; Cho et al., 1993, Science 261:1303; Carrell et al., 1994, Angew. Chem. Int. Ed. Engl. 33:2059; Carell et al., 1994, Angew. Chem. Int. Ed. Engl. 33:2061; and Gallop et al., 1994, 10 J. Med. Chem. 37:1233.

- Libraries of compounds may be presented in solution (*e.g.*, Houghten, 1992, Bio/Techniques 13:412-421), or on beads (Lam, 1991, Nature 354:82-84), chips (Fodor, 1993, Nature 364:555-556), bacteria (U.S. Patent No. 5,223,409), spores (Patent Nos. 5,571,698; 5,403,484; and 5,223,409), plasmids (Cull et al., 1992, Proc. Natl. Acad. Sci. 15 USA 89:1865-1869) or phage (Scott and Smith, 1990, Science 249:386-390; Devlin, 1990, Science 249:404-406; Cwirla et al., 1990, Proc. Natl. Acad. Sci. USA 87:6378-6382; and Felici, 1991, J. Mol. Biol. 222:301-310).

- Among the test compounds that can be tested are compounds, including small organic molecule compounds that act as protein tyrosine kinase inhibitors, including, but 20 not limited to, the tyrphostin class of protein tyrosine kinase inhibitors (see, *e.g.*, Gazit et al. 1989, J. Medicinal Chem. 32:2344-2352; and Levitski, 1992, FASEB J. 6:3275). Among such tyrphostin compounds that can be tested are A25 (available, *e.g.*, from CalBiochem) or AG-490 (B42) (Meydan, N. et al., 1996, Nature 379:645-648) or derivatives thereof. Further, among the compounds that can be tested are compounds that interfere with SH2 25 domain interactions (see, *e.g.*, U.S. Patent No. 5,710,129; 5,776,902; and 5,580,97, or derivatives of compounds therein), *e.g.*, SH2-mediated Stat 4/IL-12 receptor- β_2 interactions. Still further, among the compounds that can be tested are compounds that interfere with Jak 2/IL-12 receptor- β_2 interactions and/or ones that interfere with Tyk 2/IL-12 receptor- β_1 interactions.

- 30 Upon identification of compounds to be tested for an ability to reduce immune rejection, the compounds can be further investigated. In particular, for example, the compounds identified via the present methods can be further tested *in vivo* in accepted animal models of transplant or autoimmune disorders. Further, the compounds identified can also be analyzed with respect to their specificity. In particular, the compounds can be 35 tested for an effect on platelet aggregation and/or on NF- κ B activation. Techniques for such additional compound investigation are described below.

Accepted animal models can be utilized to determine whether the compounds identified via the methods described herein. Such models can include both transplant-related models as well as autoimmune disorder models.

For example, the ability of a compound to reduce immune rejection, including the ability of the compound to induce tolerance in a subject mammal that has undergone a transplant can include, but are not limited to, a murine allograft model in which an allogeneic heart is transplanted into a subject mouse recipient (Hancock et al., 1998, *Nature Medicine* 4:1392-1396). In addition, primate models can also be tested. Such models include, for example, a primate renal allograft model (Kirk et al., 1997, *Proc. Natl. Acad. Sci. USA* 94:8789-8794). In addition, a graft versus host disease (GVHD) model can be used (see, e.g., Guillen et al., 1986, *Laboratory Investigation* 55:35-42). In such models, chronic and acute GVHD is made to result from introduction of donor cells into a host exhibiting disparate MHC alleles. The GVHD results, therefore, from the donor cells' response to such the host's disparate MHC alleles.

The ability of a compound to reduce immune rejection can also be tested in such autoimmune disorder models as, first, an experimental allergic encephalomyelitis (EAE) model. EAE is an experimental autoimmune disease of the central nervous system (CNS) (Zamvil et al, 1990, *Ann. Rev, Immunol.* 8:579) and is a disease model for the human autoimmune condition, multiple sclerosis (MS). EAE is an example of a cell-mediated autoimmune disorder that is mediated via T cells. No direct evidence exists for an autoantibody requirement in disease progression. EAE is readily induced in mammalian species by immunizations of myelin basic protein purified from the CNS or an encephalitogenic proteolipid (PLP). SJL/J mice are a susceptible strain of mice (H-2') and, upon induction of EAE, these mice develop an acute paralytic disease and an acute cellular infiltrate is identifiable within the CNS.

In addition, a collagen-induced arthritis (CIA) model can be utilized to determine whether the compound of interest reduce immune rejection. CIA is an animal model for the human autoimmune disease rheumatoid arthritis (RA) (Trenthorn et al., 1977, *J. Exp. Med.*, 146:857). This disease can be induced in many species by the administration of heterologous type II collagen (Courtenay et al., 1980, *Nature* 283:665; Cathcart et al, 1986, *Lab. Invest.*, 54:26). With respect to animal models of arthritis see, in addition, e.g., Holmdahl, R., 1999, *Curr. Biol.* 15:R528-530.

Still further, animal models for type 1 diabetes, thyroid autoimmunity or systemic lupus erythematosus, including glomerulonephritis can be utilized to determine whether the compound of interest reduces immune rejection (see, e.g., Flanders et al., 1999,

Autoimmunity 29:235-246; Krogh et al., 1999, Biochimie 81:511-515; and Foster, N.H., 1999, Semin. Nephrol. 19:12-24, respectively).

In addition, it is preferred that compounds to be utilized as therapeutic according to the methods described herein not induce platelet aggregation. Therefore, it is preferable that
5 compounds identified via the methods described herein that are to be tested for an ability to reduce immune rejection be further tested for an ability to induce platelet aggregation. *In vitro* and *ex vivo* assays for platelet aggregation are well known and compounds of interest can easily be tested via such assays.

Specifically, such assays include, but are not limited to the turbidometric method, in
10 which aggregation is measured as an increase in transmission of visible light through a stirred or agitated platelet suspension. See, e.g., Chanarin, L., 1989, Laboratory Haematology, Chapter 30, Churchill, Livingstone, London; and Schmidt, R.M. (ed), 1979, CRC Handbook Series in Clinical Laboratory Science, CRC Press, Inc.: Boca Raton, Florida.

Platelet aggregation can also be assayed via methods such as those described in U.S.
15 Patent 5,976,532. For example, in a non-limiting example of such a method, the platelet concentration in platelet-rich plasma obtained (PRP) obtained from blood samples is adjusted to 200,000 to 300,000/mm³. In an *in vitro* assay, the PRP is aliquoted and incubated in the presence or absence of a compound of interest for a period of time (e.g., 15
20 minutes at 37° C) prior to the addition of a platelet inducing agonist (e.g., ADP, thrombin, collagen, epinephrine, and ristocetin). In an *ex vivo* assay, the PRP obtained from individuals treated with the compound of interest or a placebo is aliquoted and incubated in the presence of a platelet inducing agonist (e.g., ADP, thrombin, collagen, epinephrine, and ristocetin). Platelet aggregation is measured by assessing an increase in the transmission of
25 visible light through a platelet suspension using a spectrophotometer.

It is also preferred that compounds to be utilized as therapeutic according to the methods described herein not affect NF- κ B activation, in particular, NF- κ B activation in CD40L⁺ cells. Therefore, it is preferable that compounds identified via the methods described herein that are to be tested for an ability to reduce immune rejection be further
30 tested for possible effect on NF- κ B activation in CD40L⁺ cells. In such tests, a CD40L⁺ cell is contacted with the compound of interest, and its effect on NF- κ B activation, if any is assayed, and compared to the level of NF- κ B activation in a corresponding control CD40L⁺ cell that has not been contacted with the compound.

Standard techniques can be utilized to test for NF- κ B activation. For example, the
35 activity of NF- κ B can be assessed by detecting the binding of NF- κ B to its cognate DNA binding element in an electromobility shift assay (EMSA), detecting the expression of a

gene whose expression is controlled by a promoter that is responsive to NF- κ B, detecting the induction of the expression of a reporter gene construct that comprises a regulatory element that is responsive to NF- κ B is operably linked to a nucleic acid encoding a detectable marker, *e.g.*, luciferase.

- 5 Genes whose expression is controlled by an NF- κ B-responsive promoter are well known, and include, for example granulocyte/macrophage colony-stimulating factor (GM-CSF), granulocyte colony-stimulating factor (G-CSF), IL-2, IL-6, IL-8, tumor necrosis factor- α (TNF- α), and intercellular cell adhesion molecule 1 (ICAM-1). See, *e.g.*, Baeuerle and Henkel, 1994, *Ann. Rev. Immunol.* 12:141-179. Thus, expression of such genes in
- 10 CD40L⁺ cells in the presence and absence of a compound of interest can routinely be determined using standard techniques. Expression can be determined using standard techniques. Preferably, the compound being tested will not increase such expression and, most preferably, will have no effect on such expression. Likewise, the structure of NF- κ B-responsive promoters are well known (see, *e.g.*, Baeuerle and Henkel, 1994, *Ann. Rev.*
- 15 *Immunol.* 12:141-179; and Thanos et al., 1995, *Cell* 80:529-532), making the construction and assay of NF- κ B reporter genes routine. Thus, the induction and expression of such reporter genes in CD40L⁺ cells in the presence and absence of a compound of interest can routinely be determined using standard techniques. Preferably, the compound being tested will not induce expression of the reporter gene.
- 20 EMSAs can also routinely be utilized to assess NF- κ B activity. Such techniques are well known to those of skill in the art. See, *e.g.*, Amici et al., 1995, *Cancer Research* 55: 14452-4457. Briefly, in a representative, non-limiting example, extracts of cells treated with a test composition or control composition are mixed with ³²P-NF- κ B element or a control oligonucleotide and poly(dI-dC) (Pharmacia Biotech Inc.) in binding buffer (*e.g.*,
- 25 Tris-Cl, pH 7.8, 50 mM NaCl, 1 mM EDTA, 0.5 mM dithiothreitol, 5% glycerol). After an approximately 20 minute incubation at room temperature, NF- κ B-DNA-complexes or control oligonucleotide complexes are analyzed by non-denaturing 4% polyacrylamide gel electrophoresis and autoradiography. The amount of shifted NF- κ B probe, an indicator NF- κ B activity, respectively, can be quantitated by Molecular Dynamics PhosphorImager
- 30 (MDP) analysis.

Further, the effect, if any, of the compound of interest on NF- κ B activation can also be tested by assaying for a cellular response, for example, cellular differentiation, or cell proliferation. Cellular proliferation can be assayed by, for example, ³H-thymidine incorporation assays and trypan blue cell counts. Preferably, the compound will have no

35 effect on such cellular responses.

5.3. Methods and Compositions for Reducing Immune Rejection

Described herein are methods and compositions for reducing immune rejection in a subject mammal. In particular, such methods comprise administration of compounds that serve to reduce the amount or activity of Stat1 mRNA or protein, Stat2 mRNA or protein, Stat4 mRNA or protein, SOCS1 mRNA or protein or SOCS3 mRNA or protein, and/or
5 increase the amount or activity of Stat6 mRNA or protein. The compounds utilized herein are ones that neither induce platelet aggregation nor affect NF- κ B activation.

Immune rejection in any subject mammal that has undergone a transplant, or that exhibits or is suspected of exhibiting an autoimmune disorder can be reduced using the
10 methods presented herein. Preferably, the mammal is a human, however, such subject mammals can also include, but are not limited to, other primates, including monkeys, as well as pigs, dogs, cats, horses, cattle, sheep, mice, rats, and rabbits.

The term "reducing immune rejection," is meant to encompass prevention or inhibition of immune rejection, as well as delaying the onset or the progression of immune
15 rejection. The term is also meant to encompass prolonging survival of a transplant in a subject mammal, or reversing failure of a transplant in a subject. Further, the term is meant to encompass ameliorating a symptom of an immune rejection, including, for example, ameliorating an immunological complication associated with immune rejection, such as for example, interstitial fibrosis, chronic graft atherosclerosis, or vasculitis. The term is also
20 meant to encompass induction of tolerance in a subject mammal that has undergone a transplant.

Specifically, the present invention relates to methods for reducing immune rejection in a subject mammal, said methods comprising: administering to a subject mammal in need of such a reduction a concentration of a compound sufficient to reduce the level or activity
25 of Stat4 mRNA or protein in the subject relative to that observed in the subject in the absence of the compound, wherein said compound does not induce platelet aggregation and does not affect NF- κ B activation in CD40L⁺ cells.

The present invention also relates to methods for reducing immune rejection in a subject mammal, said methods comprising: administering to a subject mammal in need of
30 such a reduction a concentration of a compound sufficient to reduce the level or activity of Stat1 mRNA or protein, Stat2 mRNA or protein, or Stat3 mRNA or protein in the subject relative to that observed in the subject in the absence of the compound, wherein said compound does not induce platelet aggregation and does not affect NF- κ B activation in CD40L⁺ cells.

35 Such methods can also include methods for reducing immune rejection in a subject mammal, comprising administering to the subject mammal in need of such a reduction a

concentration of a compound sufficient to decrease the level or activity of Stat4 mRNA or protein in the subject relative to that observed in the subject in the absence of the compound, and wherein the level or activity of Stat6 mRNA or protein in the subject is maintained or increased relative to that observed in the subject in the absence of the compound. Further, the compound administered is one that does not induce platelet aggregation or affect NF- κ B activation in CD-40L⁺ cells.

Alternatively, such methods for reducing immune rejection in a subject mammal can comprise: administering to a subject mammal in need of such a reduction a concentration of a compound sufficient to increase the level or activity of Stat6 mRNA or protein in the subject relative to that observed in the subject in the absence of the compound, wherein said compound does not induce platelet aggregation and does not affect NF- κ B activation in CD40L⁺ cells.

Such methods for reducing immune rejection in a subject mammal can also comprise: administering to a subject mammal in need of such a reduction a concentration of a compound sufficient to decrease the level or activity of Stat4 mRNA or protein and maintain or increase the level or activity of Stat6 mRNA or protein in the subject subject relative to that observed in the subject in the absence of the compound, wherein said compound does not induce platelet aggregation and does not affect NF- κ B activation in CD40L⁺ cells.

Generally, practice of these methods does not solely entail administration of compositions that are considered signal 2-type blockers (see, *e.g.*, Gummert J.F., et al., 1999, J. Am. Soc. Nephrol. 10: 1366), that is, compounds (*e.g.*, CD40L antibodies) that act to inhibit CD40/CD40L (CD154) interactions or B7/CD28 interactions.

It is noted, however, that embodiments of the present invention further include combinatorial immune reduction therapy utilizing compositions as taught herein in conjunction with immunosuppressive or immunomodulatory drug therapies, as described in detail, below.

The methods of the present invention for reducing immune rejection can be utilized, *e.g.*, for reducing immune rejection in a subject mammal that has undergone a transplant. For example, such methods can induce tolerance in a subject mammal that has undergone a transplant. Such methods can be used to reduce immune reject in a transplant situation involving any cell, organ, organ system or tissue which can elicit an immune response in a recipient subject mammal. In general, therefore, a transplant includes an allograft, or a xenograft cell, organ, organ system or tissue. An allograft refers to a graft (cell, organ, organ system or tissue) obtained from a member of the same species as the recipient. A xenograft refers to a graft (cell, organ, organ system or tissue) obtained from a member of a

different species as the recipient. In particular, the transplant can, for example, be an allograft heart, liver, kidney, lung, bone marrow, skin, muscle, pancreatic islet, intestine or cornea transplant.

The methods of the present invention for reducing immune rejection can also be
5 utilized, *e.g.*, for reducing immune rejection in a subject mammal exhibiting an autoimmune disorder. Thus, the present invention can treat an autoimmune disorder affecting any body cell, tissue, organ or organ system, including but not limited to cutaneous, cardiac, pericardial, endocardial, vascular lining or wall, blood, blood-forming (*e.g.*, marrow or spleen), endocrine (*e.g.*, pancreatic or thyroid), gastrointestinal (*e.g.*, bowel), respiratory
10 (*e.g.*, lung), renal, central nervous system, peripheral nervous system, muscular or skeletal joint (*e.g.*, articular cartilage or synovial) tissue. The methods and compositions of the present invention can, therefore, be utilized to treat any autoimmune disorder including, but not limited to atopic dermatitis, contact dermatitis, eczematous dermatitides, seborrheic dermatitis, Lichen planus, Pemphigus, bullous pemphigus, Epidermolysis bullosa,
15 Alopecia areata, urticaria, angioedemas, erythema, eosinophilias, migraine, lupus, including cutaneous lupus (discoid lupus erythematosus), extracutaneous lupus, including systemic lupus erythematosus, acute lupus, lupus annularis, lupus discretus, lupus lymphaticus, lupus papillomatus, lupus psoriasis, lupus vulgaris, lupus sclerosis, neonatal lupus erythematosus, and drug-induced lupus; anti-phospholipid syndrome (APS), hemolytic anemia (HA),
20 idiopathic thrombocytopenia (ITP), thyroiditis, diabetes mellitus (DM), inflammatory bowel disease, *e.g.*, Crohn's disease or ulcerative colitis, rhinitis, uveitis, nephrotic syndrome, demyelinating diseases such as multiple sclerosis (MS), myasthenia gravis (MG), and arthritis, *e.g.*, rheumatoid arthritis, non-rheumatoid inflammatory arthritis, arthritis associated with Lyme disease, or osteoarthritis.

25 The compounds utilized as part of these methods include, but are not limited to, ones identified via the methods described above. A number of different points along the Jak/Stat pathway can be targeted by the compounds utilized as part of the methods for reducing immune rejection described herein. Administration methods, including gene therapy methods, and pharmaceutical preparations by which such compounds can routinely
30 be utilized as part of methods for reducing immune rejection are taught below.

For example, compounds that specifically downregulate Stat4 mRNA or protein levels or activity, while not affecting NF- κ B activation or platelet aggregation can be utilized as part of these methods. In addition, compounds that specifically downregulate Stat1 mRNA or protein levels or activity, while not affecting NF- κ B activation or platelet
35 aggregation can be utilized as part of these methods. Also, compounds that specifically downregulate Stat2 mRNA or protein levels or activity, while not affecting NF- κ B

activation or platelet aggregation can be utilized as part of these methods. In addition, compounds that specifically downregulate Stat3 mRNA or protein levels or activity, while not affecting NF- κ B activation or platelet aggregation can be utilized as part of these methods. In addition, compounds that specifically downregulate SOCS1 mRNA or protein levels or activity, while not affecting NF- κ B activation or platelet aggregation can be utilized as part of these methods. Likewise, compounds that specifically downregulate SOCS3 mRNA or protein levels or activity, while not affecting NF- κ B activation or platelet aggregation can be utilized as part of these methods. In addition, compounds or methods that specifically increase Stat 6 mRNA or protein levels or activity, while not affecting NF- κ B activation or platelet aggregation can be utilized as part of these methods. Representative, non-limiting examples of such compounds are described in detail below.

First, such compounds can include, for example, antisense, ribozyme, or triple helix compounds that can downregulate the expression or Stat1, Stat2, Stat3, Stat4, SOCS1 or SOCS3. Such compounds are described in detail in the subsection below.

Second, such compounds can include, for example, antibody compositions that can downregulate the expression or activity of Stat1, Stat2, Stat3, Stat4, SOCS1 or SOCS3. Such compounds are described in detail in the subsection below.

Further, among such compounds are ones, including ones administered via gene therapy techniques, that serve to upregulate Stat6 expression or activity, and compounds that act in maintaining Stat6 expression or activity levels as Stat4 activity or expression levels are decreased.

Compositions can include, for example ones can be utilized that compete with Stat4 for binding to the IL-12 receptor β_2 ("IL-12R β_2 "). Examples of such compounds include, but are not limited to, peptide compositions as in Naeger, L.K. et al., 1999, J. Biol. Chem. 274:1875-1878. Additional examples of compounds that can be utilized include compounds, such as small organic compounds that act as inhibitors of SH2 domain-mediated interactions such as SH2-mediated Stat4/IL-12R β_2 interactions. One example of such an embodiment involves a composition of the invention comprises one or more peptides that bind to the Stat4 SH2 domain which prevent Stat4 from binding to the IL-12R β_2 , or that comprise one or more dominant-negative Stat4 polypeptides (e.g., a Stat4 polypeptide lacking its SH2 domain or a Stat4 polypeptide lacking its DNA binding domain). Examples of such polypeptides include, but are not limited to, (using the standard one-letter amino acid code) phospho-YLPSNID peptides (Naeger, L.K. et al., 1999, J. Biol. Chem. 274:1875-1878).

In specific embodiments, Stat1 antisense oligonucleotides, Stat2 antisense oligonucleotides, Stat3 antisense oligonucleotides, or any combination thereof, are

administered to reduce immune rejection. In other embodiments one or more anti-Stat1 antibodies, anti-Stat2 antibodies, or anti-Stat3 antibodies are administered to reduce immune rejection. In other embodiments, one or more peptides that compete with Jak1 or Tyk2 for binding to the IFN- α receptor or IFN- β receptor are administered to reduce immune rejection. In yet another embodiment, one or more peptides that compete with Jak1 or Jak2 for binding to the IFN- γ receptor are administered to reduce immune rejection. In other embodiments, one or more peptides that compete with Stat1 or Stat2 for binding to the IFN- α receptor or IFN- β receptor are administered to reduce immune rejection. In other embodiments, one or more peptides that compete with Stat1 for binding to the IFN- γ receptor are administered to reduce immune rejection. In yet other embodiments, one or more dominant-negative Stat1 polypeptides (*e.g.*, a Stat1 polypeptide lacking its SH2 domain or a Stat1 polypeptide lacking its DNA binding domain), dominant-negative Stat2 polypeptides (*e.g.*, a Stat2 polypeptide lacking its SH2 domain or a Stat2 polypeptide lacking its DNA binding domain), or dominant-negative Stat3 polypeptides (*e.g.*, a Stat3 polypeptide lacking its SH2 domain or a Stat3 polypeptide lacking its DNA binding domain) are administered to reduce immune rejection.

Jak2 is involved in activation of Stat4 protein. In view of this, another composition that can be utilized as part of the methods of the invention comprises a composition that reduces the expression or activity of Jak2, while not affecting NF- κ B activation or platelet aggregation. In one embodiment, therefore, one or more peptides that compete with Jak2 for binding to the IL-12R β_2 can be utilized. In other embodiments, such compounds include Jak2 antisense molecules, triple helix molecules or ribozyme molecules that serve to downregulate the expression of Jak2. Representative antisense compositions are described in detail below. Such compounds also include antibodies or fragments thereof that specifically bind to and inhibit the activity of Jak2.

Tyk2 is also involved in activation of Stat4 protein. In view of this, another composition that can be utilized as part of the methods of the invention comprises a composition that reduces the expression or activity of Tyk2, while not affecting NF- κ B activation or platelet aggregation. In another embodiment, a composition of the invention comprises one or more peptides that compete with Tyk2 for binding to the IL-12R β_1 . In other embodiments, such compounds include Tyk2 antisense molecules, triple helix molecules or ribozyme molecules that serve to downregulate the expression of Tyk2. Representative antisense compositions are described in detail below.

In yet another embodiment, a composition that can be utilized as part of these methods comprises one or more small molecules that decrease or downregulate Stat4 expression or activity, while not affecting NF- κ B activation or platelet aggregation. For

example, among the compounds that can be utilized as part of these methods are protein tyrosine kinase inhibitors, including, but not limited to the tyrphostin class of protein tyrosine kinase inhibitors. Preferable tyrphostin compositions are ones that inhibit or downregulate Stat4 activity by (without wishing to be bound by any particular mechanism) inhibiting Jak2 or Tyk2 protein tyrosine kinase activity without deleterious effects on normal hematopoiesis. In specific embodiments, the tyrphostin is AG-490 (B42), although it is preferred that this particular tyrphostin not be utilized for treatment of autoimmune disorders, specifically multiple sclerosis (MS).

10 ANTISENSE, RIBOZYME, TRIPLE-HELIX COMPOSITIONS

Representative, non-limiting examples of Stat1 antisense molecules include the following: 5'- GCT GAA GCT CGA ACC ACT GTG ACA TCC - 3' (SEQ ID NO:19); and 5'-AAG TTC GTA CCA CTG AGA CAT CCT GCC (SEQ ID NO:20).

15 Representative, non-limiting examples of Stat2 antisense molecules include the following: 5'- CAT CTC CCA CTG CGC CAT TTG GAC TCT TCA -3' (SEQ ID NO:21); and 5'-CAG CAT TTC CCA CTG CGC CAT TTG GGC-3' (SEQ ID NO:22).

Representative, non-limiting examples of Stat3 antisense molecules include the following: 5'- CTG GTT CCA CTG AGC CAT CCT GCT GCA TCAG - 3' (SEQ ID NO:-23); and 5'-CTG TAG CTG ATT CCA TTG GGC CAT CCT-3' (SEQ ID NO:24).

20 Representative, non-limiting examples of Stat4 antisense molecules include the following: 5'- GAT TCC ACT GAG ACA TGC TGC TCT CTC TCT C-3' (SEQ ID NO:25); and 5'-GAC TTG ATT CCA CTG AGA CAT GCT AGC-3' (SEQ ID NO:26).

Representative, non-limiting examples of Jak2 antisense molecules include the following: 5'- GCC AGG CCA TTC CCA TCT AGA GCT TTT TTC -3' (SEQ ID NO:27); 25 and 5'-CGT AAG GCA GGC CAT TCC CAT GCA GAG-3' (SEQ ID NO:28).

Representative, non-limiting examples of Tyk2 antisense molecules include the following: 5'- CCC ACA CAG AGG CAT GGT CCC CAC CAT TCA -3' (SEQ ID NO:29); and 5'-GGC CAT CCC CCA GTG GCG CAG AGG CAT GCT CCC-3' (SEQ ID NO:30) .

30 Representative, non-limiting examples of SOCS1 antisense molecules include the following: 5'- CCT GGT TGC GTG CTA CCA TCC TAC TCG AGG GGC -3' (SEQ ID NO:31); and 5'-CAC CTG GTT GTG TGC TAC CAT CCT ACT-3' (SEQ ID NO:32).

Representative, non-limiting examples of SOCS3 antisense molecules include the following: 5'- GCT GTG GGT GAC CAT GGC GCA CGG AGC CAG CG -3' (SEQ ID 35 NO:33); and 5'- GGC GGG AAA CTT GCT GTG GGT GAC CAT-3' (SEQ ID NO:34).

In addition, standard techniques can be utilized to produce antisense, triple helix, or ribozyme molecules for use as part of the methods described herein.

First, standard techniques can be utilized for the production of antisense nucleic acid molecules, *i.e.*, molecules which are complementary to a sense nucleic acid encoding a polypeptide of interest (*e.g.*, Stat1, Stat2, Stat3, Stat4, Jak2, Tyk2, SOCS1, or SOCS3), *e.g.*,
5 complementary to the coding strand of a double-stranded cDNA molecule or complementary to an mRNA sequence. Accordingly, an antisense nucleic acid can hydrogen bond to a sense nucleic acid. The antisense nucleic acid can be complementary to an entire coding strand, or to only a portion thereof, *e.g.*, all or part of the protein coding
10 region (or open reading frame). An antisense nucleic acid molecule can be antisense to all or part of a non-coding region of the coding strand of a nucleotide sequence encoding a polypeptide of interest. The non-coding regions ("5' and 3' untranslated regions") are the 5' and 3' sequences which flank the coding region and are not translated into amino acids.

An antisense oligonucleotide can be, for example, about 5, 10, 15, 20, 25, 30, 35, 40,
15 45 or 50 nucleotides or more in length. An antisense nucleic acid of the invention can be constructed using chemical synthesis and enzymatic ligation reactions using procedures known in the art. For example, an antisense nucleic acid (*e.g.*, an antisense oligonucleotide) can be chemically synthesized using naturally occurring nucleotides or variously modified nucleotides designed to increase the biological stability of the molecules or to increase the
20 physical stability of the duplex formed between the antisense and sense nucleic acids, *e.g.*, phosphorothioate derivatives and acridine substituted nucleotides can be used. Examples of modified nucleotides which can be used to generate the antisense nucleic acid include 5-fluorouracil, 5-bromouracil, 5-chlorouracil, 5-iodouracil, hypoxanthine, xanthine, 4-acetylcytosine, 5-(carboxyhydroxymethyl) uracil,
25 5-carboxymethylaminomethyl-2-thiouridine, 5-carboxymethylaminomethyluracil, dihydrouracil, beta-D-galactosylqueosine, inosine, N6-isopentenyladenine, 1-methylguanine, 1-methylinosine, 2,2-dimethylguanine, 2-methyladenine, 2-methylguanine, 3-methylcytosine, 5-methylcytosine, N6-adenine, 7-methylguanine, 5-methylaminomethyluracil, 5-methoxyaminomethyl-2-thiouracil,
30 beta-D-mannosylqueosine, 5'-methoxycarboxymethyluracil, 5-methoxyuracil, 2-methylthio-N6-isopentenyladenine, uracil-5-oxyacetic acid (v), wybutoxosine, pseudouracil, queosine, 2-thiocytosine, 5-methyl-2-thiouracil, 2-thiouracil, 4-thiouracil, 5-methyluracil, uracil-5-oxyacetic acid methylester, uracil-5-oxyacetic acid (v), 5-methyl-2-thiouracil, 3-(3-amino-3-N-2-carboxypropyl) uracil, (acp3)w, and
35 2,6-diaminopurine. Alternatively, the antisense nucleic acid can be produced biologically using an expression vector into which a nucleic acid has been subcloned in an antisense

orientation (*i.e.*, RNA transcribed from the inserted nucleic acid will be of an antisense orientation to a target nucleic acid of interest).

Antisense nucleic acid molecules administered to a subject or generated *in situ* such that they hybridize with or bind to cellular mRNA encoding the polypeptide of interest to thereby inhibit expression, *e.g.*, by inhibiting transcription and/or translation. The hybridization can be by conventional nucleotide complementarity to form a stable duplex, or, for example, in the case of an antisense nucleic acid molecule which binds to DNA duplexes, through specific interactions in the major groove of the double helix. An example of a route of administration of antisense nucleic acid molecules of the invention includes direct injection at a tissue, *e.g.*, transplant or autoimmune lesion, site. Alternatively, antisense nucleic acid molecules can be modified to target selected cells and then administered systemically. For example, for systemic administration, antisense molecules can be modified such that they specifically bind to receptors or antigens expressed on a selected cell, *e.g.*, T cell, surface, *e.g.*, by linking the antisense nucleic acid molecules to peptides or antibodies which bind to cell surface receptors or antigens. The antisense nucleic acid molecules can also be delivered to cells using vectors, *e.g.*, gene therapy vectors, described below. To achieve sufficient intracellular concentrations of the antisense molecules, vector constructs in which the antisense nucleic acid molecule is placed under the control of a strong pol II or pol III promoter are preferred.

An antisense nucleic acid molecule of interest can be an α -anomeric nucleic acid molecule. An α -anomeric nucleic acid molecule forms specific double-stranded hybrids with complementary RNA in which, contrary to the usual β -units, the strands run parallel to each other (Gaultier et al. (1987) *Nucleic Acids Res.* 15:6625-6641). The antisense nucleic acid molecule can also comprise a 2'-o-methylribonucleotide (Inoue et al. (1987) *Nucleic Acids Res.* 15:6131-6148) or a chimeric RNA-DNA analogue (Inoue et al. (1987) *FEBS Lett.* 215:327-330).

Ribozymes are catalytic RNA molecules with ribonuclease activity which are capable of cleaving a single-stranded nucleic acid, such as an mRNA, to which they have a complementary region, and can also be generated using standard techniques. Thus, ribozymes (*e.g.*, hammerhead ribozymes (described in Haselhoff and Gerlach (1988) *Nature* 334:585-591)) can be used to catalytically cleave mRNA transcripts to thereby inhibit translation of the protein encoded by the mRNA. A ribozyme having specificity for a nucleic acid molecule encoding a polypeptide of interest can be designed based upon the nucleotide sequence of a cDNA disclosed herein. For example, a derivative of a *Tetrahymena* L-19 IVS RNA can be constructed in which the nucleotide sequence of the active site is complementary to the nucleotide sequence to be cleaved in a Cech et al. U.S.

Patent No. 4,987,071; and Cech et al. U.S. Patent No. 5,116,742. Alternatively, an mRNA encoding a polypeptide of interest can be used to select a catalytic RNA having a specific ribonuclease activity from a pool of RNA molecules. *See, e.g.,* Bartel and Szostak (1993) *Science* 261:1411-1418.

- 5 Triple helical structures can also be generated using well known techniques. For example, expression of a polypeptide of interest can be inhibited by targeting nucleotide sequences complementary to the regulatory region of the gene encoding the polypeptide (*e.g.,* the promoter and/or enhancer) to form triple helical structures that prevent transcription of the gene in target cells. *See generally* Helene (1991) *Anticancer Drug Des.* 6(6):569-84; Helene (1992) *Ann. N.Y. Acad. Sci.* 660:27-36; and Maher (1992) *Bioassays* 14(12):807-15.

In various embodiments, nucleic acid compositions can be modified at the base moiety, sugar moiety or phosphate backbone to improve, *e.g.,* the stability, hybridization, or solubility of the molecule. For example, the deoxyribose phosphate backbone of the nucleic
15 acids can be modified to generate peptide nucleic acids (*see* Hyrup et al. (1996) *Bioorganic & Medicinal Chemistry* 4(1): 5-23). As used herein, the terms "peptide nucleic acids" or "PNAs" refer to nucleic acid mimics, *e.g.,* DNA mimics, in which the deoxyribose phosphate backbone is replaced by a pseudopeptide backbone and only the four natural nucleobases are retained. The neutral backbone of PNAs has been shown to allow for
20 specific hybridization to DNA and RNA under conditions of low ionic strength. The synthesis of PNA oligomers can be performed using standard solid phase peptide synthesis protocols as described in Hyrup et al. (1996), *supra*; Perry-O'Keefe et al. (1996) *Proc. Natl. Acad. Sci. USA* 93: 14670-675.

PNAs can, for example, be modified, *e.g.,* to enhance their stability or cellular
25 uptake, by attaching lipophilic or other helper groups to PNA, by the formation of PNA-DNA chimeras, or by the use of liposomes or other techniques of drug delivery known in the art. For example, PNA-DNA chimeras can be generated which may combine the advantageous properties of PNA and DNA. Such chimeras allow DNA recognition enzymes, *e.g.,* RNase H and DNA polymerases, to interact with the DNA portion while the
30 PNA portion would provide high binding affinity and specificity. PNA-DNA chimeras can be linked using linkers of appropriate lengths selected in terms of base stacking, number of bonds between the nucleobases, and orientation (Hyrup (1996), *supra*). The synthesis of PNA-DNA chimeras can be performed as described in Hyrup (1996), *supra*, and Finn et al. (1996) *Nucleic Acids Res.* 24(17):3357-63. For example, a DNA chain can be synthesized
35 on a solid support using standard phosphoramidite coupling chemistry and modified nucleoside analogs. Compounds such as 5'-(4-methoxytrityl)amino-5'-deoxy-thymidine

phosphoramidite can be used as a link between the PNA and the 5' end of DNA (Mag et al. (1989) *Nucleic Acids Res.* 17:5973-88). PNA monomers are then coupled in a stepwise manner to produce a chimeric molecule with a 5' PNA segment and a 3' DNA segment (Finn et al. (1996) *Nucleic Acids Res.* 24(17):3357-63). Alternatively, chimeric molecules
5 can be synthesized with a 5' DNA segment and a 3' PNA segment (Peterser et al. (1975) *Bioorganic Med. Chem. Lett.* 5:1119-11124).

In other embodiments, the oligonucleotide may include other appended groups such as peptides (*e.g.*, for targeting host cell receptors *in vivo*), or agents facilitating transport across the cell membrane (*see, e.g.*, Letsinger et al. (1989) *Proc. Natl. Acad. Sci. USA*
10 86:6553-6556; Lemaitre et al. (1987) *Proc. Natl. Acad. Sci. USA* 84:648-652; PCT Publication No. W0 88/09810) or the blood-brain barrier (*see, e.g.*, PCT Publication No. W0 89/10134). In addition, oligonucleotides can be modified with hybridization-triggered cleavage agents (*see, e.g.*, Krol et al. (1988) *Bio/Techniques* 6:958-976) or intercalating agents (*see, e.g.*, Zon (1988) *Pharm. Res.* 5:539-549). To this end, the oligonucleotide may
15 be conjugated to another molecule, *e.g.*, a peptide, hybridization triggered cross-linking agent, transport agent, hybridization-triggered cleavage agent, etc.

ANTIBODY COMPOSITIONS

In one embodiment, anti-Stat1 antibodies, anti-Stat2 antibodies, anti-Stat3
20 antibodies, anti-Stat4 antibodies, anti-SOCS1 antibodies or anti-SOCS3 antibodies are administered to a mammal, preferably a human, to reduce immune rejection. In another embodiment, any combination of anti-Stat1 antibodies, anti-Stat2 antibodies, anti-Stat3 antibodies, anti-Stat4, anti-SOCS1 antibodies and anti-SOCS3 antibodies are administered to a mammal, preferably a human, to reduce immune rejection. In a preferred embodiment,
25 anti-Stat1 antibodies, anti-Stat2 antibodies, anti-Stat3 antibodies, anti-Stat4 antibodies, anti-SOCS1 antibodies or anti-SOCS3 antibodies are administered to a mammal, preferably a human, in combination with other types of treatments (*e.g.*, immunosuppressive agents) to reduce immune rejection. In yet another preferred embodiment, any combination of anti-Stat1 antibodies, anti-Stat2 antibodies, anti-Stat3 antibodies, anti-Stat4, anti-SOCS1
30 antibodies and anti-SOCS3 antibodies are administered to a mammal, preferably a human, in combination with other types of treatments (*e.g.*, immunosuppressive agents) to reduce immune rejection.

Anti-Stat1 antibodies, anti-Stat2 antibodies, anti-Stat3 antibodies, anti-Stat4
antibodies, anti-SOCS1 antibodies, anti-SOCS3 antibodies, or any combination thereof can
35 be administered to a mammal, preferably a human, using various delivery systems are known to those of skill in the art. For example, anti-Stat1 antibodies, anti-Stat2 antibodies,

anti-Stat3 antibodies, anti-Stat4 antibodies, anti-SOCS1 antibodies, anti-SOCS3 antibodies, or any combination thereof can be administered by encapsulation in liposomes, microparticles or microcapsules. See, *e.g.*, U.S. Patent No. 5,762,904, U.S. Patent No. 6,004,534, and PCT Publication WO 99/52563. In addition, anti-Stat1 antibodies, anti-
5 Stat2 antibodies, anti-Stat3 antibodies, anti-Stat4 antibodies, anti-SOCS1 antibodies, anti-SOCS3 antibodies, or any combination thereof can be administered using recombinant cells capable of expressing the antibodies, or retroviral, other viral vectors or non-viral vectors capable of expressing the antibodies.

Anti-Stat1 antibodies, anti-Stat2 antibodies, anti-Stat3 antibodies, anti-Stat4, anti-
10 SOCS1 antibodies and anti-SOCS3 antibodies can be obtained from any known source. For example, anti-Stat1 antibodies, anti-Stat2 antibodies, anti-Stat3 antibodies, anti-Stat4, anti-SOCS1 antibodies and anti-SOCS3 antibodies can be obtained from Santa Cruz Biotechnology, Inc. (Santa Cruz, CA), Research Diagnostics, Inc. (Flanders, NJ) or Zymed Laboratories (South San Francisco, CA). Alternatively, anti-Stat1 antibodies, anti-Stat2
15 antibodies, anti-Stat3 antibodies, anti-Stat4, anti-SOCS1 antibodies and anti-SOCS3 antibodies can be produced by any method known in the art for the synthesis of antibodies, in particular, by chemical synthesis or preferably, by recombinant expression techniques.

Antibodies of the invention include, but are not limited to, polyclonal, monoclonal, multispecific, human, humanized or chimeric antibodies, single chain antibodies, Fab
20 fragments, F(ab') fragments, fragments produced by a Fab expression library, anti-idiotypic (anti-Id) antibodies (including, *e.g.*, anti-Id antibodies to antibodies of the invention), and epitope-binding fragments. The term "antibody," as used herein, refers to immunoglobulin molecules and immunologically active portions of immunoglobulin molecules, *i.e.*, molecules that contain an antigen binding site that immunospecifically binds an antigen.
25 The immunoglobulin molecules of the invention can be of any type (*e.g.*, IgG, IgE, IgM, IgD, IgA and IgY), class (*e.g.*, IgG₁, IgG₂, IgG₃, IgG₄, IgA₁ and IgA₂) or subclass of immunoglobulin molecule. Examples of immunologically active portions of immunoglobulin molecules include F(ab) and F(ab')₂ fragments which can be generated by treating the antibody with an enzyme such as pepsin or papain.

30 An isolated Stat1, Stat2, Stat3, Stat4, SOCS1 or SOCS3, or a fragment thereof, can be used as an immunogen to generate antibodies using standard techniques for polyclonal and monoclonal antibody preparation. The full-length Stat1, Stat2, Stat3, Stat4, SOCS1 or SOCS3 polypeptide or protein can be used or, alternatively, the invention provides antigenic peptide fragments of Stat1, Stat2, Stat3, Stat4, SOCS1 or SOCS3 for use as
35 immunogens. An antigenic peptide comprises at least 8 (preferably 10, 15, 20, or 30) amino acid residues of the amino acid sequence of Stat1, Stat2, Stat3, Stat4, SOCS1 or SOCS3,

and encompasses an epitope of Stat1, Stat2, Stat3, Stat4, SOCS1 or SOCS3 such that an antibody raised against the peptide forms a specific immune complex with Stat1, Stat2, Stat3, Stat4, SOCS1 or SOCS3.

5 The antibody titer in the immunized subject can be monitored over time by standard techniques, such as with an enzyme linked immunosorbent assay (ELISA) using immobilized polypeptide. If desired, the antibody molecules can be isolated from the mammal (*e.g.*, from the blood) and further purified by well-known techniques, such as protein A chromatography to obtain the IgG fraction. Alternatively, antibodies specific for a protein or polypeptide of the invention can be selected for (*e.g.*, partially purified) or
10 purified by, *e.g.*, affinity chromatography. For example, a recombinantly expressed and purified (or partially purified) protein of the invention is produced as described herein, and covalently or non-covalently coupled to a solid support such as, for example, a chromatography column. The column can then be used to affinity purify antibodies specific for the proteins of the invention from a sample containing antibodies directed against a large
15 number of different epitopes, thereby generating a substantially purified antibody composition, *i.e.*, one that is substantially free of contaminating antibodies. By a substantially purified antibody composition is meant, in this context, that the antibody sample contains at most only 30% (by dry weight) of contaminating antibodies directed against epitopes other than those on the desired Stat1, Stat2, Stat3, Stat4, SOCS1 or SOCS3
20 protein or polypeptide, and preferably at most 20%, yet more preferably at most 10%, and most preferably at most 5% (by dry weight) of the sample is contaminating antibodies. A purified antibody composition means that at least 99% of the antibodies in the composition are directed against the desired Stat1, Stat2, Stat3, Stat4, SOCS1 or SOCS3 protein or polypeptide.

25 At an appropriate time after immunization, *e.g.*, when the specific antibody titers are highest, antibody-producing cells can be obtained from the subject and used to prepare monoclonal antibodies by standard techniques, such as the hybridoma technique originally described by Kohler and Milstein (1975) *Nature* 256:495-497, the human B cell hybridoma technique (Kozbor et al. (1983) *Immunol. Today* 4:72), the EBV-hybridoma technique (Cole
30 et al. (1985), *Monoclonal Antibodies and Cancer Therapy*, Alan R. Liss, Inc., pp. 77-96) or trioma techniques. The technology for producing hybridomas is well known (*see generally* *Current Protocols in Immunology* (1994) Coligan et al. (eds.) John Wiley & Sons, Inc., New York, NY). Hybridoma cells producing a monoclonal antibody of the invention are detected by screening the hybridoma culture supernatants for antibodies that bind the
35 polypeptide of interest, *e.g.*, using a standard ELISA assay.

Alternative to preparing monoclonal antibody-secreting hybridomas, a monoclonal antibody directed against Stat1, Stat2, Stat3, Stat4, SOCS1 or SOCS3 polypeptide can be identified and isolated by screening a recombinant combinatorial immunoglobulin library (e.g., an antibody phage display library) with Stat1, Stat2, Stat3, Stat4, SOCS1 or SOCS3.

5 Kits for generating and screening phage display libraries are commercially available (e.g., the Pharmacia *Recombinant Phage Antibody System*, Catalog No. 27-9400-01; and the Stratagene *SurfZAP™ Phage Display Kit*, Catalog No. 240612). Additionally, examples of methods and reagents particularly amenable for use in generating and screening antibody display library can be found in, for example, U.S. Patent No. 5,223,409; PCT Publication

10 No. WO 92/18619; PCT Publication No. WO 91/17271; PCT Publication No. WO 92/20791; PCT Publication No. WO 92/15679; PCT Publication No. WO 93/01288; PCT Publication No. WO 92/01047; PCT Publication No. WO 92/09690; PCT Publication No. WO 90/02809; Fuchs et al. (1991) *Bio/Technology* 9:1370-1372; Hay et al. (1992) *Hum. Antibod. Hybridomas* 3:81-85; Huse et al. (1989) *Science* 246:1275-1281; Griffiths et al.

15 (1993) *EMBO J.* 12:725-734.

Additionally, recombinant antibodies, such as chimeric and humanized monoclonal antibodies, comprising both human and non-human portions, can be made using standard recombinant DNA techniques. A chimeric antibody is a molecule in which different portions are derived from different animal species, such as those having a variable region

20 derived from a murine mAb and a human immunoglobulin constant region. (See, e.g., Cabilly et al., U.S. Patent No. 4,816,567; and Boss et al., U.S. Patent No. 4,816,397, which are incorporated herein by reference in their entirety.) Humanized antibodies are antibody molecules from non-human species having one or more complementarily determining regions (CDRs) from the non-human species and a framework region from a human

25 immunoglobulin molecule. (See, e.g., Queen, U.S. Patent No. 5,585,089, which is incorporated herein by reference in its entirety.) Such chimeric and humanized monoclonal antibodies can be produced by recombinant DNA techniques known in the art, for example using methods described in PCT Publication No. WO 87/02671; European Patent

Application 184,187; European Patent Application 171,496; European Patent Application

30 173,494; PCT Publication No. WO 86/01533; U.S. Patent No. 4,816,567; European Patent Application 125,023; Better et al. (1988) *Science* 240:1041-1043; Liu et al. (1987) *Proc. Natl. Acad. Sci. USA* 84:3439-3443; Liu et al. (1987) *J. Immunol.* 139:3521-3526; Sun et al. (1987) *Proc. Natl. Acad. Sci. USA* 84:214-218; Nishimura et al. (1987) *Canc. Res.* 47:999-1005; Wood et al. (1985) *Nature* 314:446-449; and Shaw et al. (1988) *J. Natl.*

35 *Cancer Inst.* 80:1553-1559; Morrison (1985) *Science* 229:1202-1207; Oi et al. (1986) *Bio/Techniques* 4:214; U.S. Patent 5,225,539; Jones et al. (1986) *Nature* 321:552-525;

Verhoeyan et al. (1988) *Science* 239:1534; and Beidler et al. (1988) *J. Immunol.* 141:4053-4060.

Completely human antibodies are particularly desirable for therapeutic treatment of human patients. Such antibodies can be produced, for example, using transgenic mice which are incapable of expressing endogenous immunoglobulin heavy and light chains genes, but which can express human heavy and light chain genes. The transgenic mice are immunized in the normal fashion with a selected antigen, *e.g.*, all or a portion of a polypeptide of the invention. Monoclonal antibodies directed against the antigen can be obtained using conventional hybridoma technology. The human immunoglobulin transgenes harbored by the transgenic mice rearrange during B cell differentiation, and subsequently undergo class switching and somatic mutation. Thus, using such a technique, it is possible to produce therapeutically useful IgG, IgA and IgE antibodies. For an overview of this technology for producing human antibodies, see Lonberg and Huszar (1995, *Int. Rev. Immunol.* 13:65-93). For a detailed discussion of this technology for producing human antibodies and human monoclonal antibodies and protocols for producing such antibodies, *see, e.g.*, U.S. Patent 5,625,126; U.S. Patent 5,633,425; U.S. Patent 5,569,825; U.S. Patent 5,661,016; and U.S. Patent 5,545,806. In addition, companies such as Abgenix, Inc. (Fremont, CA), can be engaged to provide human antibodies directed against a selected antigen using technology similar to that described above.

Completely human antibodies which recognize a selected epitope can be generated using a technique referred to as "guided selection." In this approach a selected non-human monoclonal antibody, *e.g.*, a mouse antibody, is used to guide the selection of a completely human antibody recognizing the same epitope. (Jespers et al. (1994) *Bio/technology* 12:899-903).

As described herein, anti-Stat1, anti-Stat2, anti-Stat3, anti-Stat4, anti-SOCS1 or anti-SOCS3 antibodies can be used diagnostically to monitor protein levels within affected tissue (*e.g.*, a transplant cell, tissue, organ or organ system, or a cell, tissue, organ or organ system that is, or is suspected of being affected by an autoimmune disorder) as part of a clinical testing procedure, *e.g.*, to, for example, determine transplant rejection or the efficacy of a given treatment regimen. Detection can be facilitated by coupling the antibody to a detectable substance. Examples of detectable substances include various enzymes, prosthetic groups, fluorescent materials, luminescent materials, bioluminescent materials, and radioactive materials. Examples of suitable enzymes include horseradish peroxidase, alkaline phosphatase, beta-galactosidase, or acetylcholinesterase; examples of suitable prosthetic group complexes include streptavidin/biotin and avidin/biotin; examples of suitable fluorescent materials include umbelliferone, fluorescein, fluorescein isothiocyanate,

rhodamine, dichlorotriazinylamine fluorescein, dansyl chloride or phycoerythrin; an example of a luminescent material includes luminol; examples of bioluminescent materials include luciferase, luciferin, and aequorin, and examples of suitable radioactive material include ^{125}I , ^{131}I , ^{35}S or ^3H .

5 Further, as described herein, anti-Stat1 antibodies, anti-Stat2 antibodies, anti-Stat3 antibodies, anti-Stat4 antibodies, anti-SOCS1 antibodies, anti-SOCS3 antibodies, or any combination thereof can be conjugated to a therapeutic moiety and administered to a mammal, preferably a human, to reduce or prevent immune rejection. Examples of therapeutic moieties that can be conjugated to antibodies include, but are not limited to, a
10 cytotoxin, a therapeutic agent or a radioactive metal ion. A cytotoxin or cytotoxic agent includes any agent that is detrimental to cells such as taxol, cytochalasin B, gramicidin D, ethidium bromide, emetine, mitomycin, etoposide, tenoposide, vincristine, vinblastine, colchicin, doxorubicin, daunorubicin, dihydroxy anthracin dione, mitoxantrone, mithramycin, actinomycin D, 1-dehydrotestosterone, glucocorticoids, procaine, tetracaine,
15 lidocaine, propranolol, and puromycin and analogs or homologs thereof. Therapeutic agents include, but are not limited to, antimetabolites (*e.g.*, methotrexate, 6-mercaptopurine, 6-thioguanine, cytarabine, 5-fluorouracil decarbazine), alkylating agents (*e.g.*, mechlorethamine, thioepa chlorambucil, melphalan, carmustine (BSNU) and lomustine (CCNU), cyclophosphamide, busulfan, dibromomannitol, streptozotocin,
20 mitomycin C, and cis-dichlorodiamine platinum (II) (DDP) cisplatin), anthracyclines (*e.g.*, daunorubicin (formerly daunomycin) and doxorubicin), antibiotics (*e.g.*, dactinomycin (formerly actinomycin), bleomycin, mithramycin, and anthramycin (AMC)), and anti-mitotic agents (*e.g.*, vincristine and vinblastine).

The antibodies can also be conjugated a drug moiety that modifies a given
25 biological response. For example, a drug moiety may be a protein or polypeptide possessing a desired biological activity. Such proteins may include, for example, a toxin such as abrin, ricin A, pseudomonas exotoxin, or diphtheria toxin; and a lymphokine such as IL-4 or IL-13.

The invention also provides kits comprising an anti-Stat1 antibody, an anti-Stat2
30 antibody, an anti-Stat3 antibody, an anti-Stat4 antibody, an anti-SOCS1 antibody, an anti-SOCS3 antibody, or any combination thereof conjugated to a detectable substance, and instructions for use. Still another aspect of the invention is a pharmaceutical composition comprising an anti-Stat1 antibody, an anti-Stat2 antibody, an anti-Stat3 antibody, an anti-Stat4 antibody, an anti-SOCS1 antibody, an anti-SOCS3 antibody and a pharmaceutically
35 acceptable carrier. In preferred embodiments, the pharmaceutical composition contains an anti-Stat1 antibody, an anti-Stat2 antibody, an anti-Stat3 antibody, an anti-Stat4 antibody,

an anti-SOCS1 antibody, or an anti-SOCS3 antibody, a therapeutic moiety, and a pharmaceutically acceptable carrier.

In instances wherein an anti-Stat1 antibody, an anti-Stat2 antibody, an anti-Stat3 antibody, an anti-Stat4 antibody, an anti-SOCS1 antibody, an anti-SOCS3 antibody is to be
5 utilized as a therapeutic, characterization of the antibody can routinely be assayed and ascertained via the methods presented herein. For example, the fact that lymphocytes and animal models for transplants and autoimmune disorders are readily available, coupled with the availability of multiple assays for Stat and SOCS expression and activity provide for routine testing and analysis (*e.g.*, for *in vitro* and *in vivo* testing and analysis) of such
10 antibodies. The antibodies described herein can be tested, for example, for their ability to modulate the expression and/or activity of Stat1, Stat2, Stat3, Stat4, Stat6, SOCS1, SOCS3, or any combination thereof, and for their specificity and toxicity.

GENE THERAPY TECHNIQUES

15 Gene therapy refers to therapy performed by the administration to a subject of an expressed or expressible nucleic acid. Any of the methods for gene therapy available in the art can be used according to the present invention. Exemplary methods are described below.

In one embodiment, Stat4 antisense oligonucleotides are administered to reduce
20 immune rejection by way of gene therapy. In another embodiment, nucleic acid molecules comprising sequences encoding one or more anti-Stat4 antibodies are administered to reduce immune rejection, by way of gene therapy. In another embodiment, nucleic acid molecules comprising sequences encoding one or more peptides that compete with Jak2 for binding to the IL-12 receptor β_2 ("IL-12R β_2 ") are administered to immune rejection, by way
25 of gene therapy. In another embodiment, nucleic acid molecules comprising sequences encoding one or more peptides that compete with Tyk2 for binding to the IL-12R β_1 are administered to reduce immune rejection, by way of gene therapy. In another embodiment, nucleic acid molecules comprising sequences encoding one or more peptides that compete with Stat4 for binding to the IL-12R β_2 are administered to reduce immune rejection, by way
30 of gene therapy. In another embodiment, nucleic acid molecules comprising sequences encoding one or more peptides that bind to the Stat4 SH2 domain which prevent Stat4 from binding to the IL-12R β_2 are administered to reduce immune rejection, by way of gene therapy. In yet another embodiment, nucleic acid molecules comprising sequences encoding one or more dominant-negative Stat4 polypeptides (*e.g.*, a Stat4 polypeptide
35 lacking its SH2 domain or a Stat4 polypeptide lacking its DNA binding domain) are administered to reduce immune rejection, by way of gene therapy.

In specific embodiments, Stat1 antisense oligonucleotides, Stat2 antisense oligonucleotides, Stat3 antisense oligonucleotides, or the combination thereof are administered to reduce immune rejection by way of gene therapy. In other embodiments, nucleic acid molecules comprising sequences encoding one or more anti-Stat1 antibodies, anti-Stat2 antibodies, or anti-Stat3 antibodies are administered to reduce immune rejection, by way of gene therapy. In other embodiments, nucleic acid molecules comprising sequences encoding one or more peptides that compete with Jak1 or Tyk2 for binding to the IFN- α receptor or IFN- β receptor are administered to reduce immune rejection, by way of gene therapy. In yet another embodiment, nucleic acid molecules comprising sequences encoding one or more peptides that compete with Jak1 or Jak2 for binding to the IFN- γ receptor are administered to reduce immune rejection, by way of gene therapy. In other embodiments, nucleic acid molecules comprising sequences encoding one or more peptides that compete with Stat1 or Stat2 for binding to the IFN- α receptor or IFN- β receptor are administered to reduce immune rejection, by way of gene therapy. In other embodiments, nucleic acid molecules comprising sequences encoding one or more peptides that compete with Stat1 for binding to the IFN- γ receptor are administered to reduce immune rejection, by way of gene therapy. In yet other embodiments, nucleic acid molecules comprising sequences encoding one or more dominant-negative Stat1 polypeptides (e.g., a Stat1 polypeptide lacking its SH2 domain or a Stat1 polypeptide lacking its DNA binding domain), dominant-negative Stat2 polypeptides (e.g., a Stat2 polypeptide lacking its SH2 domain or a Stat2 polypeptide lacking its DNA binding domain), or dominant-negative Stat3 polypeptides (e.g., a Stat3 polypeptide lacking its SH2 domain or a Stat3 polypeptide lacking its DNA binding domain) are administered to reduce immune rejection, by way of gene therapy.

For general reviews of the methods of gene therapy, see Goldspiel et al., 1993, *Clinical Pharmacy* 12:488-505; Wu and Wu, 1991, *Biotherapy* 3:87-95; Tolstoshev, 1993, *Ann. Rev. Pharmacol. Toxicol.* 32:573-596; Mulligan, 1993, *Science* 260:926-932; and Morgan and Anderson, 1993, *Ann. Rev. Biochem.* 62:191-217; May, 1993, *TIBTECH* 11(5):155-215). Methods commonly known in the art of recombinant DNA technology which can be used are described in Ausubel et al. (eds.), 1993, *Current Protocols in Molecular Biology*, John Wiley & Sons, NY; and Kriegler, 1990, *Gene Transfer and Expression*, A Laboratory Manual, Stockton Press, NY.

In one aspect, a composition of the invention comprises nucleic acid sequences encoding one or more anti-Stat 1, anti-Stat 2, anti-Stat 3, anti-Stat 4, anti-SOCS 1, or anti-SOCS 3 antibodies or fragments thereof, said nucleic acid sequences being part of expression vectors that express one or more anti-Stat 1, anti-Stat 2, anti-Stat 3, anti-Stat 4,

anti-SOCS 1, or anti-SOCS 3 antibodies or fragments thereof in a suitable host. In particular, such nucleic acid sequences have promoters operably linked to the anti-Stat 1, anti-Stat 2, anti-Stat 3, anti-Stat 4, anti-SOCS 1, or anti-SOCS 3 antibodies or fragments thereof, said promoter being inducible or constitutive, and, optionally, tissue-specific.

5 In another aspect, a composition of the invention comprises nucleic acid sequences encoding dominant-negative Stat1, Stat2, Stat3, Stat4, SOCS1, or SOCS3 polypeptides, said nucleic acid sequences being part of expression vectors that express dominant-negative Stat1, Stat2, Stat3, Stat4, SOCS1, or SOCS3 polypeptides in a suitable host. In particular, such nucleic acid sequences have promoters operably linked to the dominant-negative Stat1,
10 Stat2, Stat3, Stat4, SOCS1, or SOCS3, said promoter being inducible or constitutive, and, optionally, tissue-specific. In another particular embodiment, nucleic acid molecules are used in which the dominant-negative Stat1, Stat2, Stat3, Stat4, SOCS1, or SOCS3 coding sequences and any other desired sequences are flanked by regions that promote homologous recombination at a desired site in the genome, thus providing for intrachromosomal
15 expression of the dominant-negative Stat1, Stat2, Stat3, Stat4, SOCS1, or SOCS3 nucleic acids (Koller and Smithies, 1989, *Proc. Natl. Acad. Sci. USA* 86:8932-8935; Zijlstra et al., 1989, *Nature* 342:435-438).

In another aspect of the invention, a composition of the invention comprises nucleic acid molecules comprising sequences encoding one or more peptides that compete with
20 Tyk2 for binding to the IL-12R β_1 , said nucleic acid sequences being part of expression vectors that express one or more peptides in a suitable host. In another aspect of the invention, a composition of the invention comprises nucleic acid sequences nucleic acid molecules comprising sequences encoding one or more peptides that compete with Stat4 for binding to the IL-12R β_2 , said nucleic acid sequences being part of expression vectors that
25 express one or more peptides in a suitable host. In yet another aspect of the invention, a composition of the invention comprises nucleic acid molecules comprising sequences encoding one or more peptides that bind to the Stat4 SH2 domain which prevent Stat4 from binding to the IL-12R β_2 , said nucleic acid sequences being part of expression vectors that
30 express one or more peptides in a suitable host. In particular a embodiment of the invention, the nucleic acid sequences encoding peptides of the invention have promoters operably linked to said nucleic acid sequences, said promoter being inducible or constitutive, and, optionally, tissue-specific.

Delivery of the nucleic acids into a patient may be either direct, in which case the patient is directly exposed to the nucleic acid or nucleic acid-carrying vectors, or indirect, in
35 which case, cells are first transformed with the nucleic acids *in vitro*, then transplanted into

the patient. These two approaches are known, respectively, as *in vivo* or *ex vivo* gene therapy.

In a specific embodiment, the nucleic acid sequence is directly administered *in vivo*, where it is expressed to produce the encoded product. This can be accomplished by any of numerous methods known in the art, *e.g.*, by constructing it as part of an appropriate nucleic acid expression vector and administering it so that they become intracellular, *e.g.*, by infection using defective or attenuated retrovirals or other viral vectors (see U.S. Patent No. 4,980,286), or by direct injection of naked DNA, or by use of microparticle bombardment (*e.g.*, a gene gun; Biolistic, Dupont), or coating with lipids or cell-surface receptors or transfecting agents, encapsulation in liposomes, microparticles, or microcapsules, or by administering them in linkage to a peptide which is known to enter the nucleus, by administering it in linkage to a ligand subject to receptor-mediated endocytosis (see, *e.g.*, Wu and Wu, 1987, *J. Biol. Chem.* 262:4429-4432) (which can be used to target cell types specifically expressing the receptors), etc. In another embodiment, nucleic acid-ligand complexes can be formed in which the ligand comprises a fusogenic viral peptide to disrupt endosomes, allowing the nucleic acid to avoid lysosomal degradation. In yet another embodiment, the nucleic acid can be targeted *in vivo* for cell specific uptake and expression, by targeting a specific receptor (see, *e.g.*, PCT Publications WO 92/06180 dated April 16, 1992 (Wu et al.); WO 92/22635 dated December 23, 1992 (Wilson et al.); WO92/20316 dated November 26, 1992 (Findeis et al.); WO93/14188 dated July 22, 1993 (Clarke et al.), WO 93/20221 dated October 14, 1993 (Young)). Alternatively, the nucleic acid can be introduced intracellularly and incorporated within host cell DNA for expression, by homologous recombination (Koller and Smithies, 1989, *Proc. Natl. Acad. Sci. USA* 86:8932-8935; Zijlstra et al., 1989, *Nature* 342:435-438).

In one embodiment, viral vectors that contain Stat4 antisense oligonucleotides are used (see Miller et al., 1993, *Meth. Enzymol.* 217:581-599). In another embodiment, viral vectors that contain nucleic acids encoding one or more anti-Stat4 antibodies are used. In another embodiment, viral vectors that contain nucleic acids encoding one or more peptides that compete with Jak2 for binding to the IL-12 receptor β_2 ("IL-12R β_2 ") are used. In another embodiment, viral vectors that contain nucleic acids encoding one or more peptides that compete with Tyk2 for binding to the IL-12R β_1 are used. In another embodiment, viral vectors that contain nucleic acids encoding one or more peptides that compete with Stat4 for binding to the IL-12R β_2 are used. In another embodiment, viral vectors that contain nucleic acids encoding one or more peptides that bind to the Stat4 SH2 domain which prevent Stat4 from binding to the IL-12R β_2 are used. In yet another embodiment, viral vectors that contain nucleic acids encoding dominant-negative Stat4 polypeptides (*e.g.*, Stat4 lacking its

SH2 domain or Stat4 lacking its DNA binding domain) are used. For example, a retroviral vector can be used. These retroviral vectors have been modified to delete retroviral sequences that are not necessary for packaging of the viral genome and integration into host cell DNA. The nucleic acid sequences encoding the Stat1, Stat2, Stat3, Stat4, SOCS1 or SOCS3 antibodies, or polypeptides or peptides of the invention to be used in gene therapy are cloned into one or more vectors, which facilitates delivery of the gene into a patient. More detail about retroviral vectors can be found in Boesen et al., 1994, *Biotherapy* 6:291-302, which describes the use of a retroviral vector to deliver the *mdr1* gene to hematopoietic stem cells in order to make the stem cells more resistant to chemotherapy. Other references illustrating the use of retroviral vectors in gene therapy are: Clowes et al., 1994, *J. Clin. Invest.* 93:644-651; Kiem et al., 1994, *Blood* 83:1467-1473; Salmons and Gunzberg, 1993, *Human Gene Therapy* 4:129-141; and Grossman and Wilson, 1993, *Curr. Opin. in Genetics and Devel.* 3:110-114.

Adenoviruses are other viral vectors that can be used in gene therapy. Adenoviruses are especially attractive vehicles for delivering genes to respiratory epithelia. Adenoviruses naturally infect respiratory epithelia where they cause a mild disease. Other targets for adenovirus-based delivery systems are liver, the central nervous system, endothelial cells, and muscle. Adenoviruses have the advantage of being capable of infecting non-dividing cells. Kozarsky and Wilson, 1993, *Current Opinion in Genetics and Development* 3:499-503 present a review of adenovirus-based gene therapy. Bout et al., 1994, *Human Gene Therapy* 5:3-10 demonstrated the use of adenovirus vectors to transfer genes to the respiratory epithelia of rhesus monkeys. Other instances of the use of adenoviruses in gene therapy can be found in Rosenfeld et al., 1991, *Science* 252:431-434; Rosenfeld et al., 1992, *Cell* 68:143-155; Mastrangeli et al., 1993, *J. Clin. Invest.* 91:225-234; PCT Publication WO94/12649; and Wang, et al., 1995, *Gene Therapy* 2:775-783. In a preferred embodiment, adenovirus vectors are used.

Adeno-associated virus (AAV) has also been proposed for use in gene therapy (Walsh et al., 1993, *Proc. Soc. Exp. Biol. Med.* 204:289-300; U.S. Patent No. 5,436,146).

Another approach to gene therapy involves transferring a gene to cells in tissue culture by such methods as electroporation, lipofection, calcium phosphate mediated transfection, or viral infection. Usually, the method of transfer includes the transfer of a selectable marker to the cells. The cells are then placed under selection to isolate those cells that have taken up and are expressing the transferred gene. Those cells are then delivered to a patient.

In this embodiment, the nucleic acid is introduced into a cell prior to administration *in vivo* of the resulting recombinant cell. Such introduction can be carried out by any

method known in the art, including but not limited to transfection, electroporation, microinjection, infection with a viral or bacteriophage vector containing the nucleic acid sequences, cell fusion, chromosome-mediated gene transfer, microcell-mediated gene transfer, spheroplast fusion, etc. Numerous techniques are known in the art for the introduction of foreign genes into cells (see, *e.g.*, Loeffler and Behr, 1993, *Meth. Enzymol.* 217:599-618; Cohen et al., 1993, *Meth. Enzymol.* 217:618-644; Cline, 1985, *Pharmac. Ther.* 29:69-92) and may be used in accordance with the present invention, provided that the necessary developmental and physiological functions of the recipient cells are not disrupted. The technique should provide for the stable transfer of the nucleic acid to the cell, so that the nucleic acid is expressible by the cell and preferably heritable and expressible by its cell progeny.

The resulting recombinant cells can be delivered to a patient by various methods known in the art. Recombinant blood cells (*e.g.*, hematopoietic stem or progenitor cells) are preferably administered intravenously. The amount of cells envisioned for use depends on the desired effect, patient state, etc., and can be determined by one skilled in the art.

Cells into which a nucleic acid can be introduced for purposes of gene therapy encompass any desired, available cell type, and include but are not limited to epithelial cells, endothelial cells, keratinocytes, fibroblasts, muscle cells, hepatocytes; blood cells such as T lymphocytes, B lymphocytes, monocytes, macrophages, neutrophils, eosinophils, megakaryocytes, granulocytes; various stem or progenitor cells, in particular hematopoietic stem or progenitor cells, *e.g.*, as obtained from bone marrow, umbilical cord blood, peripheral blood, fetal liver, etc.

In a preferred embodiment, the cell used for gene therapy is autologous to the patient.

In one embodiment in which recombinant cells are used in gene therapy, nucleic acid sequences encoding Stat1, Stat2, Stat3, Stat4, SOCS1 or SOCS3 antibodies, or polypeptides or peptides of the invention are introduced into the cells such that they are expressible by the cells or their progeny, and the recombinant cells are then administered *in vivo* for therapeutic effect. In a specific embodiment, stem or progenitor cells are used. Any stem and/or progenitor cells which can be isolated and maintained *in vitro* can potentially be used in accordance with this embodiment of the present invention (see *e.g.* PCT Publication WO 94/08598, dated April 28, 1994; Stemple and Anderson, 1992, *Cell* 71:973-985; Rheinwald, 1980, *Meth. Cell Bio.* 21A:229; and Pittelkow and Scott, 1986, *Mayo Clinic Proc.* 61:771).

Promoters that may be used to control the expression of nucleic acid sequences encoding Stat1, Stat2, Stat3, Stat4, SOCS1 or SOCS3 antibodies, or polypeptides or

peptides of the invention include, but are not limited to, the SV40 early promoter region (Bernoist and Chambon, 1981, *Nature* 290:304-310), the promoter contained in the 3' long terminal repeat of Rous sarcoma virus (Yamamoto, et al., 1980, *Cell* 22:787-797), the herpes thymidine kinase promoter (Wagner et al., 1981, *Proc. Natl. Acad. Sci. USA* 78:1441-1445), the regulatory sequences of the metallothionein gene (Brinster et al., 1982, *Nature* 296:39-42); prokaryotic expression vectors such as the β -lactamase promoter (Villa-Kamaroff et al., 1978, *Proc. Natl. Acad. Sci. USA* 75:3727-3731), or the *tac* promoter (DeBoer et al., 1983, *Proc. Natl. Acad. Sci. USA* 80:21-25); see also "Useful proteins from recombinant bacteria" in *Scientific American*, 1980, 242:74-94; plant expression vectors comprising the nopaline synthetase promoter region (Herrera-Estrella et al., *Nature* 303:209-213) or the cauliflower mosaic virus 35S RNA promoter (Gardner et al., 1981, *Nucl. Acids Res.* 9:2871), and the promoter of the photosynthetic enzyme ribulose biphosphate carboxylase (Herrera-Estrella et al., 1984, *Nature* 310:115-120); promoter elements from yeast or other fungi such as the Gal 4 promoter, the ADC (alcohol dehydrogenase) promoter, PGK (phosphoglycerol kinase) promoter, alkaline phosphatase promoter, and the following animal transcriptional control regions, which exhibit tissue specificity and have been utilized in transgenic animals: elastase I gene control region which is active in pancreatic acinar cells (Swift et al., 1984, *Cell* 38:639-646; Ornitz et al., 1986, *Cold Spring Harbor Symp. Quant. Biol.* 50:399-409; MacDonald, 1987, *Hepatology* 7:425-515); insulin gene control region which is active in pancreatic beta cells (Hanahan, 1985, *Nature* 315:115-122), immunoglobulin gene control region which is active in lymphoid cells (Grosschedl et al., 1984, *Cell* 38:647-658; Adames et al., 1985, *Nature* 318:533-538; Alexander et al., 1987, *Mol. Cell. Biol.* 7:1436-1444), mouse mammary tumor virus control region which is active in testicular, breast, lymphoid and mast cells (Leder et al., 1986, *Cell* 45:485-495), albumin gene control region which is active in liver (Pinkert et al., 1987, *Genes and Devel.* 1:268-276), alpha-fetoprotein gene control region which is active in liver (Krumlauf et al., 1985, *Mol. Cell. Biol.* 5:1639-1648; Hammer et al., 1987, *Science* 235:53-58; alpha 1-antitrypsin gene control region which is active in the liver (Kelsey et al., 1987, *Genes and Devel.* 1:161-171), beta-globin gene control region which is active in myeloid cells (Mogam et al., 1985, *Nature* 315:338-340; Kollias et al., 1986, *Cell* 46:89-94; myelin basic protein gene control region which is active in oligodendrocyte cells in the brain (Readhead et al., 1987, *Cell* 48:703-712); myosin light chain-2 gene control region which is active in skeletal muscle (Sani, 1985, *Nature* 314:283-286), and gonadotropic releasing hormone gene control region which is active in the hypothalamus (Mason et al., 1986, *Science* 234:1372-1378).

In a specific embodiment, the nucleic acid to be introduced for purposes of gene therapy comprises an inducible promoter operably linked to the coding region, such that expression of the nucleic acid is controllable by controlling the presence or absence of the appropriate inducer of transcription.

5

PHARMACEUTICAL COMPOSITIONS

The nucleic acid molecules, polypeptides, antibodies and small molecules (also referred to herein as "active compounds") described herein can be incorporated into pharmaceutical compositions suitable for administration. Such compositions typically
10 comprise the active compound and a pharmaceutically acceptable carrier. As used herein the language "pharmaceutically acceptable carrier" is intended to include any and all solvents, dispersion media, coatings, antibacterial and antifungal agents, isotonic and absorption delaying agents, and the like, compatible with pharmaceutical administration. The use of such media and agents for pharmaceutically active substances is well known in
15 the art. Except insofar as any conventional media or agent is incompatible with the active compound, use thereof in the compositions is contemplated. Supplementary active compounds can also be incorporated into the compositions.

The invention includes methods for preparing pharmaceutical compositions for modulating the expression or activity of a polypeptide or nucleic acid of interest (*e.g.*, Stat1,
20 Stat2, Stat3, Stat4, Stat6, SOS 1, or SOCS3). Such methods comprise formulating a pharmaceutically acceptable carrier with an agent which modulates expression or activity of a polypeptide or nucleic acid of interest (*e.g.*, Stat1, Stat2, Stat3, Stat4, Stat6, SOS 1, or SOCS3). Such compositions can further include additional active agents. Thus, the invention further includes methods for preparing a pharmaceutical composition by
25 formulating a pharmaceutically acceptable carrier with an agent which modulates expression or activity of a polypeptide or nucleic acid of interest and one or more additional active compounds.

A pharmaceutical composition of the invention is formulated to be compatible with its intended route of administration. Examples of routes of administration include
30 parenteral, *e.g.*, intravenous, intradermal, subcutaneous, oral (*e.g.*, inhalation), transdermal (topical), transmucosal, and rectal administration. Intravenous administration is preferred. Solutions or suspensions used for parenteral, intradermal, or subcutaneous application can include the following components: a sterile diluent such as water for injection, saline solution, fixed oils, polyethylene glycols, glycerine, propylene glycol or other synthetic
35 solvents; antibacterial agents such as benzyl alcohol or methyl parabens; antioxidants such as ascorbic acid or sodium bisulfite; chelating agents such as ethylenediaminetetraacetic

acid; buffers such as acetates, citrates or phosphates and agents for the adjustment of tonicity such as sodium chloride or dextrose. pH can be adjusted with acids or bases, such as hydrochloric acid or sodium hydroxide. The parenteral preparation can be enclosed in ampoules, disposable syringes or multiple dose vials made of glass or plastic.

- 5 Pharmaceutical compositions suitable for injectable use include sterile aqueous solutions (where water soluble) or dispersions and sterile powders for the extemporaneous preparation of sterile injectable solutions or dispersions. For intravenous administration, suitable carriers include physiological saline, bacteriostatic water, Cremophor EL™ (BASF; Parsippany, NJ) or phosphate buffered saline (PBS). In all cases, the composition must be
- 10 sterile and should be fluid to the extent that easy syringability exists. It must be stable under the conditions of manufacture and storage and must be preserved against the contaminating action of microorganisms such as bacteria and fungi. The carrier can be a solvent or dispersion medium containing, for example, water, ethanol, polyol (for example, glycerol, propylene glycol, and liquid polyethylene glycol, and the like), and suitable
- 15 mixtures thereof. The proper fluidity can be maintained, for example, by the use of a coating such as lecithin, by the maintenance of the required particle size in the case of dispersion and by the use of surfactants. Prevention of the action of microorganisms can be achieved by various antibacterial and antifungal agents, for example, parabens, chlorobutanol, phenol, ascorbic acid, thimerosal, and the like. In many cases, it will be
- 20 preferable to include isotonic agents, for example, sugars, polyalcohols such as mannitol, sorbitol, sodium chloride in the composition. Prolonged absorption of the injectable compositions can be brought about by including in the composition an agent which delays absorption, for example, aluminum monostearate and gelatin.

- Sterile injectable solutions can be prepared by incorporating the active compound
- 25 (e.g., a polypeptide or antibody) in the required amount in an appropriate solvent with one or a combination of ingredients enumerated above, as required, followed by filtered sterilization. Generally, dispersions are prepared by incorporating the active compound into a sterile vehicle which contains a basic dispersion medium and the required other ingredients from those enumerated above. In the case of sterile powders for the preparation
- 30 of sterile injectable solutions, the preferred methods of preparation are vacuum drying and freeze-drying which yields a powder of the active ingredient plus any additional desired ingredient from a previously sterile-filtered solution thereof.

- Oral compositions generally include an inert diluent or an edible carrier. They can be enclosed in gelatin capsules or compressed into tablets. For the purpose of oral
- 35 therapeutic administration, the active compound can be incorporated with excipients and used in the form of tablets, troches, or capsules. Oral compositions can also be prepared

using a fluid carrier for use as a mouthwash, wherein the compound in the fluid carrier is applied orally and swished and expectorated or swallowed.

Pharmaceutically compatible binding agents, and/or adjuvant materials can be included as part of the composition. The tablets, pills, capsules, troches and the like can
5 contain any of the following ingredients, or compounds of a similar nature: a binder such as microcrystalline cellulose, gum tragacanth or gelatin; an excipient such as starch or lactose, a disintegrating agent such as alginic acid, Primogel, or corn starch; a lubricant such as magnesium stearate or Sterotes; a glidant such as colloidal silicon dioxide; a sweetening agent such as sucrose or saccharin; or a flavoring agent such as peppermint, methyl
10 salicylate, or orange flavoring.

For administration by inhalation, the compounds are delivered in the form of an aerosol spray from a pressurized container or dispenser which contains a suitable propellant, *e.g.*, a gas such as carbon dioxide, or a nebulizer.

Systemic administration can also be by transmucosal or transdermal means. For
15 transmucosal or transdermal administration, penetrants appropriate to the barrier to be permeated are used in the formulation. Such penetrants are generally known in the art, and include, for example, for transmucosal administration, detergents, bile salts, and fusidic acid derivatives. Transmucosal administration can be accomplished through the use of nasal sprays or suppositories. For transdermal administration, the active compounds are
20 formulated into ointments, salves, gels, or creams as generally known in the art.

The compounds can also be prepared in the form of suppositories (*e.g.*, with conventional suppository bases such as cocoa butter and other glycerides) or retention enemas for rectal delivery.

In one embodiment, the active compounds are prepared with carriers that will protect
25 the compound against rapid elimination from the body, such as a controlled release formulation, including implants and microencapsulated delivery systems. Biodegradable, biocompatible polymers can be used, such as ethylene vinyl acetate, polyanhydrides, polyglycolic acid, collagen, polyorthoesters, and polylactic acid. Methods for preparation of such formulations will be apparent to those skilled in the art. The materials can also be
30 obtained commercially from Alza Corporation and Nova Pharmaceuticals, Inc. Liposomal suspensions (including liposomes targeted to infected cells with monoclonal antibodies to viral antigens) can also be used as pharmaceutically acceptable carriers. These can be prepared according to methods known to those skilled in the art, for example, as described in U.S. Patent No. 4,522,811.

35 It is especially advantageous to formulate oral or parenteral compositions in dosage unit form for ease of administration and uniformity of dosage. Dosage unit form as used

herein refers to physically discrete units suited as unitary dosages for the subject to be treated; each unit containing a predetermined quantity of active compound calculated to produce the desired therapeutic effect in association with the required pharmaceutical carrier. The specification for the dosage unit forms of the invention are dictated by and
5 directly dependent on the unique characteristics of the active compound and the particular therapeutic effect to be achieved, and the limitations inherent in the art of compounding such an active compound for the treatment of individuals.

For antibodies, the preferred dosage is 0.1 mg/kg to 100 mg/kg of body weight (more preferably, 0.1 to 20 mg/kg, 0.1-10 mg/kg, or 0.1 to 1.0 mg/kg). If the antibody is
10 to act in the brain, a dosage of 50 mg/kg to 100 mg/kg is usually appropriate. Generally, partially human antibodies and fully human antibodies have a longer half-life within the human body than other antibodies. Accordingly, lower dosages and less frequent administration is often possible. Modifications such as lipidation can be used to stabilize antibodies and to enhance uptake and tissue penetration (*e.g.*, into the brain). A method for
15 lipidation of antibodies is described by Cruikshank et al. ((1997) *J. Acquired Immune Deficiency Syndromes and Human Retrovirology* 14:193).

As defined herein, a therapeutically effective amount of protein or polypeptide (*i.e.*, an effective dosage) ranges from about 0.001 to 30 mg/kg body weight, preferably about 0.01 to 25 mg/kg body weight, more preferably about 0.1 to 20 mg/kg body weight, and
20 even more preferably about 0.1 to 1.0 mg/kg, 1 to 10 mg/kg, 2 to 9 mg/kg, 3 to 8 mg/kg, 4 to 7 mg/kg, or 5 to 6 mg/kg body weight.

The skilled artisan will appreciate that certain factors may influence the dosage required to effectively treat a subject, including but not limited to the severity of the disease or disorder, previous treatments, the general health and/or age of the subject, and other
25 diseases present. Moreover, treatment of a subject with a therapeutically effective amount of a protein, polypeptide, or antibody can include a single treatment or, preferably, can include a series of treatments.

In a representative, non-limiting example, a subject is treated at the time of transplantation, or when diagnosed as exhibiting a reoccurrence of rejection, or an
30 occurrence of a rejection (*e.g.*, an autoimmune rejection) with one to several (for example, between 3 and 7) doses of an appropriate modulator of Stat1, Stat2, Stat3, Stat4, Stat6, SOS 1, and/or SOCS3 for a maximum of one week. In a preferred embodiment of such an example, treatment would further comprise additional administration approximately once per month for about 3 to 6 months. The preferred route of administration is intravenous
35 bolus injection. It will also be appreciated that the effective dosage of the modulator used for treatment may increase or decrease over the course of a particular treatment. Changes in

dosage may result and become apparent from the results of diagnostic assays as described herein.

Preferably, administration of modulator is by intravenous injection, and can also be are or near the site of the cells or tissue to be treated, *e.g.*, administration is at or near the
5 site of the transplant or autoimmune disorder lesion.

In addition to those compounds described above, the present invention encompasses agents and use of agents which modulate expression or activity of a nucleic acid or polypeptide of interest. An agent may, for example, be a small molecule. For example, such small molecules include, but are not limited to, peptides, peptidomimetics, amino
10 acids, amino acid analogs, polynucleotides, polynucleotide analogs, nucleotides, nucleotide analogs, organic or inorganic compounds (*i.e.*, including heteroorganic and organometallic compounds) having a molecular weight less than about 10,000 grams per mole, organic or inorganic compounds having a molecular weight less than about 5,000 grams per mole, organic or inorganic compounds having a molecular weight less than about 1,000 grams per
15 mole, organic or inorganic compounds having a molecular weight less than about 500 grams per mole, and salts, esters, and other pharmaceutically acceptable forms of such compounds. It is understood that appropriate doses of small molecule agents depends upon a number of factors within the ken of the ordinarily skilled physician, veterinarian, or researcher. The dose(s) of the small molecule will vary, for example, depending upon the
20 identity, size, and condition of the subject or sample being treated, further depending upon the route by which the composition is to be administered, if applicable, and the effect which the practitioner desires the small molecule to have upon the nucleic acid or polypeptide of the invention. Exemplary doses include milligram or microgram amounts of the small molecule per kilogram of subject or sample weight (*e.g.*, about 1 microgram per kilogram to
25 about 500 milligrams per kilogram, about 100 micrograms per kilogram to about 5 milligrams per kilogram, or about 1 microgram per kilogram to about 50 micrograms per kilogram. It is furthermore understood that appropriate doses of a small molecule depend upon the potency of the small molecule with respect to the expression or activity to be modulated. Such appropriate doses may be determined using the assays described herein.
30 When one or more of these small molecules is to be administered to an animal (*e.g.*, a human) in order to modulate expression or activity of a polypeptide or nucleic acid of the invention, a physician, veterinarian, or researcher may, for example, prescribe a relatively low dose at first, subsequently increasing the dose until an appropriate response is obtained. In addition, it is understood that the specific dose level for any particular animal subject will
35 depend upon a variety of factors including the activity of the specific compound employed, the age, body weight, general health, gender, and diet of the subject, the time of

administration, the route of administration, the rate of excretion, any drug combination, and the degree of expression or activity to be modulated.

The pharmaceutical compositions can be included in a container, pack, or dispenser together with instructions for administration.

5 In one embodiment, one or more compositions for modulation of Stat1, Stat2, Stat3, Stat4, Stat6, SOCS1, or SOCS3 can be used combinatorially. For example, compositions for decreasing expression or activity of Stat4 can be utilized in combination (either simultaneously or serially) with compositions or techniques for increasing expression or activity of Stat6 can be utilized.

10 In another embodiment, one or more compositions of the present invention that modulate expression or activity of Stat1, Stat2, Stat3, Stat4, Stat6, SOCS1 or SOCS3 can be administered to a mammal, preferably a human, in combination with one or more standard immunosuppressive or immunomodulatory compounds to reduce or prevent immune rejection resulting from an autoimmune disorder or an allograft. Examples of
15 immunosuppressive agents include, but are not limited to, azathioprine, corticosteroids (e.g., prednisone), cyclosporine, OKT3 (anti-CD3 monoclonal human antibody), mycophenolate mofetil, rapamycin (sirolimus), mizoribine, deoxyspergualin, macrolide antibiotics such as, for example, FK506 (tacrolimus), brequinar, malononitriloamides (e.g., leflunamide), and anti-IL-2R antibodies (e.g., anti-Tac monoclonal antibody and BT 536).
20 See, e.g., Grummet et al., 1999, J. Am. Soc. Nephrol. 10:1366-1388; and Norman and Wadi, eds., 1998, "Primer on Transplantation," Am. Soc. Tx. Phys, 1st ed.).

Immunosuppressive agents may be administered at high doses initially and then tapered off over time to reduce or prevent immune rejection. For example, one or more compositions of the invention in combination with an initial dose of cyclosporine ranging
25 from between 5 and 10 mg/kg per day, an initial dose of 10mg/kg per day prednisone, or an initial dose of 10mg/kg per day mycophenolate mofetil may be administered to animal to reduce or prevent immune rejection. Alternatively, one or more compositions of the invention in combination with an initial dose of cyclosporine ranging from between 5 and 10 mg/kg per day, an initial dose of 10mg/kg per day prednisone, and an initial dose of
30 10mg/kg per day mycophenolate mofetil may be administered to animal to reduce or prevent immune rejection. Preferably, corticosteroids are not administered children.

In yet another embodiment, one or more compositions of the present invention that modulate expression or activity of Stat1, Stat2, Stat3, Stat4, Stat6, SOCS1 or SOCS3 can be administered to a mammal, preferably a human, in combination with one or more standard
35 autoimmune therapeutic agents. used for treating a particular autoimmune disorder. For example, one or more compositions of the present invention may be administered in

combination with one or more conventional anti-lupus therapeutic agents or drugs such as, for example, salicylates, corticosteroids, and immunosuppressants.

In another embodiment, one or more compositions of the present invention for modulating the expression or activity of Stat1, Stat2, Stat3, Stat4, Stat6, SOCS1, or SOCS3 are administered to a mammal, preferably a human, in combination with one or more T cell-targeted or B cell-targeted agents. Examples of such agents include, but are limited to, CTLA-4Ig, IL-2 antagonists (*e.g.*, anti-IL-2 receptor antibodies and IL-2 toxin conjugates), B7 monoclonal antibodies, anti-CD40L monoclonal antibodies, CD4 antagonists (*e.g.*, anti-CD4 monoclonal antibodies), CD3 antagonists (*e.g.*, anti-CD3 monoclonal antibodies), and IL-12 antagonists (*e.g.*, anti-IL-12 monoclonal antibodies and IL-12 toxin conjugates) to reduce or prevent immune rejection an autoimmune disorder or an allograft.

In order that the invention described herein may be more fully understood, the following examples are set forth. It should be understood that these examples are for illustrative purposes only and are not to be construed as limiting this invention in any manner.

6. EXAMPLE: QUANTITATIVE ANALYSIS OF NF- κ B AND I κ B PROTEINS IN MOUSE CARDIAC ALLOGRAFTS

NF- κ B proteins are transcription factors complexed with I κ B proteins in the cytoplasm but which upon cell activation are released, translocate to the nucleus and bind κ B motifs in the promoters of many genes, in particular of the promoters of genes whose expression is involved the immune response. Since NF- κ B plays an important role in the transcription of genes involved in immune responses, the expression levels of NF- κ B and I κ B proteins and their localization were determined in mouse cardiac allografts. The data generated and analyzed represents the first comprehensive analysis of NF- κ B and I κ B protein expression, phosphorylation, and localization as detected by Western blotting and immunohistology in serially harvested allografts (BALB/c \rightarrow B6), isografts and native hearts from recipients treated with IgG (rejection by day 8) or CD40 ligand monoclonal antibody ("CD40L mAb"; permanent survival).

Heterotopic cardiac allografting was performed with anastomoses to the abdominal aorta and vena cava (Hancock et al., 1998, *Nature Medicine* 4:1392-1396), using BALB/c donors and B6/129 wild-type or other Balb/c wild-type mice as recipients. Recipients were treated with hamster IgG or hamster anti-mouse (CD40L mAb; 250 μ g, administered intravenously) plus DST (5×10^6 splenic mononuclear cells) at the time of transplantation (Hancock et al., 1998, *Nature Medicine* 4:1392-1396). The protein expression, phosphorylation and localization of NF- κ B and I κ B were detected by Western blot analysis

and immunohistology using serially harvested allografts, isografts and native hearts from recipients treated with IgG (rejection by day 8) or CD40L mAb (permanent survival).

The following tissue-specific expression patterns in the basal cardiac state relative to other organs were detected: low p50; low p52; low p65; low C-rel; low relB; low IkBa; 5 high IkBb; low IkBe; low BCL-3 and high p105. The level of NF- κ B proteins was significantly increased (2-4 fold) upon allografting and these levels were only modestly affected by CD40L mAb. Significant increases in the levels of IkBa (greater than 2 fold) and IkBe (greater than 6 fold) proteins were also detected in cardiac allografts. In contrast, a significant decrease in the level of IkBb protein, low to undetectable levels of p105 10 protein, and trace levels of BCL-3 were detected in cardiac allografts, but were only modestly affected by CD40L mAb. Thus, these results demonstrate that in cardiac allografts the NF- κ B regulatory apparatus is highly activated at the protein level and is only modestly affected by CD40L mAb.

Cardiac samples principally express p105 and IkBb, but these are down-regulated 15 during rejection, presumably through the action of the proteasome. By contrast, cardiac allograft rejection is associated primarily with expression by infiltrating leukocytes of p65, p50 and c-rel NF- κ B proteins, plus IkBa and IkBc proteins.

The results, therefore, suggest that monitoring of the levels of NF- κ B and IkB proteins in biopsies from transplant recipients may be of diagnostic and/or prognostic 20 significance.

7. EXAMPLE: DIFFERENTIAL EFFECTS OF IMMUNOSUPPRESSIVE AGENTS ON ANTI-CD40L ANTIBODY-MEDIATED TOLERANCE INDUCTION

The data presented herein demonstrate that concomitant use of the 25 immunosuppressive agents cyclosporin A or methylprednisolone, but not rapamycin, blocks CD154 mAb efficacy in experimental allograft recipients. Indeed, the differential effects of these agents on CD154 mAb-induced tolerance correlates with their capacity to inhibit activation-induced CD154 expression on CD4+ T cells. Full expression of CD154 expression was found to require NF- κ B activation, and CD154 mAb was ineffective in 30 NF- κ B/p50 deficient allograft recipients or control mice in which NF- κ B activation was blocked by a proteasome inhibitor. Hence, these data indicate that strategies to use CD154 mAb clinically must take into account the effects of immunosuppressive agents on CD154 induction, which appears to be at least partially NF- κ B dependent, and suggest that ligation of surface-expressed CD154 provides an important signal that modulates T cell activation. 35

MATERIALS & METHODS

Media and Reagents:

Cell culture media, serum and supplements were purchased from Gibco BRL (Rockville, MD) and all mAbs were from PharMingen (San Diego, CA). Cyclosporin A (catalog C-3662, Sigma, St. Louis, MO) was prepared as a 5 mg/ml stock solution in 0.9% saline; rapamycin (catalog 380-004-M001, Alexis, San Diego, CA) as a 1 mg/ml stock solution in ethanol; 6 α -methylprednisolone (catalog M-0369, Sigma) as a 5 mg/ml stock solution in 80% ethanol; mycophenolate mofetil (catalog M-5255, Sigma) as a 20 mM stock solution in DMSO; and the 3 proteasome inhibitors (Grisham, M.B., et al., 1999, Methods Enzymol 300:345-63), clasto-lactacystin β -lactone (catalog 426102, Calbiochem, San Diego, CA) and its derivative PS-519 (Proscript, Cambridge, MA), and dipeptide-boronate (MG-273, ProScript), each as a 10 mM stock solution in DMSO.

Mice:

BALB/c (H-2^d) and B6/129 (H-2^b) mice were obtained from Jackson Labs (Bar Harbor, ME), and NF- κ B/p50 knockout (p50 KO) B6/129 mice (H-2^b) (Sha, W.C. et al., 1995, Cell 80:321-30.) were provided by Dr. David Baltimore (MIT, Cambridge, MA), and were housed under specific pathogen-free conditions.

Cardiac Transplantation:

Heterotopic cardiac allografting was performed with anastomoses to the abdominal aorta and vena cava (Hancock, W.W. et al., 1998, Nature Medicine 4:1392-1396), using BALB/c donors and B6/129 wild-type or NF- κ B/p50 KO mice as recipients (n=6/group). Recipients were treated with hamster IgG or hamster anti-mouse CD 154 mAb (250 μ g, i.v.) plus DST ("donor specific transfusion"; 5 x 10⁶ splenic mononuclear cells) at the time of transplantation (Hancock, W.W. et al., 1998, Nature Medicine 4:1392-1396). Additional groups of allografted wild-type mice were treated with CD 154 mAb/DST plus (i) rapamycin (0.2 mg/kg/d i.p.) or (ii) cyclosporin A (10 mg/kg/d i.p.) on day 0 and every other day until day 14; (iii) methylprednisolone (1 mg/kg i.p.) on day 0, 1 and 2; and (iv) PS-519 (1 mg/kg/d i.p.) daily from the time of transplantation. Graft survival was monitored by daily palpation, and rejection was confirmed by laparotomy and histologic evaluation.

Measurement of activation-induced CD154 expression:

Six-well plates (Costar) were pre-coated overnight with 1 μ g/ml of rat anti-mouse CD3 mAb (2C11). After washing with media (RPMI supplemented with 10% fetal bovine

serum, 50 U/ml penicillin, 50 µg/ml streptomycin, 2 mM glutamine and 50 µM 2-mercaptoethanol). one ml of media containing 2X final drug concentration was added to each well. After testing of a range of concentrations of each agent, the optimal final physiologic drug concentrations for the data to be reported were 125 ng/ml cyclosporin A, 20 ng/ml rapamycin. 100 µg/ml methylprednisolone, 1 µM mycophenolate mofetil, 10 µM lactacystin and 10 µM MG-273. Cells were obtained from mechanically disrupted spleens and, after collection by centrifugation and washing once with media, one ml aliquots containing two million viable splenocytes were added to each well. Cultures were incubated at 37°C in 5% CO₂ for 7 hr. and were then diluted two-fold with chilled media containing 0.1% sodium azide (FACS media). Cells were collected by centrifugation at 4°C, resuspended in FACS media containing 1 µg Fc Block (PharMingen) and incubated on ice for 15 min. Samples were then split and either a mixture of 1 µg each of FITC-conjugated CD4-specific mAb and PE-conjugated control hamster IgG mAb or FITC-conjugated CD4-specific mAb and PE-conjugated CD 154-specific mAb were added. After a 30 min incubation on ice, cells were washed twice and analyzed on a Becton Dickinson FACScan using Cell Quest software. Viable CD4+ lymphocytes were gated using a combination of forward/side scatter and CD4 staining.

20 Statistics

Flow cytometry data was analyzed using student t-test and cardiac allograft survival was evaluated by the two-tailed Mann-Whitney U test using the program InStat (GraphPad Software, San Diego. CA); p.<0.05 indicated a significant result.

25

RESULTS

Given the apparent efficacy of CD154 mAb in inducing long-term cardiac allograft survival in various mouse strains (Larsen, C.P. et.al., 1996, Nature 381:434-438; Hancock. W. W. et al., 1996 Proc. Natl. Acad. Sci. USA. 93:13967-13972), without development of transplant arteriosclerosis or other sequelae of chronic rejection (Hancock, W.W. et al., 1998, Nature Medicine 4:1392-1396), investigators have begun clinical testing. As human allograft recipients presently derive significant benefit from conventional immunosuppressant therapy, initial CD 154 mAb trial designs have included pharmacologic immunosuppression. However, conventional immunosuppression with glucocorticoids or cyclosporin A has prevented the success of otherwise potent experimental approaches for tolerance induction, such as that seen with intrathymic injection of donor MHC antigen in rodents (Perico, N. et al., 1995, J. Amer. Soc. Nephrol. 5:1618-1623). Likewise, CD154

mAb-induced prolongation of renal allograft survival in the initial primate studies was diminished by concomitant use of either glucocorticoids or FK506 (Kirk, A.D. et. Al, 1999, Nature Medicine 5:686-693): FK-506, like cyclosporin A, blocks activation of the calcium-dependent serine phosphatase calcineurin.

5 To understand the potential for inhibitory effects of immunosuppressive agents on the therapeutic efficacy of CD154 mAb, the effectiveness of CD154 mAb was investigated in conjunction with adjunct therapies in a completely MHC-mismatched (H2^d->N2^b) mouse cardiac allograft model. The results of concomitant administration of cyclosporin A, methylprednisolone or rapamycin on allograft survival in mice treated with CD154 mAb
10 plus DST are summarized in Figure 10. Whereas CD154 mAb therapy induced permanent cardiac allograft survival (>100 days), the effects of CD154 mAb were blocked by addition of cyclosporin A or methylprednisolone ($p < 0.001$ vs. CD154 mAb/DST alone), but not by rapamycin (Figure 10).

Given these markedly contrasting differences in effects of standard
15 immunosuppressive agents on the efficacy of CD154 mAb therapy, and recent evidence that CD154 may signal to T cells (Blair, P.J. et al., 2000, J Exp Med 191:651-660), it was hypothesized that inhibition of CD 154 expression was involved. Accordingly, the in vitro effects of standard immunosuppressive agents on the expression of CD154 by activated T cells were tested in vitro. It was found that whereas resting CD4⁺ splenic cells lacked CD
20 154 expression, 25-30% of cells expressed the molecule within 7 hours of activation with plate-bound CD3 mAb (Fig. 11A). This upregulation was markedly suppressed by therapeutic doses of cyclosporin A or methylprednisolone. but not by rapamycin or mycophenolate mofetil (Figure 11a). Analysis of three separate experiments showed >90% suppression by methylprednisolone ($p < 0.01$) and >70% suppression by cyclosporin A
25 ($p < 0.01$) whereas the effects of rapamycin (<20% inhibition) and mycophenolate mofetil (<10% inhibition) were not statistically distinguishable from control activated cells (Figure 11B). Thus, pharmacologic immunosuppressives that reduce the efficacy of CD154 mAb in vivo also inhibit activation-induced CD154 expression in vitro.

Since the allograft response is highly T cell-dependent, immunosuppressants must
30 modulate one or more aspects of the T cell response. As reviewed in this context (Gummert, J.F. et al., 1999, J. Am. Soc. Nephrol. 10:1366-1380), full T cell activation requires three signals: signal 1 is the triggering of the T cell antigen receptor, signal 2 is costimulation through CD28 and related molecules, and signal 3 is provided by cytokines. Cyclosporin A, like FK-506, blocks activation of calcineurin, an early event in T cell
35 activation, just downstream of signal 1. Calcineurin dephosphorylates the transcription factor, nuclear factor of activated T cells (NFAT), and though it is well known that NFAT is

essential to the transcription of IL-2 and (FN- γ (Ullman, K.S. et al., 1990, *Ann Rev Immunol* 8:421-52), it is less appreciated that NFATp-binding sites are also present in the CD154 promoter (Schubert, L.A., et al., 1995, *J. Biol Chem* 270:29624-7). It was found here that CD154 induction on murine CD4+ T cells was markedly inhibited by cyclosporin A, as was reported for human T cells (Fuleihan, R., et al., 1994, *J. Clin Invest* 93:1315-20).

In contrast to cyclosporin A, the macrolide rapamycin blocks a relatively late stage of T cell activation, as the target of rapamycin ("TOR"), is downstream of signal 3, cytokine receptor activation. Notably, though rapamycin is known to block various CD28-mediated events, and costimulation though CD28 can augment CD154 expression (Klaus, S.J., et al., 1994, *J. Immunol* 152:5643-5652), it was found here that rapamycin had no significant inhibitory effect on CD154 induction, suggesting that the effects of CD28 costimulation on CD154 expression are not critical. Hence, our data suggest that the contrasting efficacies of rapamycin and other immunophilin-binding agents such as cyclosporin A or FK506 in trials of CD154 mAb correlate with their capacities to block CD154 induction and may reflect inhibition of late and early T cell activation events, respectively.

The immunosuppressive agent mycophenolate mofetil blocks the final stage of T cell activation. T cell proliferation, which requires de novo synthesis of purine and pyrimidine nucleotides. Mycophenolate mofetil is converted within lymphocytes to its active metabolite, mycophenolic acid, which is a reversible inhibitor of a key enzyme in the de-novo purine synthesis pathway, inosine monophosphate dehydrogenase (Gummert, J.F. et al., 1999, *J. Am. Soc. Nephrol.* 10:1366-1380). Mycophenolate mofetil was notable in the current studies for its complete lack of effect on CD154 expression by T cells, consistent with an antagonistic function at late stages of T cell activation. Accordingly, use of CD 154 mAb with mycophenolate mofetil and/or rapamycin may be particularly efficacious in the management of transplant recipients.

Methylprednisolone resulted in almost complete inhibition of CD154 induction in mouse CD4+ T cells, consistent with a single previous report of the effects of dexamethasone on human CD4+ T cells in which activation was induced by a non-TCR-dependent mechanism (PMA/ionomycin) (Bischof, F. et al., 1998, *Cell Immunol* 187:3844). Glucocorticoids enter target cells, bind cytoplasmic receptors and form complexes which translocate to the nucleus and bind specific response elements in the promoters of target genes (Gummert, J.F. et al., 1999, *J. Am. Soc. Nephrol.* 10:1366-1380). In addition to inhibiting late events in T cell activation including proliferation and cytokine production, glucocorticoids block early events such as the activation of transcription factors AP-1 and NF- κ B. Indeed, ligand-bound glucocorticoid receptors bind RelA and NF- κ B p50 subunits

in vitro (Epinat, J.C. et al., 1999, *Oncogene* 18:6896-6909), affecting the transactivation potential of RelA/p65, and also enhance transcription of the NF- κ B antagonist I κ B- α . Thus, we considered whether NF- κ B activation was required for CD154 expression.

Compared with the responses in normal CD3 mAb-activated CD4⁺ T cells, CD154
5 induction was consistently inhibited by about 65% in NF- κ B/p50 KG cells (Figure 12) ($p < 0.02$). Since activation of NF- κ B requires the signal-coupled phosphorylation and proteolysis of I κ B- α through the 26S proteasome (Lin, Y.C., et al., 1995, *Proc Natl Acad Sci USA* 92:552-6; Traenckner, E.B. et al., 1995, *EMBO J* 14:2876-83), we also tested the effects of 2 different types of proteasome inhibitors on CD154 induction by activated T
10 cells (Fig. 12). Lactacystin irreversibly blocks proteasome activity by acylating a threonine residue in the active site of the mammalian proteasome subunit X, whereas dipeptide boronates, such as MG-273, act by irreversible inhibition of proteasomal chymotryptic activity (Grisham, M.B. et al., 1999, *Methods Enzymol* 300:345-63). Use of either agent in vitro significantly decreased CD154 expression by CD3-stimulated CD4⁺ cells ($p < 0.05$)
15 (Figure 12). Together, these data indicate that NF- κ B activation is required for optimal induction of CD154 by activated T cells.

To assess the validity of these in vitro data as guides to the in vivo efficacy of CD154 mAb therapy, CD154 mAb-induced cardiac allograft survival in normal vs. NF- κ B/p50 KO mice, as well as in wild-type mice treated with a proteasome inhibitor; all
20 experiments involved the same H2^d->H2^b MHC disparity. As anticipated from the in vitro data using spleen cells from NF- κ B/p50 KO mice was evaluated, the efficacy of CD154 mAb therapy in vivo was abrogated in NF- κ B/p50 KG mice ($p < 0.001$) (Figure 13). Moreover, administration of a proteasome inhibitor to wild-type mice also blocked the effects of CD154 mAb therapy in vivo ($p < 0.001$) (Figure 13). In this light, even agents
25 such as the calcineurin-inhibitors, cyclosporin A and FK-506, which are known to inhibit NF- κ B activation (Epinat, J.C. et al., 1999, *Oncogene* 18:6896-6909), may exert their effects on CD154 induction and CD154 mAb therapy as consequences of NF- κ B inhibition.

The first conclusion of these in vitro and in vivo studies is that CD154 mAb fails to prolong allograft survival under conditions that diminish upregulation of CD154 upon
30 CD4⁺ T cell activation. Indeed, the flow cytometric assay of CD154 expression provides a ready approach to evaluating potential clinical utility of CD154 mAb in combination with other immunosuppressive agents, and predicting those combinations which are likely to be successful (e.g. CD154 mAb plus rapamycin or mycophenolate mofetil) or not (e.g. CD154 mAb plus cyclosporin A or glucocorticoids). These studies further suggest that agents
35 blocking early stages of T cell activation, thereby suppressing CD154 induction, will not be useful adjuncts to CD154 mAb therapy.

A second conclusion from the data is that NP- κ B is required for CD154 induction, a finding that has not been previously described. Interestingly, the phenotype of NF- κ B/p50 KO is one of moderate immunodeficiency associated with defective antibody responses (Sha, W.C. et al., 1995, Cell, 80:321-30). These data indicate, therefore, that much of this phenotype may relate to defects in CD154 induction, which is key to development of B cell responses.

In addition, these findings provide an alternate interpretation for recent experimental data emphasizing an important role for CD40/CD154 interactions in the development of apoptosis in vivo, and which have suggested that concomitant use of cyclosporin A prevents tolerance induction by impairing activation-induced cell death (Li, X.C. et al., 1999, J. Immunol 163:2500-2 507; Wells. A.D. et al., 1999 Nat Med 5:1303-7). These data are more consistent with models suggesting that CD154mAb functions by stimulating CD154-dependent events (Blair, P.J. et al., 2000, J Exp Med 191:651-660), at least transiently. Further, these data demonstrate, for the first time, that NF- κ B is required for CD154 induction.

8. EXAMPLE: Post-Transplantation Stat and SOCS Levels and Their Involvement in Immune Reduction and Tolerance Induction

This Example presents the first data that has been generated on the post-transplantation levels of Stat and SOCS members post-transplantation. Using a vascularized cardiac transplant model, the levels of Stats (Stats 1-6) and SOCS (inhibitors of Stats) were measured during rejection and tolerance (via CD40L antibody treatment). During rejection, a rapid upregulation of Stat1, Stat2 and Stat3 mRNA was observed, followed by upregulation of these Stats in the animals' own hearts, indicating a systemic expression pattern. In contrast, RNA and protein analysis demonstrate that changes in Stat4 and Stat6 expression are confined to the transplant tissue. Specifically, Stat4 mRNA, however, was upregulated only later, and only in the transplant heart. Likewise, Stat6 mRNA was upregulated locally in the transplant tissue. Treatment with CD40L monoclonal antibody (MR-1) resulted in the downregulation of all the Stats, with the exception of Stat6, which was upregulated upon MR-1 treatment. Thus, reduction of immune rejection, via induction of tolerance, was accompanied by a downregulation of Stat4 and an increase in Stat6 levels. Cardiac transplants in Stat6 knockout mice were rejected despite treatment with MR-1. These results indicate that an immune deviation from TH1 (mediated by IL-12 and Stat4) to TH2 (mediated by IL-4 and Stat6) is a prerequisite in the induction of tolerance, and that Stat4 and Stat6, at a minimum, are critical signals in graft survival.

Further, these results indicate that Stat6, at a minimum, is a prerequisite in the induction of tolerance.

MATERIALS & METHODS

5 Cloning by RT-PCR:

5 μ g of IL-6 (Pharmingen, San Diego, CA) was injected intravenously to a B6/129 strain female mouse. The heart, liver and spleen from the mouse was collected 1 hr after the intravenous injection of IL-6. Total RNA from these three organs were prepared using the acid-guanidine thiocyanate-phenol-chloroform method (Promega, Madison, WI). The RNA
10 was then dissolved in water, quantitated, and a portion of the RNA was combined in equal amounts, and 1 μ g of the combined RNA was used in a 50 μ l reaction volume for the synthesis of first-strand cDNA. The ProStar Ultra HF RT-PCR System (Stratagene, La Jolla, CA) reagents were used both for the first-strand cDNA generation and for the subsequent steps during the amplification of the cDNA template. The following upstream
15 and downstream primers were used:

```

Stat1:  5'-GAACTTTCAGCTGTTACTTTCC-3' (SEQ ID NO:35)
        5'-CTGTGCTCATCATACTGTC-3' (SEQ ID NO:36)
Stat2:  5'-GTGTTACAGTCACTCCCACTG-3' (SEQ ID NO:37)
        5'-CCTCAGGCAAATCTGACTCTG-3' (SEQ ID NO:38)
20 Stat3:  5'-GAAAGTACTGTAGGCCCGAG-3' (SEQ ID NO:39)
        5'-CTGGAACCACAAAGTTAGGAG-3' (SEQ ID NO:40)
Stat4:  5'-GAAGTGAGATTCCACTCTGTAG-3' (SEQ ID NO:41)
        5'-CACTCTCCAGTTTCATCTGC-3' (SEQ ID NO:42)
Stat5A: 5'-CGAAAGCAGTTGACGGATACG-3' (SEQ ID NO:43)
25      5'-CTCCAACCTTAGTTGCCTAAACC-3' (SEQ ID NO:44)
Stat5B: 5'-CAAGCCGTTAGAAGCAGGAG-3' (SEQ ID NO:45)
        5'-CCATGGTTCACAACCTACAG-3' (SEQ ID NO:46)
Stat6:  5'-GATGAGGCTTTCCGGAGTCAC-3' (SEQ ID NO:47)
        5'-CAGTTGTATCACATTCGAGC-3' (SEQ ID NO:48)
30 SOCS1: 5'-CTGTGCCGCAGCATTAAGTG-3' (SEQ ID NO:49)
        5'-GTTTATTACCTAAACTGGCTG-3' (SEQ ID NO:50)
      SOCS2: 5'-CCAGGTATAAGTATTTCTCTC-3' (SEQ ID NO:51)
        5'-GGCCATTTGATCTTGAGCAGC-3' (SEQ ID NO:52)
      SOCS3: 5'-GCAGATTGGCTTCTTCCTCAG-3' (SEQ ID NO:53)
        5'-GGCATTTAAGGCGAGTCTCC-3' (SEQ ID NO:54)
35 SOCS5:  5'-GGAGCTTACTCGCAGTAGGCTC-3' (SEQ ID NO:55)
        5'-GTAGGAGTCTCTCCGTGCAAGC-3' (SEQ ID NO:56)
  
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CIS: 5' -CCAACTCTGACTGAGCCAGG-3' (SEQ ID NO:57)
 5' -CATCCATACGCAGGTGGATG-3' (SEQ ID NO:58)

Amplification reactions included 5 µl 10x Ultra HF PCR buffer, 1 µl dNTP (40 mM) mix, 0.5 µl upstream primer (approximately 500 ng/µl), 0.5 µl downstream primer (~500 ng/µl), 1 µl first-strand cDNA reaction, 41 µl H₂O, 1 µl Pfu Turbo DNA polymerase (2.5 U/µl). The same PCR program was used for all the amplifications: samples were heated for 1 min at 95°C, followed by 40 cycles of 1 min at 95°C, 1 min at 58°C, 2 min at 68°C, and final extension at 68°C for 5 min. The PCR samples were then loaded onto agarose or acrylamide gels, the cDNA fragments were isolated and cloned directly into SfrI cut PCR-Script (Stratagene). Sequence analysis of the plasmids were performed by Tufts Core Facility (Boston, MA). The length of the cloned fragments were as follows: Stat1 (334 bp); Stat2 (694 bp); Stat3 (373 bp); Stat4 (442 bp); stat 5A (634 bp); Stat 5B (458 bp); Stat6 (894 bp); SOCS1 (381 bp); SOCS 2 (266 bp); SOCS3 (381 bp); SOCS 5 (558 bp); and CIS (688 bp).

Cardiac Transplantation:

Heterotopic cardiac allografting was performed with anastomoses to the abdominal aorta and vena cava, using BALB/c donors and B6/129 wild-type or Stat-6 KO mice as recipients (n = 6/group). See, Hancock et al., 1998, Nature Med. 4:1392-1396. Recipients were intravenously administered donor specific transfusion ("DST"; 5 x 10⁶ splenic mononuclear cells) ("DST + IgG") plus 250 µg hamster anti-mouse CD40L mAb (CD154 mAb ; BioExpress, West Lebanon, NH) and DST ("DST + MR-1") or control hamster IgG ("DST + IgG") at the time of transplantation. Graft survival was monitored by daily palpation and rejection was confirmed by laparotomy and histologic evaluation.

RNA isolations and Northern blot analysis:

Total RNA from native or transplanted hearts of Balb/c or B6/129 mice was prepared using the acid-guanidine thiocyanate-phenol-chloroform method (Chomezynski, P. and Sacchi, N., 1987, Anal. Biochem. 162:156-159). 25 µg of RNA was loaded onto each lane of 1.2 % agarose-formaldehyde gels. The 0.24 kB RNA ladder (GIBCO-BRL, Rockville, MD) was used as a size control. After electrophoresis the RNA was blotted overnight onto Nytran Supercharge membranes (Schleicher & Schuell, Keene, NH) with 20 x SSC and cross-linked onto the membranes by irradiation with UV light using a Stratalinker (Stratagene). ³²P-labeled probes were prepared by using the Multiprime DNA labelling system and ³²P-dCTP (both from Amersham Pharmacia Biotech, Piscataway, NJ). Hybridizations with the ³²P-labeled probes were done at 68°C in roller bottles using

ExpressHyb Solution (Clontech Laboratories, Palo Alto, CA). For re-use, the membranes were deprobed in 0.5 % SDS at 95-100°C and exposed to film to assure complete removal of previous hybridization signals.

The nucleotide sequences of the probes utilized are presented below:

- 5 Stat1 (SEQ ID NO:59):
gaactttcagctgttactttccagatattattcgcaactacaaagtcattggctgccgagaacataccagagaatcccctgaa
gtatctgtaccccaatattgacaaagaccacgcctttgggaagtattattccagaccaaaggaagcaccagaaccgatgga
gcttgacgacctaaagcgaactggatacatcaagactgagttgatttctgtgtctgaagtcacccttctagacttcagacca
cagacaacctgtctcccatgtctccagaggagttgatgagatgtcccgatagtgggccccgaatttgacagtatgatga
gcacag
- 10 Stat2 (SEQ ID NO:60):
gtgttacagtcactcccactgacagagatcatccgccactaccaggttcttggcgaagagaacatccccgagaaccact
ccgcttccctctatccccgaatccctcgggacgaagcttttgggtgttactaccaggaaaaagttaatttgaagaacaggag
gaatatttgaacataaactcattgtgatcttaacagacaggtggacgagctgcagcagcctctggagctcaaacaggat
tcagagtccttagaagtgaatgcagagctcttgttagcacacgaccaggagttgccattgatgatgcagactgggctggtt
ctgggcacagagctgaaagtggacccatactgagtacagccccacaagtcctgctggagccagccccacaagtcctg
ctggagccagccccacaagtcctgctggagccagccccacaagtcctgctggagccagccccacaagtcctgctggag
ccagccccacaagtcctgctggagccagccccacaagtcctgctggagccagccccacaagtcctgctggagccagc
15 ccacaagtcctgctggagctagccccacaagtcctgctggagccagccccacaagtcctgctggagctagccccaca
gtccagctggagccagcacacttgctgcagcagccatcagagtcagattgcctgagg
- 20 Stat3 (SEQ ID NO:61):
gaaagtactgtaggcccgagagccaggagcacccccgaagccgaccaggtagtgtctgccccgtactgaagaccaag
ttcatctgtgtgacaccaacgacctgcagcaataaccattgacctgcccgtatgtccccccgaccttagattcattgatgcagttt
ggaaataacgggtgaaggtgctgagccctcagcaggaggggcagtttgagtcgctcacgtttgacatggatctgacctcgga
gtgtgctacctccccatgtgaggagctgaaaccagaagctgcagagacgtgacttgagacacctgccccgtgtccac
ccctaagcagccgaacccatactgtctgaaactcctaactttgtggtccag
- 25 Stat4 (SEQ ID NO:62):
gaagttagattccactctgtagaacctacaacaaagggagactgtcggtctggccttcgctgacatcctgcgagactac
aaggttatcatggctgaaaacatccctgaaaacctctgaagtacctctaccctgacattcccaagacaaagcctttggca
aacactacagctcccagccgtgcgaagtctcaagaccaaccgaacggggagacaagggttacgtccccctctgttttctc
cccatttcaacaatccgaagcgattccacggagccacaatctccttcagaccttctccccatgtctccaagtgcataatgctgt
gctgagagaaaacctgagcccaacgacaattgaaactgcaattgcaattccccatattctgctgaatgacgggtgcaaacgga
cactttaagaaggaagcagatgaaactggagagt
- 30 Stat 5A (SEQ ID NO:63):
cgaaagcagttgacggatacgtgaagccacagatcaagcaagtggccttgagttcgtcaatgcacccacagatgccgg
agccagcgcacctacatggaccaggctccttccccagtcgtgtgccctcaacctcactacaacatgtaccacccaacc
ctgacctgtccttgaccaagatggcgagtttgacctggatgagagcatggatgttgcaggcagctggaagaacttttac
gccggcccatggacagtctgcagcccgcctctccccacctgctggctctctcacctccgctagaagctccctgtcctgaa
cgctggactccatgtttcttggaaaccaccttcagtgtgaaggagcccacgtcagttgtagtatctctgttcataccaacaat
ggctttgcaggtttcacagggtaccttggccacacagttctgggttgggtaaaagcggtgggtgacctttttgttcagacct
caaggggccccagggcctctctgtgaagagctgaacctatcattgctgacaaacctatttctccgggtgctcttttctgtcca
atggccatttcagtgaatttctagaaaaggcaggaggaggtttaggcaactaagttggag
- 35 Stat 5B (SEQ ID NO:64):
caagccaagccgttagaagcaggagccccctggccagtgccctggctcacggagctgagctgtgttagatgtgttgctgt
gcgtggtgaaaggaagaccgctctccagaaaagcaatttaggcacaaagggattccgttgatggcagagtcaccagtgcata
gaaaggtagcgaaggtggacagcttacagtcacacttctcgtctgtaaatgtcctcgtaacgacattgattctctacctg

gataaccttttgtttgtttgtttgtttgtttgttttccccgtgaaccattttttctgacaagaaaacattttaattttctaag
caagaagcatttttcaataccatgtctgtgacccaaagtaaaatggatgataattcatgtaaatgtgtgcaacatagcaac
ctgaacctgcacgcgattcgggctctgtaggtgtgaacctgg

Stat6 (SEQ ID NO:65):

5 gatgaggctttccggagtcactataagcccgaacagatggggaaggacgggaggggttatgtctctactactatcaagat
gactgtggaaaggaccagcccccttactccagagccccagatgctgcatggtgccaccttatgatcttggatgg
ccccgatgcttccatgcaactcagctcagatatggggtatcctccacagtccatccacitcattcagagcctagaagagtc
catgagtgtactgccatctttcaggagcctcacctgcaaatgcccccaacatgagccagataacctatgccccttgaccag
cctcacccccagggtctgtgcagtgcagtcacaggaacatgctgtgtccagccctgaacctatgcttggcagatgtg
actatggtagaggacagttgcctaactcagcctgtgggaggtttccccaaggcacctgggtcagtgagacatgtacc
tccccgtctgctccctactgaacaggacctcaccaagcttctctggagaaccaaggggagggagggaggtccttagga
agccagccccctctgaaacctctccttatgggcaatcagggtatctcactgtcccacctggacctaaggaccaacccag
10 ctgggtatcccagctggagaagcccagaaacaaagcctctctgtctctatggaccagctctggacacctgctcatgcagg
tgccttcogtctcaactgttcttgggtaagagaaaagaactggctgggagaccatgtggtgtatggaactgctgtgctctgt
cctacctgccatacagggccccctttccagcactgggtgcaagggatgagtggggtgttaatgctcgaatgtgatac
aactg

SOCS1 (SEQ ID NO:66):

15 ctgtgccgcagcattaagtgggggcgccttattatttcttattattaattattatttttctggaaccacgtgggagccctccc
cgcttgggtcggaggagtggtgtggagggtgagatgcctcccacttctggctggagacctatcccacctctcaggg
gtgggggtgctccccctctgtgtcctccgggtccccctgtgtgtagcagcttgtgtctggggccaggacctgaattc
cactctacctctccatgtttacatattccagtatctttgcacaaaccaggggtcggggaggggtctctggcttcattttctgc
tgtgcagaatatcctattttatattttacagccagtttaggtaataaac

SOCS2 (SEQ ID NO:67):

20 ccagggtataagtatttctctctctttttcgttttttaaaaaaaaaaacacatgcctcatatagactatctccgaatgcagct
atgtgaaagagaacccagaggccctcctctggataactgcgcagaattctctcttaaggacagttgggtcagcttaactta
aagggtgtgaagatgtagctaggtattttaaagttcccccttaggtagtttagctgaatgatgctttcttctatggctgctcaag
atcaaatggcc

SOCS3 (SEQ ID NO:68):

25 gcagattggcttcttctcagggccctccactcccgagagtagagctggcaggacctggaattcgtctgaggggagggg
gagctgccacctgctttccccctccccagctccagcttcttcaagtggagccagccggcctggctgtgggacaata
cctttgacaagcggactctccccctcccttctccacccccctctgcttccaaggaggtggggacacctccaagtgtt
gaactagaactgcaagggggaatcttcaaaccttcccgtggaactgtttgctgttattgatttgggttgaatgaagagcaggca
cctggggggaaggatggaagagaaaagggtgtgtgaagggttttatgtctggccaaagaaataaccactcccactgccc
acctaggtgaggagtggtggtcctggctctggggagagtggaagggtgacctgaagagagctatactggtgccag
gtcctctccatggggcagctaataaacctcgcagatcccttgacccccagaacctccccgttgtgaagaggcagtag
catttagaaggagagacagatgaggctggtgagctggccgctttccaacaccgaaggggagggcagatcaacagatgag
ccatcttgagaccaggtttccccctggagcagatggagggttctgcttctctctctatgtggggctaggagactgcctt
30 aatgcc

SOCS 5 (SEQ ID NO:69):

35 ggagcttactgcagtaggctctcgtcttctaatcaatggataaagtggggaaaatgtggaacaacttaaaatacagatgc
cagaatctctcagccacgaggaggaagccgtaatgagaacgtggagatgaacccaacagatgtccgtctgtcaag
agaaaagcatcagctctgggagaggcagctccccagcaagagagcagctcccttaagagaaaatgttgccttacagctggg
actgagcccttcaagacctttccaggcggaaacaaaactgtgccgcagagatccctcaagtgttgaatcagcatcg
agaaagacagtgactcgggtgccacccaggaacgaggcttgcacggagagactcctac

CIS (SEQ ID NO:70):

ccaactctgactgagccaggcaccctgctctgctcacacagtcacatcctggagggaacacagtccccagctggacttg
 gggttctgctgtcctttctcagtcacatcctggtgctgcatgtgacagctggaccagagaatgccagcaagaacaag
 gcaggtggaggagggtgtcacacaactctgaggtcaacgcctctaggtacaatatggctcttltgtgtgagccatgtat
 cagagcgagacaggcaggacctcgtctctccacagaggctggacctaggtctccactcacttgcctgcccttgccacctg
 aactgtgtctattctcccagccctggtttctcagtcgtgagtagggcaggccccctacccatgtatagaatagcgagcct
 5 gtttctgggagaatatcagccagaggtgatcatgccaaggccccttatggggacgcagactgggctaggggactacac
 agttatacagtattttatttatttctccttgagggggtgggggtggaatgatggcgtgagccatcccacttctctgccctgt
 gctctgggtggtccagagacccccaggctggttcttccctgtggagacccccatcccaaacattgttgggcccacaaagta
 gtctcgaatgtcctgggcccacccacctgcgtatggatg

10 Western blotting:

Hearts obtained from mice were frozen in liquid nitrogen and homogenized with
 Tissue Tearor (model 985370, Biospec Products, Inc.) in 800 µl lysis buffer containing 4%
 SDS, 125 mM Tris-HCl, pH 6.8, and protease inhibitors Antipain, Benzamidin, BeStatin,
 Chymostatin, Leupeptin, Pefabloc C, Pepstatin A, PMSF, TLCK, TPCCK. The protease
 15 inhibitors were prepared and used as suggested by the manufacturer, Roche Molecular
 Biochemicals, Indianapolis, IN. Following homogenization, the DNA in the samples was
 sheared by sonication for 30 sec at 5 Watts (RMS) output power, with a Virtis sonicator
 (model Virsonic 60, Virtis Company, Gardiner, NY). Samples were then heated at 95°C for
 10 minutes and centrifuged at 14,000 x g for 30 minutes, to get rid of particulate material.
 20 Protein concentrations in the extracts were determined using DC Protein Assay (Bio-Rad
 Laboratories, Hercules, CA). The protein concentration in all the samples was equalized to
 10 mg/ml by adding lysis buffer, and 1 vol. 2 x Sample buffer (20 % Glycerol, 0.005 %
 Bromophenol blue) was added to yield samples with a protein concentration of 5 mg/ml.
 The samples were kept at -80°C until use. Prior to loading onto 10% SDS-polyacrylamide
 25 gels, 2.5 % β-mercaptoethanol was added to every sample (100 µg) followed by heating to
 95°C for 10 min. After electrophoresis, proteins were transferred onto Immobilon-P
 membranes (Millipore Corporation, Bedford, MA 01730) for 2.5 hrs at 100 V, using a
 Hoefer Transphor Electrophoresis unit. The Transfer buffer consisted of 12 mM Tris base,
 96 mM Glycine, and 200 ml Methanol in 1 liter. After the transfer, the membranes were
 30 placed in roller bottles and rotated for 1 hr (at 4°C) in a Blocking solution (1 x PBS + 0.1 %
 Tween-20 + 5 % non-fat dry milk + 0.02 % sodium azide), the membranes were rinsed once
 with the Rinse solution (1 x PBS + 0.1 % Tween-20) and rolled overnight with the primary
 antibody in the Blocking solution. After washing times in the Rinse solution (15, 10, 5
 minutes), the blots were incubated for 1 hour with the secondary antibody conjugated to
 35 horseradish peroxidase, followed by another set of washes with the Rinse solution and one
 final wash in 1 x TBS (10 mM Tris-HCl, pH 8.0, 150 mM NaCl). Chemiluminescence was
 performed by using the Luminol Reagent (Santa Cruz Biotechnology, Inc., Santa Cruz, CA)

and detection was with Biomax MR film (Eastman Kodak Company, Rochester, NY). All the antibodies were used at 1:1000 dilution. Stat1, phospho-Stat1, Stat3, phospho-Stat3, Stat4, and Stat5A were from UpState Biotechnology (Lake Placid, NY). Stat2, Stat6, HSC70, and the secondary antibodies were from Santa Cruz Biotechnology.

5

Quantification of RNA and Protein

Quantification of the RNA and protein bands on the Kodak Biomax MR film was performed on a Macintosh Performa 6300CD computer using the public domain NIH Image program (developed at the U.S. National Institutes of Health and available on the Internet at

10 <http://rsb.info.nih-image/>.

RESULTS

STAT and SOCS RNA levels in heart allografts during the 5 days following transplantation

15 Hearts from Balb/c mice were transplanted into B6/129 female mice using a vascularized mouse cardiac allograft model. The heart transplant resulted with animals having two hearts: their own hearts and a second functional heart grafted adjacent to the kidneys. To determine the kinetics of Stat and SOCS RNA expression following cardiac allograft surgery, total RNA was prepared from the animals' own hearts (native), the
20 transplanted hearts and from the hearts of healthy controls (designated as "N", "T", and "C", respectively) one, two, three, and five days post-transplantation. RNA (25 µg/lane) was then electrophoresed on three identical gels, transferred to Nytran Supercharge membranes and the membranes were hybridized with Stat probes, SOCS probes, and a GAPDH probe as a control (Figures 14-16).

25 As shown in Figures 14A and 14B, increases of approximately 5-fold in Stat1, Stat2 and Stat3 RNA levels were detected in the transplanted hearts relative to the control hearts as early as one day post-transplantation, and were more evident by the second day. During the first two days, the increase appeared to be localized to only the transplanted tissue. By the third day, and much more strongly by the fifth day, higher levels of Stat1, 2, and 3 RNA
30 were also detected in the native hearts.

Stat4 RNA levels, almost undetectable the first three days post-transplantation, increased significantly (approximately 60-fold) in the transplanted hearts at around day five post-transplantation, but not earlier. Stat5A RNA levels did not change in the native and transplanted hearts during the five days following the transplant surgery. An increase in
35 Stat6 RNA expression was detected in transplanted hearts as early as day one post-transplantation (approximately 2-fold), and remained at approximately the same level

during the five days following transplantation. The fact that increases in Stat4 and Stat6 RNA expression were only detected in the transplanted hearts by day five post-transplantation suggests that the up-regulation of the expression of these two Stats was localized to the transplanted hearts (Figures 14A, 14B).

5 An increase in SOCS1 RNA expression levels in the transplanted hearts began to increase by day two post-transplantation and by day five post-transplantation SOCS1 RNA expression in the transplanted hearts had increased approximately 300-fold (Figure 15). Between days 1 and 3, there was an approximately 6-fold difference in such SOCS1 RNA levels, and between days 3 and 5, there was an approximately 25-fold difference in such
10 SOCS1 RNA levels. At approximately 38-fold, the SOCS1 RNA increase in native hearts was much less pronounced (Figure 15).

A considerably high level of SOCS3 RNA expression was detected in the transplanted hearts as early as one day post-transplantation (approximately 100-fold increase), and reached an approximately 230-fold increase at five days post-transplantation.
15 (Figure 15). However, SOCS3 RNA expression remained low in native hearts, and was hardly detectable in the native hearts five days following the transplant surgery. As shown in Figure 16, Stat4 and SOCS3 RNA expression patterns were found to be quite similar to each other, both of them being expressed only in the transplanted hearts, and both of them being significantly up-regulated at approximately day 5 post-transplantation..

20 Two species of SOCS5 RNA were detected (4.4 kb and 3.8 kb) in the naive and transplanted hearts. The shorter species 3.8 kb SOCS5 RNA species was much more abundant in the transplanted hearts than in naive hearts or in control hearts. The level of CIS RNA expression detected in naive hearts remained almost unchanged relative to the control hearts, while a lower level of CIS RNA expression was detected in the transplanted
25 hearts relative to the naive hearts or control hearts (Figure 15).

STAT Protein Levels in Heart Allografts and Native Hearts During the 5 Days Post-transplantation

To determine the effect of cardiac allografts on Stat and SOCS protein expression,
30 protein extracts were prepared from transplanted and native hearts at day one, day two, day three, and day five, post-transplantation. In the transplanted hearts an increase in Stat1 protein levels relative to the control hearts was detected as early as one day post-transplantation, increased steadily, and by day five post-transplantation Stat1 protein levels were approximately 17-fold higher (Figure 17). In the native hearts an increase in Stat1
35 protein levels began increasing at day two post-transplantation and by day five post-transplantation Stat1 levels were 15-fold higher.

An increase in Stat2 protein expression levels in the transplanted hearts was initially detected three days post-transplantation, and by day five post-transplantation had increased sharply by approximately 27-fold. In native hearts an increase in Stat2 protein levels was detected by day three post-transplantation and by day five post-transplantation a 10-fold increase in the levels of Stat2 protein was detected.

An increase in Stat3 protein levels in the transplanted hearts was detected as early as day two post-transplantation and by day five post-transplantation the levels of Stat3 protein in the transplanted hearts exhibited an approximately 3.5-fold increase. In the native hearts a 2-fold increase in the levels of Stat3 protein was detectable only by day five post-transplantation.

Stat4 levels began increasing around one day post-transplantation in the transplanted heart, with a dramatic increase (approximately 16-fold) between day 2 and day 5 post-transplantation. Stat4 was detected only in the transplant tissue, not in the native heart tissue (Figure 17).

Stat5A protein levels did not change in the transplanted or native hearts relative to the control hearts.

Stat6 is normally made at a low level in hearts (see, e.g., Figure 17, control lanes). As was observed with Stat4, Stat6 expression, however, only increased (approximately 1.5-3.5-fold) in transplanted hearts.

20

Phosphorylated forms of Stat1 and Stat3 during graft rejection

To determine whether the levels of phosphorylated Stat1 and phosphorylated Stat3 protein parallel that of the newly synthesized Stat1 and Stat3 in the native and transplanted hearts following cardiac allograft surgery, the levels of these Stats and their phosphorylated forms were analyzed (Figure 18). As shown in Figure 18, an increase in phosphorylated Stat1 protein levels paralleled the increase in the total Stat1 protein levels detected both in native and transplanted hearts on days three and five post-transplantation. In contrast, the level of phosphorylated Stat3 protein detected in native and transplanted hearts over the five day post-transplantation period did not parallel the increase in the total Stat3 protein levels detected during this period. In particular, although there was a steady increase of total Stat3 over the five day period, there was no major change in the levels of phosphorylated Stat3 protein detected over this same period, with the exception of the appearance of a slightly higher molecular weight form of phosphorylated Stat3 protein.

35 The Effect of Anti-CD4OL Monoclonal Antibody MR-1 on the Stat and SOCS RNA Levels in 5-day Post-Transplant Hearts

To determine how Stat and SOCS RNA expression levels are affected in cardiac allografts when tolerance has been induced, six mice having undergone cardiac transplant surgery were divided into three groups. One group of mice were intravenously administered DST ("donor specific transfusion") + MR-1 anti-CD40L antibody which has
5 been shown to induce tolerance), on the day of the transplant surgery. The second group of mice were intravenously administered DST + IgG as control on the day of the transplant surgery and the third group received no treatment. The animals were sacrificed five days post-transplantation and total RNA was prepared from the animals' own hearts (native), the transplanted hearts and from the hearts of healthy controls (designated as "N", "T", and
10 "C", respectively). The RNA (25 µg /lane) was electrophoresed on four identical gels and the Northern blots were hybridized to Stat probes, SOCS probes and a GAPDH probe as a control. (Figures 19, 20)

Stat1, Stat2, and Stat3 mRNA were found to be expressed at some level in the normal heart (see control lanes of the no treatment group). Five days post-transplantation,
15 the transplant allograft tissue exhibited a dramatic upregulation of Stat1, Stat2 (approximately 45-fold), and Stat3 (approximately 6-fold) mRNAs. This upregulation was found to be systemic in nature in that the increases were also observed in the native heart tissue (approximately 43-fold, 48-fold, and 5.5-fold, respectively). Treatment with DST + IgG caused minimal changes in the transplant Stat RNA levels, but the single dose of DST +
20 MR-1 resulted in lower levels of Stat1, Stat2, and Stat3 mRNAs (62%, 76%, and 50% lower than the animals that received no treatment).

Stat4 mRNA was found only in the transplanted heart tissue, as a local signal, and was not detected in the control or native hearts (Figure 19; see the control and native lanes of the no treatment group). Stat4 mRNA was significantly reduced (approximately 6-fold)
25 in transplants obtained from MR-1-treated animals.

Only minimal differences in Stat 5A mRNA expression in transplant tissue of the group that received no treatment and the controls (approximately 1.4-fold increase) were observed. MR-1 treatment, however, lowered the Stat 5 A expression level by 70%.. MR-1 treatment resulted in Stat 5A levels that were 60% lower than control heart levels. Stat 5B
30 mRNA levels were too low to be detected (data not shown).

Stat6 mRNA is normally expressed at some level in the heart, and five days post-transplantation, Stat6 mRNA levels doubled only in the transplanted, not native, hearts. Upon MR-1 treatment, however, Stat6 mRNA levels were downregulated to pre-transplantation (control) levels.

35 The same membranes were used to determine the mRNA expression levels of SOCS1, SOCS 2, SOC , and CIS (Figure 20). SOCS1 and SOCS3 mRNA levels were

almost undetectable in control hearts. SOCS1 and SOCS3 mRNA levels were increased five days post-transplantation in transplant tissue in both DST + IgG treated mice and untreated mice (some low level of SOCS1 expression was also detected in the native hearts of the untreated groups). Treatment with DST + MR-1 resulted in a 93% and 73%
 5 reduction in the level of SOCS1 and SOCS3 mRNA expression, respectively, relative to that in transplanted hearts of untreated mice.

SOCS 2 RNA levels remained unchanged in the control, native, and transplanted hearts in the three experimental groups. Interestingly, CIS levels were 50% higher in the transplanted hearts of mice, regardless of treatment.

10

The Effect of Anti-CD40L Monoclonal Antibody MR-1 on Stat Protein Levels in 5-day and 7-day Post-Transplant Hearts

To determine how Stat protein levels are affected in cardiac allografts when tolerance has been induced, the day of the surgery mice were intravenously administered a
 15 single dose of either DST + IgG or DST + MR-1, and the Stat protein levels were analyzed five and seven days post-transplantation. As shown in Figure 21, at day 5 post-transplantation there were minimal changes in the Stat levels between IgG and MR-1 treated animals, but at day 7 post-transplantation, there was significant changes in such levels.

Stat1 and Stat2 levels were reduced 64% and 52%, respectively, in native hearts of
 20 MR-1 treated animals. Stat3 levels were reduced in the transplant tissue by 29%. The most dramatic change at day 7 post-transplantation was the amount of Stat4 reduction (approximately 63%) in the transplanted hearts. Stat6, on the other hand, tended to increase about 2-fold upon MR-1 treatment.

Stat and Bax Protein Levels in Isografts and Allografts

The level of Stat1, Stat2, Stat3, Stat4, Stat5A, and Stat6 were compared in isografts and allografts to exclude the possibility that the observed changes in Stats were due to the surgery and the healing process that follows the surgery. As shown in Figure 22, between
 25 day 1 and day 2 post-transplantation, the isografts showed a small increase in different Stats. The low levels, however, of Stat1, Stat2, and Stat5A remained constant between days
 30 2 and 7 post-transplantation, and Stat3 and Stat6 levels declined between these days. In allografts, on the other hand, all of the Stats reached high levels by day 5. Bax, an unrelated protein, showed a very different profile, with its levels increasing in both isografts and allografts.

35

Transplantation in Stat6 Knockout Mice

Stat6 is activated by IL-4 and IL-13, and Stat6 knock-out animals have been shown to be defective in TH2 differentiation (Kaplan, M.H. et al., 1996, Imm. 4:313-319; Takeda, K. et al., 1996, Nature 380:627-630; and Shimoda, K. et al., 1996, Nature 380:630-633). The results presented herein indicate that induction of tolerance, by causing lower Stat4 and higher Stat6 levels, shifts the differentiation of T cells toward the TH2 lineage.

To assess the importance of Stat6 and the TH2 response in MR-1 mediated tolerance induction, hearts from B6/129 background mice were transplanted into three Stat6 (-/-) mice with a Balb/c background. The day of the surgery the animals were injected with MR-1 and splenocytes from the donor animals. In normal animals, such a transplant results in indefinite graft survival. Stat6 (-/-) animals, however, readily rejected the transplanted hearts at day 11. This result points out the importance of Stat6 in the induction of tolerance, e.g., the induction of tolerance via MR-1 administration.

The contents of all references, patents and published patent applications cited throughout this application are hereby incorporated by reference.

Equivalents

Those skilled in the art will recognize, or be able to ascertain using no more than routine experimentation, many equivalents to the specific embodiments of the invention described herein. Such equivalents are intended to be encompassed by the following claims.

WHAT IS CLAIMED IS:

1. A method for monitoring acceptance of a transplant in a subject mammal that has undergone a transplant, comprising: determining the amount of at least one of (i) to
5 (iv), as follows:
- (i) Stat4 mRNA or Stat4 protein,
 - (ii) Stat6 mRNA or Stat6 protein,
 - (iii) SOCS1 mRNA or SOCS1 protein, or
 - (iv) SOCS3 mRNA or SOCS3 protein,
- 10 present in a transplant sample from the subject.
2. The method of Claim 1, comprising determining the amount of at least two of (i) to (iv) present in the transplant sample.
- 15 3. The method of Claim 1, comprising determining the amount of at least three of (i) to (iv) present in the transplant sample.
4. The method of Claim 1, comprising determining the amount of (i) to (iv) present in the transplant sample.
- 20 5. A method for monitoring acceptance of a transplant in a subject mammal that has undergone a transplant, comprising: determining the amount of Stat4 and Stat6 mRNA or Stat4 and Stat6 protein present in a transplant sample from the subject.
- 25 6. The method of Claim 5, further comprising determining the ratio of Stat4 to Stat6 amounts.
7. The method of Claim 1, 2, 3, 4, 5, or 6, comprising comparing the amount or ratio determined to that present in a control sample.
- 30 8. The method of Claim 7, wherein the control sample is a corresponding pre-transplant subject sample.
9. The method of Claim 7, wherein the control sample is a subject blood
35 sample.

10. The method of Claim 7, wherein the amount of Stat4, SOCS1, or SOCS3 mRNA or protein in the transplant sample is greater than, or the amount of Stat6 mRNA or protein in the transplant sample is less than that of the control sample, thereby indicating that acceptance of the transplant has not been induced or is not being maintained.

5

11. The method of Claim 7, wherein the amount of Stat4, SOCS1, or SOCS3 mRNA or protein in the transplant sample is less than, or the amount of Stat6 mRNA or protein in the transplant sample is equal to or greater than that of the control sample, thereby indicating that acceptance of the transplant has occurred, is being induced or is
10 being maintained.

12. The method of Claim 1 or 5, wherein the amount of mRNA is determined.

13. The method of Claim 1 or 5, wherein the amount of protein is determined.

15

14. The method of Claim 1 or 5, wherein the transplant is an allograft.

15. The method of Claim 1 or 5, wherein the transplant is a heart, liver, kidney, lung, bone marrow, skin, muscle, pancreatic islet, intestine or cornea transplant.

20

16. The method of Claim 1, further comprising assaying the transplant sample for evidence of cell injury.

17. A method for monitoring an autoimmune disorder in a subject mammal,
25 comprising: determining the amount of at least one of (i) to (iv), as follows:

- (i) Stat4 mRNA or Stat4 protein,
- (ii) Stat6 mRNA or Stat6 protein,
- (iii) SOCS1 mRNA or SOCS protein, or
- (iv) SOCS3 mRNA or SOCS protein,

30 present in a sample mammal from a subject being treated for or suspected of exhibiting the disorder, wherein the sample is obtained from a tissue affected by the disorder.

18. The method of Claim 17, comprising determining the amount of at least two of (i) to (iv) present in the sample.

35

19. The method of Claim 17, comprising determining the amount of at least three of (i) to (iv) present in the sample.

20. The method of Claim 17, further comprising determining the amount of (i) to (iv) present in the sample.

21. A method for monitoring an autoimmune disorder in a subject mammal comprising: determining the amount of Stat4 and Stat6 mRNA or Stat4 and Stat6 protein present in a sample from a subject mammal being treated for or suspected of exhibiting the disorder, wherein the sample is obtained from a tissue affected by the disorder.

22. The method of Claim 21, further comprising determining the ratio of Stat4 to Stat6 amounts.

23. The method of Claim 17, 18, 19, 20, 21 or 22, further comprising comparing the amount or ratio determined to that present in a control sample.

24. The method of Claim 23, wherein the control sample is a corresponding tissue not affected by the disorder.

25. The method of Claim 23, wherein the control sample is a subject blood sample.

26. The method of Claim 17, wherein the amount of Stat4, SOCS1, or SOCS3 mRNA or protein in the sample is greater than, or the amount of Stat6 mRNA or protein in the sample is less than, that of the control sample, thereby indicating that the subject mammal exhibits or continues to exhibit the disorder.

27. The method of Claim 17, wherein the amount of Stat4, SOCS1, or SOCS3 mRNA or protein in the sample is less than, or the amount of Stat6 mRNA or protein in the sample is equal to or greater than, that of the control sample, thereby indicating that the subject mammal does not exhibit the disorder or that treatment for the disorder is effective.

28. The method of Claim 17 or 21, wherein the amount of mRNA is determined.

29. The method of Claim 17 or 21, wherein the amount of protein is determined.

30. The method of Claim 17 or 21, wherein the autoimmune disorder is systemic lupus erythematosus, glomerulonephritis, rheumatoid arthritis, Wegener's granulomatosis, chronic active hepatitis, or vasculitis.

5 31. A method for identifying a compound to be tested for an ability to reduce immune rejection, comprising:

- (a) contacting an activated T cell sample with a test compound;
- (b) determining the amount of at least one of (i) to (iv), as follows:
 - 10 (i) Stat4 mRNA or Stat4 protein,
 - (ii) Stat6 mRNA or Stat6 protein,
 - (iii) SOCS1 mRNA or SOCS1 protein, or
 - (iv) SOCS3 mRNA or SOCS3 protein, present in (a); and
- (c) comparing the amount(s) in (a) to that/those present in a
15 corresponding control activated T cell sample that has not been contacted with the test compound,

so that if the amount of (i), (iii) or (iv) is decreased, or the amount of (ii) is increased, relative to the amount in the control sample, a compound to be tested for an ability to reduce immune rejection is identified.

20 32. The method of Claim 31, comprising determining the amount of at least two of (i) to (iv) present in (a), and comparing the amounts to those present in the control sample.

33. The method of Claim 31, further comprising determining the amount of at
25 least three of (i) to (iv) present in (a), and comparing the amounts to those present in the control sample.

34. The method of Claim 31, further comprising determining the amounts of (i)
to (iv) present in (a), and comparing the amounts to those present in the control sample.
30

35. The method of Claim 31, wherein the amount of mRNA is determined.

36. The method of Claim 31, wherein the amount of protein is determined.
35

37. The method of Claim 32, comprising:

- (a) contacting an activated T cell sample with a test compound;
- (b) determining the amount of Stat4 and Stat6 mRNA or Stat4 and Stat6 protein present in the sample; and
- (c) comparing the amounts in (a) to those present in a corresponding control activated T cell sample that has not been contacted with the test compound,

so that if the amount of Stat4 is decreased or the amount of Stat6 is increased relative to the amount in the control sample, a compound to be tested for an ability to reduce immune rejection is identified.

10

38. A method for identifying a compound to be tested for an ability to reduce immune rejection, comprising:

- (a) contacting an activated T cell sample with a test compound;
- (b) determining the ratio Stat4 mRNA to Stat6 mRNA or Stat4 protein to Stat6 protein present in the sample; and
- (c) comparing the ratio in (a) to that present in a corresponding control activated T cell sample that has not been contacted with the test compound,

15

so that if the ratio in the sample is decreased relative to that in the control sample, a compound to be tested for an ability to reduce immune rejection is identified.

20

39. A method for identifying a compound to be tested for an ability to reduce immune rejection, comprising:

- (a) contacting a resting T cell sample, a T cell activator and a test compound;
- (b) determining the amount of at least one of (i) to (iv), as follows:
 - (i) Stat4 mRNA or Stat4 protein,
 - (ii) Stat6 mRNA or Stat6 protein,
 - (iii) SOCS1 mRNA or SOCS1 protein, or
 - (iv) SOCS3 mRNA or SOCS3 protein, present in (a); and
- (c) comparing the amount(s) in (a) to that/those present in a corresponding control activated T cell sample that has not been contacted with the test compound,

25

30

so that if the amount of (i), (iii) or (iv) is decreased, or the amount of (ii) is increased, relative to the amount in the control sample, a compound to be tested for an ability to reduce immune rejection is identified.

35

40. The method of Claim 39, comprising determining the amount of at least two of (i) to (iv) present in (a), and comparing the amounts to those present in the control sample.

5 41. The method of Claim 39, further comprising determining the amount of at least three of (i) to (iv) present in (a), and comparing the amounts to those present in the control sample.

42. The method of Claim 39, comprising determining the amounts of (i) to (iv)
10 present in (a), and comparing the amounts to those present in the control sample.

43. The method of Claim 39, wherein the resting T cell is a primary T cell.

15 44. The method of Claim 37, wherein the resting T cell is a T cell line.

45. The method of Claim 39, wherein the T cell activator is a mitogen or a T cell receptor stimulatory factor.

20 46. The method of Claim 45, wherein the T cell activator is an anti-CD3 antibody.

47. The method of Claim 40, comprising:
25 (a) contacting a resting T cell sample, a T cell activator and a test compound;
(b) determining the amount of Stat4 and Stat6 mRNA or Stat4 and Stat6 protein present in the sample; and
(c) comparing the amounts in (b) to those present in a corresponding control activated T cell sample that has not been contacted with the
30 test compound,
so that if the amount of Stat4 is decreased or the amount of Stat6 is increased relative to the amounts in the control sample, a compound to be tested for an ability to reduce immune rejection is identified.

35 48. A method for identifying a compound to be tested for an ability to reduce immune rejection, comprising:

- (a) contacting a resting T cell sample, a T cell activator and a test compound;
- (b) determining the ratio of Stat4 mRNA to Stat6 mRNA or Stat4 mRNA to Stat6 protein present in the sample; and
- 5 (c) comparing the ratio to in (a) to that present in a corresponding control activated T cell sample that has not been contacted with the test compound,

so that if the ratio in the sample is decreased relative to that in the control sample, a compound to be tested for an ability to reduce immune rejection is identified.

10

49. A method for identifying a compound to be tested for an ability to reduce immune rejection, comprising:

- (a) contacting a T cell sample, a cytokine and a test compound, wherein the T cell sample is responsive to the cytokine;
- 15 (b) determining the amount of at least one of (i) to (iv), as follows:
- (i) Stat4 mRNA or Stat4 protein,
 - (ii) Stat6 mRNA or Stat6 protein,
 - (iii) SOCS1 mRNA or SOCS1 protein, or
 - (iv) SOCS3 mRNA or SOCS3 protein, present in (a); and
- 20 (c) comparing the amount(s) in (a) to that/those present in a corresponding control T cell sample that has not been contacted with the test compound,

so that if the amount of (i), (iii) or (iv) is decreased, or the amount of (ii) is increased, relative to the amount in the control sample, a compound to be tested for an ability to
25 reduce immune rejection is identified.

50. The method of Claim 49, comprising determining the amount of at least two of (i) to (iv) present in (a), and comparing the amounts to those present in the control sample.

30

51. The method of Claim 49, further comprising determining the amount of at least three of (i) to (iv) present in (a), and comparing the amounts to those present in the control sample.

35

52. The method of Claim 49, comprising determining the amounts of (i) to (iv) present in (a), and comparing the amounts to those present in the control sample.

5 53. The method of Claim 49, wherein the T cell is a T cell line.

54. The method of Claim 49, wherein the cytokine is IL-2, IL-4, IL-12, or IL-13.

55. The method of Claim 50, comprising:

- 10 (a) contacting a resting T cell sample, a cytokine and a test compound, wherein the T cell sample is responsive to the cytokine;
- (b) determining the amount of Stat4 and Stat6 mRNA or Stat4 and Stat6 protein present in the sample; and
- 15 (c) comparing the amounts in (b) to those present in a corresponding control T cell sample that has not been contacted with the test compound,

so that if the amount of Stat4 is decreased or the amount of Stat6 is increased relative to the amounts in the control sample, a compound to be tested for an ability to reduce immune rejection is identified.

20

56. A method for identifying a compound to be tested for an ability to reduce immune rejection, comprising:

- (a) contacting a T cell sample, a cytokine and a test compound, wherein the T cell sample is responsive to the cytokine;
- 25 (b) determining the ratio of Stat4 mRNA to Stat6 mRNA or Stat4 mRNA to Stat6 protein present in the sample; and
- (c) comparing the ratio to in (b) to that present in a corresponding control T cell sample that has not been contacted with the test compound,

30 so that if the ratio in the sample is decreased relative to that in the control sample, a compound to be tested for an ability to reduce immune rejection is identified.

57. A method for reducing immune rejection in a subject mammal, comprising: administering to a subject mammal in need of such a reduction a concentration of a
35 compound sufficient to reduce the level of Stat4 mRNA or protein in the subject relative to

that observed in the subject in the absence of the compound, wherein said compound does not induce platelet aggregation and does not affect NF- κ B activation in CD40L⁺ cells.

58. The method of Claim 57, wherein upon administration of the compound, the
5 level of Stat6 mRNA or protein in the subject is maintained or increased relative to that observed in the subject in the absence of the compound.

59. A method for reducing immune rejection in a subject mammal, comprising:
administering to a subject mammal in need of such a reduction a concentration of a
10 compound sufficient to increase the level of Stat6 mRNA or protein in the subject relative to that observed in the subject in the absence of the compound, wherein said compound does not induce platelet aggregation and does not affect NF- κ B activation in CD40L⁺ cells.

60. The method of Claim 57, 58 or 59, wherein the subject mammal has
15 undergone a transplant.

61. The method of Claim 60, wherein administering the compound induces tolerance in the subject mammal.

20 62. The method of Claim 57, 58 or 59, wherein the subject mammal being treated exhibits an autoimmune disorder.

63. The method of Claim 57, 58 or 59, wherein the compound is a tyrosine kinase inhibitor.
25

64. The method of Claim 63, wherein the tyrosine kinase inhibitor is a tyrphostin compound.

65. The method of Claim 63, wherein the tyrosine kinase inhibitor inhibits a Jak
30 enzyme.

66. The method of Claim 65, wherein the Jak inhibited is Jak2.

67. The method of Claim 57, 58, or 59, wherein the compound is a Jak2
35 inhibitor.


```

attaaacctc tcgccgagcc cctccgcaga ctctgcgccg gaaagtttca tttgctgtat      60
gccatcctcg agagctgtct aggttaacgt tcgcactctg tgtatataac ctgcacagtc      120
ttggcaccta acgtgctgtg cgtagctgct cctttggttg aatccccagg cccttggttg      180
ggcacaaggt ggcagg atg tct cag tgg tac gaa ctt cag cag ctt gac tca      232
      Met Ser Gln Trp Tyr Glu Leu Gln Gln Leu Asp Ser
              1              5              10

aaa ttc ctg gag cag gtt cac cag ctt tat gat gac agt ttt ccc atg      280
Lys Phe Leu Glu Gln Val His Gln Leu Tyr Asp Asp Ser Phe Pro Met
              15              20              25

gaa atc aga cag tac ctg gca cag tgg tta gaa aag caa gac tgg gag      324
Glu Ile Arg Gln Tyr Leu Ala Gln Trp Leu Glu Lys Gln Asp Trp Glu
              30              35              40

cac gct gcc aat gat gtt tca ttt gcc acc atc cgt ttt cat gac ctc      376
His Ala Ala Asn Asp Val Ser Phe Ala Thr Ile Arg Phe His Asp Leu
              45              50              55              60

ctg tca cag ctg gat gat caa tat agt cgc ttt tct ttg gag aat aac      424
Leu Ser Gln Leu Asp Asp Gln Tyr Ser Arg Phe Ser Leu Glu Asn Asn
              65              70              75

ttc ttg cta cag cat aac ata agg aaa agc aag cgt aat ctt cag gat      472
Phe Leu Leu Gln His Asn Ile Arg Lys Ser Lys Arg Asn Leu Gln Asp
              80              85              90

aat ttt cag gaa gac cca atc cag atg tct atg atc att tac agc tgt      520
Asn Phe Gln Glu Asp Pro Ile Gln Met Ser Met Ile Ile Tyr Ser Cys
              95              100              105

ctg aag gaa gaa agg aaa att ctg gaa aac gcc cag aga ttt aat cag      568
Leu Lys Glu Glu Arg Lys Ile Leu Glu Asn Ala Gln Arg Phe Asn Gln
              110              115              120

gct cag tcg ggg aat att cag agc aca gtg atg tta gac aaa cag aaa      616
Ala Gln Ser Gly Asn Ile Gln Ser Thr Val Met Leu Asp Lys Gln Lys
              125              130              135              140

gag ctt gac agt aaa gtc aga aat gtg aag gac aag gtt atg tgt ata      664
Glu Leu Asp Ser Lys Val Arg Asn Val Lys Asp Lys Val Met Cys Ile
              145              150              155

gag cat gaa atc aag agc ctg gaa gat tta caa gat gaa tat gac ttc      712
Glu His Glu Ile Lys Ser Leu Glu Asp Leu Gln Asp Glu Tyr Asp Phe
              160              165              170

aaa tgc aaa acc ttg cag aac aga gaa cac gag acc aat ggt gtg gca      760
Lys Cys Lys Thr Leu Gln Asn Arg Glu His Glu Thr Asn Gly Val Ala
              175              180              185

aag agt gat cag aaa caa gaa cag ctg tta ctc aag aag atg tat tta      808
Lys Ser Asp Gln Lys Gln Glu Gln Leu Leu Leu Lys Lys Met Tyr Leu
              190              195              200

atg ctt gac aat aag aga aag gaa gta gtt cac aaa ata ata gag ttg      856
Met Leu Asp Asn Lys Arg Lys Glu Val Val His Lys Ile Ile Glu Leu
              205              210              215              220

```

Figure 1

ctg aat gtc act gaa ctt acc cag aat gcc ctg att aat gat gaa cta Leu Asn Val Thr Glu Leu Thr Gln Asn Ala Leu Ile Asn Asp Glu Leu 225 230 235	904
gtg gag tgg aag cgg aga cag cag agc gcc tgt att ggg ggg ccg ccc Val Glu Trp Lys Arg Arg Gln Gln Ser Ala Cys Ile Gly Gly Pro Pro 240 245 250	952
aat gct tgc ttg gat cag ctg cag aac tgg ttc act ata gtt gcg gag Asn Ala Cys Leu Asp Gln Leu Gln Asn Trp Phe Thr Ile Val Ala Glu 255 260 265	1000
agt ctg cag caa gtt cgg cag cag ctt aaa aag ttg gag gaa ttg gaa Ser Leu Gln Gln Val Arg Gln Gln Leu Lys Lys Leu Glu Glu Leu Glu 270 275 280	1048
cag aaa tac acc tac gaa cat gac cct atc aca aaa aac aaa caa gtg Gln Lys Tyr Thr Tyr Glu His Asp Pro Ile Thr Lys Asn Lys Gln Val 285 290 295 300	1096
tta tgg gac cgc acc ttc agt ctt ttc cag cag ctc att cag agc tcg Leu Trp Asp Arg Thr Phe Ser Leu Phe Gln Gln Leu Ile Gln Ser Ser 305 310 315	1144
ttt gtg gtg gaa aga cag ccc tgc atg cca acg cac cct cag agg ccg Phe Val Val Glu Arg Gln Pro Cys Met Pro Thr His Pro Gln Arg Pro 320 325 330	1192
ctg gtc ttg aag aca ggg gtc cag ttc act gtg aag ttg aga ctg ttg Leu Val Leu Lys Thr Gly Val Gln Phe Thr Val Lys Leu Arg Leu Leu 335 340 345	1240
gtg aaa ttg caa gag ctg aat tat aat ttg aaa gtc aaa gtc tta ttt Val Lys Leu Gln Glu Leu Asn Tyr Asn Leu Lys Val Lys Val Leu Phe 350 355 360	1288
gat aaa gat gtg aat gag aga aat aca gta aaa gga ttt agg aag ttc Asp Lys Asp Val Asn Glu Arg Asn Thr Val Lys Gly Phe Arg Lys Phe 365 370 375 380	1336
aac att ttg ggc acg cac aca aaa gtg atg aac atg gag gag tcc acc Asn Ile Leu Gly Thr His Thr Lys Val Met Asn Met Glu Glu Ser Thr 385 390 395	1384
aat ggc agt ctg gcg gct gaa ttt cgg cac ctg caa ttg aaa gaa cag Asn Gly Ser Leu Ala Ala Glu Phe Arg His Leu Gln Leu Lys Glu Gln 400 405 410	1432
aaa aat gct ggc acc aga acg aat gag ggt cct ctc atc gtt act gaa Lys Asn Ala Gly Thr Arg Thr Asn Glu Gly Pro Leu Ile Val Thr Glu 415 420 425	1480
gag ctt cac tcc ctt agt ttt gaa acc caa ttg tgc cag cct ggt ttg Glu Leu His Ser Leu Ser Phe Glu Thr Gln Leu Cys Gln Pro Gly Leu 430 435 440	1528
gta att gac ctc gag acg acc tct ctg ccc gtt gtg gtg atc tcc aac Val Ile Asp Leu Glu Thr Thr Ser Leu Pro Val Val Val Ile Ser Asn 445 450 455 460	1576

Figure 1 (cont'd)

gtc agc cag ctc ccg agc ggt tgg gcc tcc atc ctt tgg tac aac atg	1624
Val Ser Gln Leu Pro Ser Gly Trp Ala Ser Ile Leu Trp Tyr Asn Met	
465 470 475	
ctg gtg gcg gaa ccc agg aat ctg tcc ttc ttc ctg act cca cca tgt	1672
Leu Val Ala Glu Pro Arg Asn Leu Ser Phe Phe Leu Thr Pro Pro Cys	
480 485 490	
gca cga tgg gct cag ctt tca gaa gtg ctg agt tgg cag ttt tct tct	1720
Ala Arg Trp Ala Gln Leu Ser Glu Val Leu Ser Trp Gln Phe Ser Ser	
495 500 505	
gtc acc aaa aga ggt ctc aat gtg gac cag ctg aac atg ttg gga gag	1768
Val Thr Lys Arg Gly Leu Asn Val Asp Gln Leu Asn Met Leu Gly Glu	
510 515 520	
aag ctt ctt ggt cct aac gcc agc ccc gat ggt ctc att ccg tgg acg	1816
Lys Leu Leu Gly Pro Asn Ala Ser Pro Asp Gly Leu Ile Pro Trp Thr	
525 530 535 540	
agg ttt tgt aag gaa aat ata aat gat aaa aat ttt ccc ttc tgg ctt	1864
Arg Phe Cys Lys Glu Asn Ile Asn Asp Lys Asn Phe Pro Phe Trp Leu	
545 550 555	
tgg att gaa agc atc cta gaa ctc att aaa aaa cac ctg ctc cct ctc	1912
Trp Ile Glu Ser Ile Leu Glu Leu Ile Lys Lys His Leu Leu Pro Leu	
560 565 570	
tgg aat gat ggg tgc atc atg ggc ttc atc agc aag gag cga gag cgt	1960
Trp Asn Asp Gly Cys Ile Met Gly Phe Ile Ser Lys Glu Arg Glu Arg	
575 580 585	
gcc ctg ttg aag gac cag cag ccg ggg acc ttc ctg ctg cgg ttc agt	2008
Ala Leu Leu Lys Asp Gln Gln Pro Gly Thr Phe Leu Leu Arg Phe Ser	
590 595 600	
gag agc tcc cgg gaa ggg gcc atc aca ttc aca tgg gtg gag cgg tcc	2056
Glu Ser Ser Arg Glu Gly Ala Ile Thr Phe Thr Trp Val Glu Arg Ser	
605 610 615 620	
cag aac gga ggc gaa cct gac ttc cat gcg gtt gaa ccc tac acg aag	2104
Gln Asn Gly Gly Glu Pro Asp Phe His Ala Val Glu Pro Tyr Thr Lys	
625 630 635	
aaa gaa ctt tct gct gtt act ttc cct gac atc att cgc aat tac aaa	2152
Lys Glu Leu Ser Ala Val Thr Phe Pro Asp Ile Ile Arg Asn Tyr Lys	
640 645 650	
gtc atg gct gct gag aat att cct gag aat ccc ctg aag tat ctg tat	2200
Val Met Ala Ala Glu Asn Ile Pro Glu Asn Pro Leu Lys Tyr Leu Tyr	
655 660 665	
cca aat att gac aaa gac cat gcc ttt gga aag tat tac tcc agg cca	2248
Pro Asn Ile Asp Lys Asp His Ala Phe Gly Lys Tyr Tyr Ser Arg Pro	
670 675 680	

Figure 1 (cont'd)

aag gaa gca cca gag cca atg gaa ctt gat ggc cct aaa gga act gga Lys Glu Ala Pro Glu Pro Met Glu Leu Asp Gly Pro Lys Gly Thr Gly 685 690 695 700	2296
tat atc aag act gag ttg att tct gtg tct gaa gtt cac cct tct aga Tyr Ile Lys Thr Glu Leu Ile Ser Val Ser Glu Val His Pro Ser Arg 705 710 715	2344
ctt cag acc aca gac aac ctg ctc ccc atg tct cct gag gag ttt gac Leu Gln Thr Thr Asp Asn Leu Leu Pro Met Ser Pro Glu Glu Phe Asp 720 725 730	2392
gag gtg tct cgg ata gtg ggc tct gta gaa ttc gac agt atg atg aac Glu Val Ser Arg Ile Val Gly Ser Val Glu Phe Asp Ser Met Met Asn 735 740 745	2440
aca gta tag agcatgaatt tttttcatct tctctggcga cagttttcct Thr Val * 750	2489
tctcatctgt gattccctcc tgctactctg ttccttcaca tccgtgtgttt ctagggaat gaaagaaagg ccagcaaatt cgctgcaacc tgttgatagc aagtgaattt ttctctaact cagaaacatc agttactctg aagggcacatc tgcactctac tgaaggtaaa attgaaaggc attctctgaa gagtgggttt cacaagttaa aaacatccag atacacccaa agtatcagga cgagaatgag ggtccttttg gaaaggagaa gtttaagcaac atctagcaaa tgttatgcat aaagtcagt cccaactgtt ataggttgtt ggataaatca gtgggtattt agggaaactgc ttgacgtagg aacggtaaat ttctgtggga gaattcttac atgttttctt tgctttaagt gtaactggca gttttccatt ggtttacctg tgaaatagtt caaagccaag tttatatata attatatcag tctcttttca aaggtagcca tcatggatct ggtaggggga aaatgtgtat tttattacat ctttcacatt ggctatttaa agacaaagac aaattctgtt tcttgagaag agaatattag ctttactgtt tgttatggct taatgacact agctaataac aatagaagga tgtacatttc caaattcaca agttgtgttt gatattccaa gctgaataca ttctgctttc atcttggtea catacaatta tttttacagt tctcccaagg gagttaggct attcacaacc actcattcaa aagttgaaat taaccataga tgtagataaa ctcagaaatt taattcatgt ttcttaaatg ggctactttg tcttttttgt tattaggggtg gtatttagtc tattagccac aaaattggga aaggagtaga aaaagcagta actgacaact tgaataatac accagagata atatgagaat cagatcattt caaaactcat ttcctatgta actgcattga gaactgcata tgtttcgctg atatatgtgt ttttcacatt tgcgaatggt tccattctct ctctgtact ttttccagac acttttttga gtggatgatg tttcgtgaag tatactgtat ttttaccttt ttccttcctt atcactgaca caaaaagtag attaagagat gggtttgaca aggttcttcc cttttacata ctgctgtcta tgtggctgta tcttggtttt ccactactgc taccacaact atattatcat gcaaatgctg tattcttctt tgggtggagat aaagatttct tgagttttgt tttaaaatta aagctaaagt atctgtattg cattaaatat aatatcgaca cagtgttttc cgtggcactg catacaatct gaggcctcct ctctcagttt ttatatagat ggcgagaacc taagtttcag ttgattttac aattgaaatg actaaaaaac aaagaagaca acattaaaaa caatattgtt tcta	2549 2609 2669 2729 2789 2849 2909 2969 3029 3089 3149 3209 3269 3329 3389 3449 3509 3569 3629 3689 3749 3809 3869 3929 3989 4003

Figure 1 (cont'd)

aagtcg	cgcgac	cagagccatt	ggagggcgcg	gggactgcaa	ccctaatacag	agcccaa	atg	60								
							Met									
							1									
gcg	cag	tgg	gaa	atg	ctg	cag	aat	ctt	gac	agc	ccc	ttt	cag	gat	cag	108
Ala	Gln	Trp	Glu	Met	Leu	Gln	Asn	Leu	Asp	Ser	Pro	Phe	Gln	Asp	Gln	
		5						10					15			
ctg	cac	cag	ctt	tac	tgc	cac	agc	ctc	ctg	cct	gtg	gac	att	cga	cag	156
Leu	His	Gln	Leu	Tyr	Ser	His	Ser	Leu	Leu	Pro	Val	Asp	Ile	Arg	Gln	
		20					25					30				
tac	ttg	gct	gtc	tgg	att	gaa	gac	cag	aac	tgg	cag	gaa	gct	gca	ctt	204
Tyr	Leu	Ala	Val	Trp	Ile	Glu	Asp	Gln	Asn	Trp	Gln	Glu	Ala	Ala	Leu	
	35					40					45					
ggg	agt	gat	gat	tcc	aag	gct	acc	atg	cta	ttc	ttc	cac	ttc	ttg	gat	252
Gly	Ser	Asp	Asp	Ser	Lys	Ala	Thr	Met	Leu	Phe	Phe	His	Phe	Leu	Asp	
	50				55					60					65	
cag	ctg	aac	tat	gag	tgt	ggc	cgt	tgc	agc	cag	gac	cca	gag	tcc	ttg	300
Gln	Leu	Asn	Tyr	Glu	Cys	Gly	Arg	Cys	Ser	Gln	Asp	Pro	Glu	Ser	Leu	
				70					75						80	
ttg	ctg	cag	cac	aat	ttg	cgg	aaa	ttc	tgc	cgg	gac	att	cag	ccc	ttt	348
Leu	Leu	Gln	His	Asn	Leu	Arg	Lys	Phe	Cys	Arg	Asp	Ile	Gln	Pro	Phe	
			85					90					95			
tcc	cag	gat	cct	acc	cag	ttg	gct	gag	atg	atc	ttt	aac	ctc	ctt	ctg	396
Ser	Gln	Asp	Pro	Thr	Gln	Leu	Ala	Glu	Met	Ile	Phe	Asn	Leu	Leu	Leu	
		100					105					110				
gaa	gaa	aaa	aga	att	ttg	atc	cag	gct	cag	agg	gcc	caa	ttg	gaa	caa	444
Glu	Glu	Lys	Arg	Ile	Leu	Ile	Gln	Ala	Gln	Arg	Ala	Gln	Leu	Glu	Gln	
	115					120					125					
gga	gag	cca	gtt	ctc	gaa	aca	cct	gtg	gag	agc	cag	caa	cat	gag	att	492
Gly	Glu	Pro	Val	Leu	Glu	Thr	Pro	Val	Glu	Ser	Gln	Gln	His	Glu	Ile	
	130				135					140				145		
gaa	tcc	cgg	atc	ctg	gat	tta	agg	gct	atg	atg	gag	aag	ctg	gta	aaa	540
Glu	Ser	Arg	Ile	Leu	Asp	Leu	Arg	Ala	Met	Met	Glu	Lys	Leu	Val	Lys	
				150					155					160		
tcc	atc	agc	caa	ctg	aaa	gac	cag	cag	gat	gtc	ttc	tgc	ttc	cga	tat	588
Ser	Ile	Ser	Gln	Leu	Lys	Asp	Gln	Gln	Asp	Val	Phe	Cys	Phe	Arg	Tyr	
			165					170					175			
aag	atc	cag	gcc	aaa	ggg	aag	aca	ccc	tct	ctg	gac	ccc	cat	cag	acc	636
Lys	Ile	Gln	Ala	Lys	Gly	Lys	Thr	Pro	Ser	Leu	Asp	Pro	His	Gln	Thr	
		180					185					190				
aaa	gag	cag	aag	att	ctg	cag	gaa	act	ctc	aat	gaa	ctg	gac	aaa	agg	684
Lys	Glu	Gln	Lys	Ile	Leu	Gln	Glu	Thr	Leu	Asn	Glu	Leu	Asp	Lys	Arg	
	195					200					205					
aga	aag	gag	gtg	ctg	gat	gcc	tcc	aaa	gca	ctg	cta	ggc	cga	tta	act	732
Arg	Lys	Glu	Val	Leu	Asp	Ala	Ser	Lys	Ala	Leu	Leu	Gly	Arg	Leu	Thr	
	210				215					220				225		

Figure 2

acc cta atc gag cta ctg ctg cca aag ttg gag gag tgg aag gcc cag	780
Thr Leu Ile Glu Leu Leu Leu Pro Lys Leu Glu Glu Trp Lys Ala Gln	
230 235 240	
cag caa aaa gcc tgc atc aga gct ccc att gac cac ggg ttg gaa cag	828
Gln Gln Lys Ala Cys Ile Arg Ala Pro Ile Asp His Gly Leu Glu Gln	
245 250 255	
ctg gag aca tgg ttc aca gct gga gca aag ctg ttg ttt cac ctg agg	876
Leu Glu Thr Trp Phe Thr Ala Gly Ala Lys Leu Leu Phe His Leu Arg	
260 265 270	
cag ctg ctg aag gag ctg aag gga ctg agt tgc ctg gtt agc tat cag	924
Gln Leu Leu Lys Glu Leu Lys Gly Leu Ser Cys Leu Val Ser Tyr Gln	
275 280 285	
gat gac cct ctg acc aaa ggg gtg gac cta cgc aac gcc cag gtc aca	972
Asp Asp Pro Leu Thr Lys Gly Val Asp Leu Arg Asn Ala Gln Val Thr	
290 295 300 305	
gag ttg cta cag cgt ctg ctc cac aga gcc ttt gtg gta gaa acc cag	1020
Glu Leu Leu Gln Arg Leu Leu His Arg Ala Phe Val Val Glu Thr Gln	
310 315 320	
ccc tgc atg ccc caa act ccc cat cga ccc ctc atc ctc aag act ggc	1068
Pro Cys Met Pro Gln Thr Pro His Arg Pro Leu Ile Leu Lys Thr Gly	
325 330 335	
agc aag ttc acc gtc cga aca agg ctg ctg gtg aga ctc cag gaa ggc	1116
Ser Lys Phe Thr Val Arg Thr Arg Leu Leu Val Arg Leu Gln Glu Gly	
340 345 350	
aat gag tca ctg act gtg gaa gtc tcc att gac agg aat cct cct caa	1164
Asn Glu Ser Leu Thr Val Glu Val Ser Ile Asp Arg Asn Pro Pro Gln	
355 360 365	
tta caa ggc ttc cgg aag ttc aac att ctg act tca aac cag aaa act	1212
Leu Gln Gly Phe Arg Lys Phe Asn Ile Leu Thr Ser Asn Gln Lys Thr	
370 375 380 385	
ttg acc ccc gag aag ggg cag agt cag ggt ttg att tgg gac ttt ggt	1260
Leu Thr Pro Glu Lys Gly Gln Ser Gln Gly Leu Ile Trp Asp Phe Gly	
390 395 400	
tac ctg act ctg gtg gag caa cgt tca ggt ggt tca gga aag ggc agc	1308
Tyr Leu Thr Leu Val Glu Gln Arg Ser Gly Gly Ser Gly Lys Gly Ser	
405 410 415	
aat aag ggg cca cta ggt gtg aca gag gaa ctg cac atc atc agc ttc	1356
Asn Lys Gly Pro Leu Gly Val Thr Glu Glu Leu His Ile Ile Ser Phe	
420 425 430	
acg gtc aaa tat acc tac cag ggt ctg aag cag gag ctg aaa acg gac	1404
Thr Val Lys Tyr Thr Tyr Gln Gly Leu Lys Gln Glu Leu Lys Thr Asp	
435 440 445	

Figure 2 (cont'd)

acc ctc cct gtg gtg att att tcc aac atg aac cag ctc tca att gcc	1452
Thr Leu Pro Val Val Ile Ile Ser Asn Met Asn Gln Leu Ser Ile Ala	
450 455 460 465	
tgg gct tca gtt ctc tgg ttc aat ttg ctc agc cca aac ctt cag aac	1500
Trp Ala Ser Val Leu Trp Phe Asn Leu Leu Ser Pro Asn Leu Gln Asn	
470 475 480	
cag cag ttc ttc tcc aac ccc ccc aag gcc ccc tgg agc ttg ctg ggc	1548
Gln Gln Phe Phe Ser Asn Pro Pro Lys Ala Pro Trp Ser Leu Leu Gly	
485 490 495	
cct gct ctc agt tgg cag ttc tcc tcc tat gtt ggc cga ggc ctc aac	1596
Pro Ala Leu Ser Trp Gln Phe Ser Ser Tyr Val Gly Arg Gly Leu Asn	
500 505 510	
tca gac cag ctg agc atg ctg aga aac aag ctg ttc ggg cag aac tgt	1644
Ser Asp Gln Leu Ser Met Leu Arg Asn Lys Leu Phe Gly Gln Asn Cys	
515 520 525	
agg act gag gat cca tta ttg tcc tgg gct gac ttc act aag cga gag	1692
Arg Thr Glu Asp Pro Leu Leu Ser Trp Ala Asp Phe Thr Lys Arg Glu	
530 535 540 545	
agc cct cct ggc aag tta cca ttc tgg aca tgg ctg gac aaa att ctg	1740
Ser Pro Pro Gly Lys Leu Pro Phe Trp Thr Trp Leu Asp Lys Ile Leu	
550 555 560	
gag ttg gta cat gac cac ctg aag gat ctc tgg aat gat gga cgc atc	1788
Glu Leu Val His Asp His Leu Lys Asp Leu Trp Asn Asp Gly Arg Ile	
565 570 575	
atg ggc ttt gtg agt cgg agc cag gag cgc cgg ctg ctg aag aag acc	1836
Met Gly Phe Val Ser Arg Ser Gln Glu Arg Arg Leu Leu Lys Lys Thr	
580 585 590	
atg tct ggc acc ttt cta ctg cgc ttc agt gaa tgc tca gaa ggg ggc	1884
Met Ser Gly Thr Phe Leu Leu Arg Phe Ser Glu Ser Ser Glu Gly Gly	
595 600 605	
att acc tgc tcc tgg gtg gag cac cag gat gat gac aag gtg ctc atc	1932
Ile Thr Cys Ser Trp Val Glu His Gln Asp Asp Asp Lys Val Leu Ile	
610 615 620 625	
tac tct gtg caa ccg tac acg aag gag gtg ctg cag tca ctc ccg ctg	1980
Tyr Ser Val Gln Pro Tyr Thr Lys Glu Val Leu Gln Ser Leu Pro Leu	
630 635 640	
act gaa atc atc cgc cat tac cag ttg ctc act gag gag aat ata cct	2028
Thr Glu Ile Ile Arg His Tyr Gln Leu Leu Thr Glu Glu Asn Ile Pro	
645 650 655	
gaa aac cca ctg cgc ttc ctc tat ccc cga atc ccc cgg gat gaa gct	2076
Glu Asn Pro Leu Arg Phe Leu Tyr Pro Arg Ile Pro Arg Asp Glu Ala	
660 665 670	
ttt ggg tgc tac tac cag gag aaa gtt aat ctc cag gaa cgg agg aaa	2124
Phe Gly Cys Tyr Tyr Gln Glu Lys Val Asn Leu Gln Glu Arg Arg Lys	
675 680 685	

Figure 2 (cont'd)

tac ctg aaa cac agg ctc att gtg gtc tct aat aga cag gtg gat gaa	2172
Tyr Leu Lys His Arg Leu Ile Val Val Ser Asn Arg Gln Val Asp Glu	
690 695 700 705	
ctg caa caa ccg ctg gag ctt aag cca gag cca gag ctg gag tca tta	2220
Leu Gln Gln Pro Leu Glu Leu Lys Pro Glu Pro Glu Leu Glu Ser Leu	
710 715 720	
gag ctg gaa cta ggg ctg gtg cca gag cca gag ctc agc ctg gac tta	2268
Glu Leu Glu Leu Gly Leu Val Pro Glu Pro Glu Leu Ser Leu Asp Leu	
725 730 735	
gag cca ctg ctg aag gca ggg ctg gat ctg ggg cca gag cta gag tct	2316
Glu Pro Leu Leu Lys Ala Gly Leu Asp Leu Gly Pro Glu Leu Glu Ser	
740 745 750	
gtg ctg gag tcc act ctg gag cct gtg ata gag ccc aca cta tgc atg	2364
Val Leu Glu Ser Thr Leu Glu Pro Val Ile Glu Pro Thr Leu Cys Met	
755 760 765	
gta tca caa aca gtg cca gag cca gac caa gga cct gta tca cag cca	2412
Val Ser Gln Thr Val Pro Glu Pro Asp Gln Gly Pro Val Ser Gln Pro	
770 775 780 785	
gtg cca gag cca gat ttg ccc tgt gat ctg aga cat ttg aac act gag	2460
Val Pro Glu Pro Asp Leu Pro Cys Asp Leu Arg His Leu Asn Thr Glu	
790 795 800	
cca atg gaa atc ttc aga aac tgt gta aag att gaa gaa atc atg ccg	2508
Pro Met Glu Ile Phe Arg Asn Cys Val Lys Ile Glu Glu Ile Met Pro	
805 810 815	
aat ggt gac cca ctg ttg gct ggc cag aac acc gtg gat gag gtt tac	2556
Asn Gly Asp Pro Leu Leu Ala Gly Gln Asn Thr Val Asp Glu Val Tyr	
820 825 830	
gtc tcc cgc ccc agc cac ttc tac act gat gga ccc ttg atg cct tct	2604
Val Ser Arg Pro Ser His Phe Tyr Thr Asp Gly Pro Leu Met Pro Ser	
835 840 845	
gac ttc tag gaaccacatt tcctctgttc tttcatatc tctttgccct	2653
Asp Phe *	
850	
tctactcct catagcatga tattgtttct caaggatggg aatcaggcat gtgtcccttc	2713
caagctgtgt taactgttca aactcaggcc tgtgtgactc cattgggggtg agaggtgaaa	2773
gcataacatg ggtacagagg ggacaacaat gaatcagaac agatgctgag ccataggtct	2833
aaataggatc ctggaggctg cctgctgtgc tgggaggtat aggggtcctg ggggcaggcc	2893
agggcagttg acaggtactt ggagggctca gggcagtggc ttctttccag tatggaagga	2953
tttcaacatt ttaatagtgt gttaggctaa actggtgcat actggcattg gccttggtgg	3013
ggagcacaga cacaggatag gactccattt ctttcttcca ttccttcattg tctaggataa	3073
cttgctttct tctttccttt actcctggct caagccctga atttcttctt ttcctgcagg	3133
ggttgagagc tttctgcctt agcctaccat gtgaaactct accctgaaga aagggatgga	3193
taggaagtag acctcttttt cttaccagtc tcctccccta ctctgcccc taagctggct	3253
gtacctgttc ctccccata aaatgatcct gccaatct	3291

Figure 2 (cont'd)

cagctggaat	tcggggcggc	ggcgagact	gggaggggga	gccgggggtt	ccgacgtcgc	60
agccgagggg	acaagcccca	accggatcct	ggacaggcac	cccggcttgg	cgctgtctct	120
ccccctcggc	tcggagaggc	ccttcggcct	gagggagcct	cgccgcccgt	ccccggcaca	180
cgcgagcccc	cggcctctcg	gcctctgccg	gagaaacagg	atg gcc caa tgg aat		235
				Met Ala Gln Trp Asn		
				1	5	
cag cta cag cag ctt gac aca cgg tac ctg gag cag ctc cat cag ctc						283
Gln Leu Gln Gln Leu Asp Thr Arg Tyr Leu Glu Gln Leu His Gln Leu						
	10		15		20	
tac agt gac agc ttc cca atg gag ctg cgg cag ttt ctg gcc cct tgg						331
Tyr Ser Asp Ser Phe Pro Met Glu Leu Arg Gln Phe Leu Ala Pro Trp						
	25		30		35	
att gag agt caa gat tgg gca tat gcg gcc agc aaa gaa tca cat gcc						379
Ile Glu Ser Gln Asp Trp Ala Tyr Ala Ala Ser Lys Glu Ser His Ala						
	40		45		50	
act ttg gtg ttt cat aat ctc ctg gga gag att gac cag cag tat agc						427
Thr Leu Val Phe His Asn Leu Leu Gly Glu Ile Asp Gln Gln Tyr Ser						
	55		60		65	
cgc ttc ctg caa gag tcg aat gtt ctc tat cag cac aat cta cga aga						475
Arg Phe Leu Gln Glu Ser Asn Val Leu Tyr Gln His Asn Leu Arg Arg						
	70		75		80	85
atc aag cag ttt ctt cag agc agg tat ctt gag aag cca atg gag att						523
Ile Lys Gln Phe Leu Gln Ser Arg Tyr Leu Glu Lys Pro Met Glu Ile						
	90		95		100	
gcc cgg att gtg gcc cgg tgc ctg tgg gaa gaa tca cgc ctt cta cag						571
Ala Arg Ile Val Ala Arg Cys Leu Trp Glu Glu Ser Arg Leu Leu Gln						
	105		110		115	
act gca gcc act gcg gcc cag caa ggg ggc cag gcc aac cac ccc aca						619
Thr Ala Ala Thr Ala Ala Gln Gln Gly Gly Gln Ala Asn His Pro Thr						
	120		125		130	
gca gcc gtg gtg acg gag aag cag cag atg ctg gag cag cac ctt cag						667
Ala Ala Val Val Thr Glu Lys Gln Gln Met Leu Glu Gln His Leu Gln						
	135		140		145	
gat gtc cgg aag aga gtg cag gat cta gaa cag aaa atg aaa gtg gta						715
Asp Val Arg Lys Arg Val Gln Asp Leu Glu Gln Lys Met Lys Val Val						
	150		155		160	165
gag aat ctc cag gat gac ttt gat ttc aac tat aaa acc ctc aag agt						763
Glu Asn Leu Gln Asp Asp Phe Asp Phe Asn Tyr Lys Thr Leu Lys Ser						
	170		175		180	
caa gga gac atg caa gat ctg aat gga aac aac cag tca gtg acc agg						811
Gln Gly Asp Met Gln Asp Leu Asn Gly Asn Asn Gln Ser Val Thr Arg						
	185		190		195	
cag aag atg cag cag ctg gaa cag atg ctc act gcg ctg gac cag atg						859
Gln Lys Met Gln Gln Leu Glu Gln Met Leu Thr Ala Leu Asp Gln Met						
	200		205		210	

Figure 3

egg aga agc atc gtg agt gag ctg gcg ggg ctt ttg tca gcg atg gag	907
Arg Arg Ser Ile Val Ser Glu Leu Ala Gly Leu Leu Ser Ala Met Glu	
215 220 225	
tac gtg cag aaa act ctc acg gac gag gag ctg gct gac tgg aag agg	955
Tyr Val Gln Lys Thr Leu Thr Asp Glu Glu Leu Ala Asp Trp Lys Arg	
230 235 240 245	
egg caa cag att gcc tgc att gga ggc ccg ccc aac atc tgc cta gat	1003
Arg Gln Gln Ile Ala Cys Ile Gly Gly Pro Pro Asn Ile Cys Leu Asp	
250 255 260	
egg cta gaa aac tgg ata acg tca tta gca gaa tct caa ctt cag acc	1051
Arg Leu Glu Asn Trp Ile Thr Ser Leu Ala Glu Ser Gln Leu Gln Thr	
265 270 275	
cgt caa caa att aag aaa ctg gag gag ttg cac caa aaa gtt tcc tac	1099
Arg Gln Gln Ile Lys Lys Leu Glu Glu Leu His Gln Lys Val Ser Tyr	
280 285 290	
aaa ggg gac ccc att gta cag cac ccg ccg atg ctg gag gag agg atc	1147
Lys Gly Asp Pro Ile Val Gln His Arg Pro Met Leu Glu Glu Arg Ile	
295 300 305	
gtg gag ctg ttc aga aac tta atg aaa agt gcc ttt gtg gtg gag ccg	1195
Val Glu Leu Phe Arg Asn Leu Met Lys Ser Ala Phe Val Val Glu Arg	
310 315 320 325	
cag ccc tgc atg ccc atg cat cct gac ccg ccc ctc gtc atc aag acc	1243
Gln Pro Cys Met Pro Met His Pro Asp Arg Pro Leu Val Ile Lys Thr	
330 335 340	
ggc gtc cag ttc act act aaa gtc agg ttg ctg gtc aag ttc cct gag	1291
Gly Val Gln Phe Thr Thr Lys Val Arg Leu Leu Val Lys Phe Pro Glu	
345 350 355	
ttg aat tat cag ctt aaa att aaa gtg tgc att gac aaa gac tct ggg	1339
Leu Asn Tyr Gln Leu Lys Ile Lys Val Cys Ile Asp Lys Asp Ser Gly	
360 365 370	
gac gtt gca gct ctc aga gga tcc ccg aaa ttt aac att ctg ggc aca	1387
Asp Val Ala Ala Leu Arg Gly Ser Arg Lys Phe Asn Ile Leu Gly Thr	
375 380 385	
aac aca aaa gtg atg aac atg gaa gaa tcc aac aac ggc agc ctc tct	1435
Asn Thr Lys Val Met Asn Met Glu Glu Ser Asn Asn Gly Ser Leu Ser	
390 395 400 405	
gca gaa ttc aaa cac ttg acc ctg agg gag cag aga tgt ggg aat ggg	1483
Ala Glu Phe Lys His Leu Thr Leu Arg Glu Gln Arg Cys Gly Asn Gly	
410 415 420	
ggc cga gcc aat tgt gat gct tcc ctg att gtg act gag gag ctg cac	1531
Gly Arg Ala Asn Cys Asp Ala Ser Leu Ile Val Thr Glu Glu Leu His	
425 430 435	
ctg atc acc ttt gag acc gag gtg tat cac caa ggt ctc aag att gac	1579
Leu Ile Thr Phe Glu Thr Glu Val Tyr His Gln Gly Leu Lys Ile Asp	
440 445 450	

Figure 3 (cont'd)

cta gag acc cac tcc ttg tca gtt gtg gtg atc tcc aac atc tgt cag Leu Glu Thr His Ser Leu Ser Val Val Val Ile Ser Asn Ile Cys Gln 455 460 465	1627
atg cca aat gcc tgg gcg tcc atc ctg tgg tac aac atg ctg acc aac Met Pro Asn Ala Trp Ala Ser Ile Leu Trp Tyr Asn Met Leu Thr Asn 470 475 480 485	1675
aat ccc aag aat gtg aac ttc ttc act aag ccg cca att gga acc tgg Asn Pro Lys Asn Val Asn Phe Phe Thr Lys Pro Pro Ile Gly Thr Trp 490 495 500	1723
gac caa gtg gcc gag gtg ctc agc tgg cag ttc tcg tcc acc acc aag Asp Gln Val Ala Glu Val Leu Ser Trp Gln Phe Ser Ser Thr Thr Lys 505 510 515	1771
cgg ggg ctg agc atc gag cag ctg aca acg ctg gct gag aag ctc cta Arg Gly Leu Ser Ile Glu Gln Leu Thr Thr Leu Ala Glu Lys Leu Leu 520 525 530	1819
ggg cct ggt gtg aac tac tca ggg tgt cag atc aca tgg gct aac ttc Gly Pro Gly Val Asn Tyr Ser Gly Cys Gln Ile Thr Trp Ala Asn Phe 535 540 545	1867
tgc aaa gaa aac atg gct ggc aag ggc ttc tcc tac tgg gtc tgg cta Cys Lys Glu Asn Met Ala Gly Lys Gly Phe Ser Tyr Trp Val Trp Leu 550 555 560 565	1915
gac aat atc atc gac ctt gtg aaa aag tat atc ttg gcc ctt tgg aat Asp Asn Ile Ile Asp Leu Val Lys Lys Tyr Ile Leu Ala Leu Trp Asn 570 575 580	1963
gaa ggg tac atc atg ggt ttc atc agc aag gag cgg gag cgg gcc atc Glu Gly Tyr Ile Met Gly Phe Ile Ser Lys Glu Arg Glu Arg Ala Ile 585 590 595	2011
ttg agc act aag ccc cca ggc acc ttc ctg ctg cgc ttc agt gaa agc Leu Ser Thr Lys Pro Pro Gly Thr Phe Leu Leu Arg Phe Ser Glu Ser 600 605 610	2059
agc aaa gaa gga ggc gtc act ttc act tgg gtg gag aag gac atc agc Ser Lys Glu Gly Gly Val Thr Phe Thr Trp Val Glu Lys Asp Ile Ser 615 620 625	2107
ggt aag acc cag atc cag tcc gtg gaa cca tac aca aag cag cag ctg Gly Lys Thr Gln Ile Gln Ser Val Glu Pro Tyr Thr Lys Gln Gln Leu 630 635 640 645	2155
aac aac atg tca ttt gct gaa atc atc atg ggc tat aag atc atg gat Asn Asn Met Ser Phe Ala Glu Ile Ile Met Gly Tyr Lys Ile Met Asp 650 655 660	2203
gct acc aat atc ctg ttg tct cca ctt gtc tat ctc tat cct gac att Ala Thr Asn Ile Leu Leu Ser Pro Leu Val Tyr Leu Tyr Pro Asp Ile 665 670 675	2251

Figure 3 (cont'd)

ccc aag gag gag gca ttc ggg aag tat tgt cgg cca gag agc cag gag	2299
Pro Lys Glu Glu Ala Phe Gly Lys Tyr Cys Arg Pro Glu Ser Gln Glu	
680 685 690	
cat cct gaa gct gac cca ggt agc gct gcc cca tac ctg aag acc aag	2347
His Pro Glu Ala Asp Pro Gly Ser Ala Ala Pro Tyr Leu Lys Thr Lys	
695 700 705	
ttt atc tgt gtg aca cca acg acc tgc agc aat acc att gac ctg ccg	2395
Phe Ile Cys Val Thr Pro Thr Thr Cys Ser Asn Thr Ile Asp Leu Pro	
710 715 720 725	
atg tcc ccc cgc gct tta gat tca ttg atg cag ttt gga aat aat ggt	2443
Met Ser Pro Arg Ala Leu Asp Ser Leu Met Gln Phe Gly Asn Asn Gly	
730 735 740	
gaa ggt gct gaa ccc tca gca gga ggg cag ttt gag tcc ctc acc ttt	2491
Glu Gly Ala Glu Pro Ser Ala Gly Gly Gln Phe Glu Ser Leu Thr Phe	
745 750 755	
gac atg gag ttg acc tcg gag tgc gct acc tcc ccc atg tga	2533
Asp Met Glu Leu Thr Ser Glu Cys Ala Thr Ser Pro Met *	
760 765 770	
ggagctgaga acggaagctg cagaaagata cgactgaggc gcctacctgc attctgccac	2593
ccctcacaca gccaaacccc agatcatctg aaactactaa ctttgtggtt ccagattttt	2653
tttaaatctcc tactttctgct atctttgagc aatctgggca cttttaaaaa tagagaaatg	2713
agtgaatgtg ggtgatctgc ttttatctaa atgcaaataa ggatgtgttc tctgagaccc	2773
atgatcaggg gatg	2787

Figure 3 (cont'd)

gcttttctcct agggactgtg aggggcgctt ctgactttgg aattgagcac tgcctgggac	60
ctgtgctgag agagcgctag c atg tct cag tgg aat caa gtc caa cag tta	111
Met Ser Gln Trp Asn Gln Val Gln Gln Leu	
1 5 10	
gaa atc aag ttt ttg gag cag gtg gat caa ttc tat gat gac aac ttt	159
Glu Ile Lys Phe Leu Glu Gln Val Asp Gln Phe Tyr Asp Asp Asn Phe	
15 20 25	
ccc atg gaa att cgg cat ctg ttg gcc caa tgg att gaa aat caa gac	207
Pro Met Glu Ile Arg His Leu Leu Ala Gln Trp Ile Glu Asn Gln Asp	
30 35 40	
tgg gag gca gct tct aac aat gaa acc atg gca acg att ctt ctt caa	255
Trp Glu Ala Ala Ser Asn Asn Glu Thr Met Ala Thr Ile Leu Leu Gln	
45 50 55	
aac ttg tta ata caa ctg gat gaa cag tta ggt cgt gtt tcc aaa gag	303
Asn Leu Leu Ile Gln Leu Asp Glu Gln Leu Gly Arg Val Ser Lys Glu	
60 65 70	
aaa aac cta ctc ttg ata cac aat cta aaa aga att agg aag gtc ctt	351
Lys Asn Leu Leu Leu Ile His Asn Leu Lys Arg Ile Arg Lys Val Leu	
75 80 85 90	
cag gga aaa ttt cat gga aat cca atg cat gta gct gtg gtt att tca	399
Gln Gly Lys Phe His Gly Asn Pro Met His Val Ala Val Val Ile Ser	
95 100 105	
aac tgt tta agg gaa gag agg aga ata ttg gct gca gcc aac atg cct	447
Asn Cys Leu Arg Glu Glu Arg Arg Ile Leu Ala Ala Ala Asn Met Pro	
110 115 120	
gtc cag ggg cct cta gag aaa tcc tta caa agt tct tca gtt tca gaa	495
Val Gln Gly Pro Leu Glu Lys Ser Leu Gln Ser Ser Ser Val Ser Glu	
125 130 135	
aga cag agg aat gtg gag cac aaa gtg gct gcc att aaa aac agt gtg	543
Arg Gln Arg Asn Val Glu His Lys Val Ala Ala Ile Lys Asn Ser Val	
140 145 150	
cag atg aca gaa caa gat acc aaa tac tta gaa gat ctg caa gac gaa	591
Gln Met Thr Glu Gln Asp Thr Lys Tyr Leu Glu Asp Leu Gln Asp Glu	
155 160 165 170	
ttt gac tac agg tat aaa aca att cag aca atg gat cag agt gac aag	639
Phe Asp Tyr Arg Tyr Lys Thr Ile Gln Thr Met Asp Gln Ser Asp Lys	
175 180 185	
aat agt gcc atg gtg aat cag gaa gtt ttg aca ctg cag gaa atg ctt	687
Asn Ser Ala Met Val Asn Gln Glu Val Leu Thr Leu Gln Glu Met Leu	
190 195 200	
aac agc ctc gat ttc aag aga aag gag gct ctc agt aaa atg acc caa	735
Asn Ser Leu Asp Phe Lys Arg Lys Glu Ala Leu Ser Lys Met Thr Gln	
205 210 215	

Figure 4

atc atc cat gag aca gac ctg tta atg aac acc atg ctc ata gaa gag Ile Ile His Glu Thr Asp Leu Leu Met Asn Thr Met Leu Ile Glu Glu 220 225 230	783
ctg caa gac tgg aag cgg cgg cag caa atc gcc tgc atc ggg ggt cca Leu Gln Asp Trp Lys Arg Arg Gln Gln Ile Ala Cys Ile Gly Gly Pro 235 240 245 250	831
ctc cac aat ggg ctc gac cag ctt cag aac tgc ttt aca cta ttg gca Leu His Asn Gly Leu Asp Gln Leu Gln Asn Cys Phe Thr Leu Leu Ala 255 260 265	879
gaa agt ctt ttc caa ctg aga agg caa ttg gag aaa cta gag gag caa Glu Ser Leu Phe Gln Leu Arg Arg Gln Leu Glu Lys Leu Glu Glu Gln 270 275 280	927
tct acc aaa atg aca tat gaa ggt gat ccc att cca atg caa aga act Ser Thr Lys Met Thr Tyr Glu Gly Asp Pro Ile Pro Met Gln Arg Thr 285 290 295	975
cac atg cta gaa aga gtc acc ttc ttg atc tac aac ctt ttc aag aac His Met Leu Glu Arg Val Thr Phe Leu Ile Tyr Asn Leu Phe Lys Asn 300 305 310	1023
tca ttt gtg gtt gag cga cag cca tgt atg cca acc cac cct cag agg Ser Phe Val Val Glu Arg Gln Pro Cys Met Pro Thr His Pro Gln Arg 315 320 325 330	1071
cgg ttg gta ctt aaa acc cta att cag ttc act gta aaa cta agg cta Pro Leu Val Leu Lys Thr Leu Ile Gln Phe Thr Val Lys Leu Arg Leu 335 340 345	1119
cta ata aaa ttg cca gaa cta aac tat cag gta aag gtt aag gca tca Leu Ile Lys Leu Pro Glu Leu Asn Tyr Gln Val Lys Val Lys Ala Ser 350 355 360	1167
att gac aag aat gtt tca act cta agc aac cga aga ttt gta ctt tgt Ile Asp Lys Asn Val Ser Thr Leu Ser Asn Arg Arg Phe Val Leu Cys 365 370 375	1215
gga act aat gtc aaa gcc atg tct att gaa gaa tct tcc aat ggg agt Gly Thr Asn Val Lys Ala Met Ser Ile Glu Glu Ser Ser Asn Gly Ser 380 385 390	1263
ctc tca gta gaa ttt cga cat ttg caa cca aag gaa atg aag tcc agt Leu Ser Val Glu Phe Arg His Leu Gln Pro Lys Glu Met Lys Ser Ser 395 400 405 410	1311
gct gga ggt aaa gga aat gag ggc tgt cac atg gtg act gaa gaa ctt Ala Gly Gly Lys Gly Asn Glu Gly Cys His Met Val Thr Glu Glu Leu 415 420 425	1359
cat tcc ata acg ttt gaa aca cag atc tgc ctc tat ggc ctg acc ata His Ser Ile Thr Phe Glu Thr Gln Ile Cys Leu Tyr Gly Leu Thr Ile 430 435 440	1407
gat ttg gag acc agc tca ttg cct gtg gtg atg att tcc aat gtc agt Asp Leu Glu Thr Ser Ser Leu Pro Val Val Met Ile Ser Asn Val Ser 445 450 455	1455

Figure 4 (cont'd)

cag tta cct aat gct tgg gca tcc atc att tgg tac aac gtg tca acc	1503
Gln Leu Pro Asn Ala Trp Ala Ser Ile Ile Trp Tyr Asn Val Ser Thr	
460 465 470	
aac gat tcc cag aac ttg gtt ttc ttt aat aat cct cca cct gcc aca	1551
Asn Asp Ser Gln Asn Leu Val Phe Phe Asn Asn Pro Pro Pro Ala Thr	
475 480 485 490	
ttg agt caa cta ctg gag gtg atg agc tgg cag ttt tca tcg tac gtt	1599
Leu Ser Gln Leu Leu Glu Val Met Ser Trp Gln Phe Ser Ser Tyr Val	
495 500 505	
ggt cgt ggt ctt aac tca gat caa ctc cat atg ctg gca gag aag ctt	1647
Gly Arg Gly Leu Asn Ser Asp Gln Leu His Met Leu Ala Glu Lys Leu	
510 515 520	
aca gtc caa tct agc tac agt gat ggt cac ctc acc tgg gcc aag ttc	1695
Thr Val Gln Ser Ser Tyr Ser Asp Gly His Leu Thr Trp Ala Lys Phe	
525 530 535	
tgc aag gaa cat tta cct ggt aaa tca ttt acc ttt tgg aca tgg ctt	1743
Cys Lys Glu His Leu Pro Gly Lys Ser Phe Thr Phe Trp Thr Trp Leu	
540 545 550	
gaa gca ata ttg gat cta att aag aaa cac att ctt ccc ctt tgg att	1791
Glu Ala Ile Leu Asp Leu Ile Lys Lys His Ile Leu Pro Leu Trp Ile	
555 560 565 570	
gat ggg tat gtc atg ggc ttt gtt agc aaa gag aag gaa cgg ctg ttg	1839
Asp Gly Tyr Val Met Gly Phe Val Ser Lys Glu Lys Glu Arg Leu Leu	
575 580 585	
cta aag gat aaa atg cct ggc acc ttt tta tta aga ttc agt gaa agc	1887
Leu Lys Asp Lys Met Pro Gly Thr Phe Leu Leu Arg Phe Ser Glu Ser	
590 595 600	
cat ctc gga gga ata act ttc acc tgg gtg gac cat tct gaa agt ggg	1935
His Leu Gly Gly Ile Thr Phe Thr Trp Val Asp His Ser Glu Ser Gly	
605 610 615	
gaa gtg aga ttc cac tct gta gaa ccc tac aat aaa ggc cgg ttg tct	1983
Glu Val Arg Phe His Ser Val Glu Pro Tyr Asn Lys Gly Arg Leu Ser	
620 625 630	
gct ctg cca ttc gct gac atc ctg cga gac tac aaa gtt att atg gct	2031
Ala Leu Pro Phe Ala Asp Ile Leu Arg Asp Tyr Lys Val Ile Met Ala	
635 640 645 650	
gaa aac att cct gaa aac cct ctg aag tac cta tat cct gac att ccc	2079
Glu Asn Ile Pro Glu Asn Pro Leu Lys Tyr Leu Tyr Pro Asp Ile Pro	
655 660 665	
aaa gac aaa gcc ttc ggt aaa cac tac agc tct cag cct tgc gaa gtt	2127
Lys Asp Lys Ala Phe Gly Lys His Tyr Ser Ser Gln Pro Cys Glu Val	
670 675 680	

Figure 4 (cont'd)

tca	aga	cca	aca	gaa	agg	ggg	gac	aaa	ggg	tat	gtt	cct	tct	gtt	ttt	2175
Ser	Arg	Pro	Thr	Glu	Arg	Gly	Asp	Lys	Gly	Tyr	Val	Pro	Ser	Val	Phe	
		685					690					695				
atc	ccc	atc	tca	aca	atc	cga	agt	gat	tca	aca	gag	cca	cat	tct	cca	2223
Ile	Pro	Ile	Ser	Thr	Ile	Arg	Ser	Asp	Ser	Thr	Glu	Pro	His	Ser	Pro	
	700					705					710					
tca	gac	ctt	ctt	ccc	atg	tct	cca	agt	gtg	tat	gcg	gtg	ttg	aga	gaa	2271
Ser	Asp	Leu	Leu	Pro	Met	Ser	Pro	Ser	Val	Tyr	Ala	Val	Leu	Arg	Glu	
	715				720					725					730	
aac	ctg	agt	ccc	aca	aca	att	gaa	act	gca	atg	aag	tct	cct	tat	tct	2319
Asn	Leu	Ser	Pro	Thr	Thr	Ile	Glu	Thr	Ala	Met	Lys	Ser	Pro	Tyr	Ser	
				735					740					745		
gct	gaa	tga	caggataaac	tctgacgcac	caagaaagga	agcaaatagaa										2368
Ala	Glu	*														
aaagtttaaa	gactgttctt	tgcccaataa	ccacatttta	tttcttcagc	tttgtaaata											2428
ccaggttcta	ggaaatgttt	gacatctgaa	gctctcttca	cactcccggtg	gcactcctca											2488
attgggagtg	ttgtgactga	aatgcttgaa	accaaagctt	cagataaaact	tgcaagataa											2548
gacaacttta	agaaaccagt	gttaataaca	atattaacag													2588

Figure 4 (cont'd)

atcttatttt	tctttttggt	ggtggtggtg	gaagggggga	ggtgctagca	gggccagcct	60
tgaactcgct	ggacagagct	acagacctat	ggggcctgga	agtgcctcgct	gagaaagggg	120
gaagacagca	gaggggttgc	cgaggcaacc	tccaagtccc	agatc atg tct ctg tgg		177
				Met Ser Leu Trp		
				1		
ggt ctg gtc tcc aag atg ccc cca gaa aaa gtg cag cgg ctc tat gtc						225
Gly Leu Val Ser Lys Met Pro Pro Glu Lys Val Gln Arg Leu Tyr Val						
5 10 15 20						
gac ttt ccc caa cac ctg cgg cat ctt ctg ggt gac tgg ctg gag agc						273
Asp Phe Pro Gln His Leu Arg His Leu Leu Gly Asp Trp Leu Glu Ser						
25 30 35						
cag ccc tgg gag ttc ctg gtc ggc tcc gac gcc ttc tgc tgc aac ttg						321
Gln Pro Trp Glu Phe Leu Val Gly Ser Asp Ala Phe Cys Cys Asn Leu						
40 45 50						
gct agt gcc cta ctt tca gac act gtc cag cac ctt cag gcc tcg gtg						369
Ala Ser Ala Leu Leu Ser Asp Thr Val Gln His Leu Gln Ala Ser Val						
55 60 65						
gga gag cag ggg gag ggg agc acc atc ttg caa cac atc agc acc ctt						417
Gly Glu Gln Gly Glu Gly Ser Thr Ile Leu Gln His Ile Ser Thr Leu						
70 75 80						
gag agc ata tat cag agg gac ccc ctg aag ctg gtg gcc act ttc aga						465
Glu Ser Ile Tyr Gln Arg Asp Pro Leu Lys Leu Val Ala Thr Phe Arg						
85 90 95 100						
caa ata ctt caa gga gag aaa aaa gct gtt atg gaa cag ttc cgc cac						513
Gln Ile Leu Gln Gly Glu Lys Lys Ala Val Met Glu Gln Phe Arg His						
105 110 115						
ttg cca atg cct ttc cac tgg aag cag gaa gaa ctc aag ttt aag aca						561
Leu Pro Met Pro Phe His Trp Lys Gln Glu Glu Leu Lys Phe Lys Thr						
120 125 130						
ggc ttg cgg agg ctg cag cac cga gta ggg gag atc cac ctt ctc cga						609
Gly Leu Arg Arg Leu Gln His Arg Val Gly Glu Ile His Leu Leu Arg						
135 140 145						
gaa gcc ctg cag aag ggg gct gag gct ggc caa gtg tct ctg cac agc						657
Glu Ala Leu Gln Lys Gly Ala Glu Ala Gly Gln Val Ser Leu His Ser						
150 155 160						
ttg ata gaa act cct gct aat ggg act ggg cca agt gag gcc ctg gcc						705
Leu Ile Glu Thr Pro Ala Asn Gly Thr Gly Pro Ser Glu Ala Leu Ala						
165 170 175 180						
atg cta ctg cag gag acc act gga gag cta gag gca gcc aaa gcc cta						753
Met Leu Leu Gln Glu Thr Thr Gly Glu Leu Glu Ala Ala Lys Ala Leu						
185 190 195						
gtg ctg aag agg atc cag att tgg aaa cgg cag cag cag ctg gca ggg						801
Val Leu Lys Arg Ile Gln Ile Trp Lys Arg Gln Gln Gln Leu Ala Gly						
200 205 210						

Figure 5

aat ggc gca ccg ttt gag gag agc ctg gcc cca ctc cag gag agg tgt Asn Gly Ala Pro Phe Glu Glu Ser Leu Ala Pro Leu Gln Glu Arg Cys 215 220 225	849
gaa agc ctg gtg gac att tat tcc cag cta cag cag gag gta ggg gcg Glu Ser Leu Val Asp Ile Tyr Ser Gln Leu Gln Gln Glu Val Gly Ala 230 235 240	897
gct ggt ggg gag ctt gag ccc aag acc cgg gca tcg ctg act ggc cgg Ala Gly Gly Glu Leu Glu Pro Lys Thr Arg Ala Ser Leu Thr Gly Arg 245 250 255 260	945
ctg gat gaa gtc ctg aga acc ctc gtc acc agt tgc ttc ctg gtg gag Leu Asp Glu Val Leu Arg Thr Leu Val Thr Ser Cys Phe Leu Val Glu 265 270 275	993
aag cag ccc ccc cag gta ctg aag act cag acc aag ttc cag gct gga Lys Gln Pro Pro Gln Val Leu Lys Thr Gln Thr Lys Phe Gln Ala Gly 280 285 290	1041
gtt cga ttc ctg ttg ggc ttg agg ttc ctg ggg gcc cca gcc aag cct Val Arg Phe Leu Leu Gly Leu Arg Phe Leu Gly Ala Pro Ala Lys Pro 295 300 305	1089
ccg ctg gtc agg gcc gac atg gtg aca gag aag cag gcg cgg gag ctg Pro Leu Val Arg Ala Asp Met Val Thr Glu Lys Gln Ala Arg Glu Leu 310 315 320	1137
agt gtg cct cag ggt cct ggg gct gga gca gaa agc act gga gaa atc Ser Val Pro Gln Gly Pro Gly Ala Gly Ala Glu Ser Thr Gly Glu Ile 325 330 335 340	1185
atc aac aac act gtg ccc ttg gag aac agc att cct ggg aac tgc tgc Ile Asn Asn Thr Val Pro Leu Glu Asn Ser Ile Pro Gly Asn Cys Cys 345 350 355	1233
tct gcc ctg ttc aag aac ctg ctt ctc aag aag atc aag cgg tgt gag Ser Ala Leu Phe Lys Asn Leu Leu Leu Lys Lys Ile Lys Arg Cys Glu 360 365 370	1281
cgg aag ggc act gag tct gtc aca gag gag aag tgc gct gtg ctc ttc Arg Lys Gly Thr Glu Ser Val Thr Glu Glu Lys Cys Ala Val Leu Phe 375 380 385	1329
tct gcc agc ttc aca ctt ggc ccc ggc aaa ctc ccc atc cag ctc cag Ser Ala Ser Phe Thr Leu Gly Pro Gly Lys Leu Pro Ile Gln Leu Gln 390 395 400	1377
gcc ctg tct ctg ccc ctg gtg gtc atc gtc cat ggc aac caa gac aac Ala Leu Ser Leu Pro Leu Val Val Ile Val His Gly Asn Gln Asp Asn 405 410 415 420	1425
aat gcc aaa gcc act atc ctg tgg gac aat gcc ttc tct gag atg gac Asn Ala Lys Ala Thr Ile Leu Trp Asp Asn Ala Phe Ser Glu Met Asp 425 430 435	1473
cgc gtg ccc ttt gtg gtg gct gag cgg gtg ccc tgg gag aag atg tgt Arg Val Pro Phe Val Val Ala Glu Arg Val Pro Trp Glu Lys Met Cys 440 445 450	1521

Figure 5 (cont'd)

gaa act ctg aac ctg aag ttc atg gct gag gtg ggg acc aac cgg ggg Glu Thr Leu Asn Leu Lys Phe Met Ala Glu Val Gly Thr Asn Arg Gly 455 460 465	1569
ctg ctc cca gag cac ttc ctc ttc ctg gcc cag aag atc ttc aat gac Leu Leu Pro Glu His Phe Leu Phe Leu Ala Gln Lys Ile Phe Asn Asp 470 475 480	1617
aac agc ctc agt atg gag gcc ttc cag cac cgt tct gtg tcc tgg tcg Asn Ser Leu Ser Met Glu Ala Phe Gln His Arg Ser Val Ser Trp Ser 485 490 495 500	1665
cag ttc aac aag gag atc ctg ctg ggc cgt ggc ttc acc ttt tgg cag Gln Phe Asn Lys Glu Ile Leu Leu Gly Arg Gly Phe Thr Phe Trp Gln 505 510 515	1713
tgg ttt gat ggt gtc ctg gac ctc acc aaa cgc tgt ctc cgg agc tac Trp Phe Asp Gly Val Leu Asp Leu Thr Lys Arg Cys Leu Arg Ser Tyr 520 525 530	1761
tgg tct gac cgg ctg atc att ggc ttc atc agc aaa cag tac gtt act Trp Ser Asp Arg Leu Ile Ile Gly Phe Ile Ser Lys Gln Tyr Val Thr 535 540 545	1809
agc ctt ctt ctc aat gag ccc gac gga acc ttt ctc ctc cgc ttc agc Ser Leu Leu Leu Asn Glu Pro Asp Gly Thr Phe Leu Leu Arg Phe Ser 550 555 560	1857
gac tca gag att ggg ggc atc acc att gcc cat gtc atc cgg ggc cag Asp Ser Glu Ile Gly Gly Ile Thr Ile Ala His Val Ile Arg Gly Gln 565 570 575 580	1905
gat ggc tct cca cag ata gag aac atc cag cca ttc tct gcc aaa gac Asp Gly Ser Pro Gln Ile Glu Asn Ile Gln Pro Phe Ser Ala Lys Asp 585 590 595	1953
ctg tcc att cgc tca ctg ggg gac cga atc cgg gat ctt gct cag ctc Leu Ser Ile Arg Ser Leu Gly Asp Arg Ile Arg Asp Leu Ala Gln Leu 600 605 610	2001
aaa aat ctc tat ccc aag aag ccc aag gat gag gct ttc cgg agc cac Lys Asn Leu Tyr Pro Lys Lys Pro Lys Asp Glu Ala Phe Arg Ser His 615 620 625	2049
tac aag cct gaa cag atg ggt aag gat ggc agg ggt tat gtc cca gct Tyr Lys Pro Glu Gln Met Gly Lys Asp Gly Arg Gly Tyr Val Pro Ala 630 635 640	2097
acc atc aag atg acc gtg gaa agg gac caa cca ctt cct acc cca gag Thr Ile Lys Met Thr Val Glu Arg Asp Gln Pro Leu Pro Thr Pro Glu 645 650 655 660	2145
ctc cag atg cct acc atg gtg cct tct tat gac ctt gga atg gcc cct Leu Gln Met Pro Thr Met Val Pro Ser Tyr Asp Leu Gly Met Ala Pro 665 670 675	2193
gat tcc tcc atg agc atg cag ctt ggc cca gat atg gtg ccc cag gtg Asp Ser Ser Met Ser Met Gln Leu Gly Pro Asp Met Val Pro Gln Val 680 685 690	2241

Figure 5 (cont'd)

tac cca cca cac tct cac tcc atc ccc ccg tat caa ggc ctc tcc cca	2289
Tyr Pro Pro His Ser His Ser Ile Pro Pro Tyr Gln Gly Leu Ser Pro	
695 700 705	
gaa gaa tca gtc aac gtg ttg tca gcc ttc cag gag cct cac ctg cag	2337
Glu Glu Ser Val Asn Val Leu Ser Ala Phe Gln Glu Pro His Leu Gln	
710 715 720	
atg ccc ccc agc ctg ggc cag atg agc ctg ccc ttt gac cag cct cac	2385
Met Pro Pro Ser Leu Gly Gln Met Ser Leu Pro Phe Asp Gln Pro His	
725 730 735 740	
ccc cag ggc ctg ctg ccg tgc cag cct cag gag cat gct gtg tcc agc	2433
Pro Gln Gly Leu Leu Pro Cys Gln Pro Gln Glu His Ala Val Ser Ser	
745 750 755	
cct gac ccc ctg ctc tgc tca gat gtg acc atg gtg gaa gac agc tgc	2481
Pro Asp Pro Leu Leu Cys Ser Asp Val Thr Met Val Glu Asp Ser Cys	
760 765 770	
ctg agc cag cca gtg aca gcg ttt cct cag ggc act tgg att ggt gaa	2529
Leu Ser Gln Pro Val Thr Ala Phe Pro Gln Gly Thr Trp Ile Gly Glu	
775 780 785	
gac ata ttc cct cct ctg ctg cct ccc act gaa cag gac ctc act aag	2577
Asp Ile Phe Pro Pro Leu Leu Pro Pro Thr Glu Gln Asp Leu Thr Lys	
790 795 800	
ctt ctc ctg gag ggg caa ggg gag tgc ggg gga ggg tcc ttg ggg gca	2625
Leu Leu Leu Glu Gly Gln Gly Glu Ser Gly Gly Gly Ser Leu Gly Ala	
805 810 815 820	
cag ccc ctc ctg cag ccc tcc cac tat ggg caa tct ggg atc tca atg	2673
Gln Pro Leu Leu Gln Pro Ser His Tyr Gly Gln Ser Gly Ile Ser Met	
825 830 835	
tcc cac atg gac cta agg gcc aac ccc agt tgg tga tcccagctgg	2719
Ser His Met Asp Leu Arg Ala Asn Pro Ser Trp *	
840 845	
agggagaacc caaagagaca gctctttctac tacccccaca gacctgctct ggacaattgc	2779
tcatgccctg ccaagcagca gatggggagg gtgccctcct atccccacct actcctgggt	2839
caggaggaaa agactaacag gagaatgcac agtgggtgga gccaatccac tccttccttt	2899
ctatcattcc cctgcccacc tccttcacagc actgactgga agggaagtgc aggctctgag	2959
acacgcccac acatgcctgc acctgcagcg cgcacacgca cgcacacaca catacagagc	3019
tcctctgaggg tgatggggct gagcagg	3046

Figure 5 (cont'd)

cccccttctgt agg atg gta gca cac aac cag gtg gca gcc gac aat gca	49
Met Val Ala His Asn Gln Val Ala Ala Asp Asn Ala	
1 5 10	
gtc tcc aca gca gca gag ccc cga cgg cgg cca gaa cct tcc tcc tct	97
Val Ser Thr Ala Ala Glu Pro Arg Arg Arg Pro Glu Pro Ser Ser Ser	
15 20 25	
tcc tcc tcc tcc ccc gcg gcc ccc gcg cgc ccg cgg ccg tgc ccc gcg	145
Ser Ser Ser Ser Pro Ala Ala Pro Ala Arg Pro Arg Pro Cys Pro Ala	
30 35 40	
gtc ccg gcc ccg gcc ccc ggc gac acg cac ttc cgc aca ttc cgt tcc	193
Val Pro Ala Pro Ala Pro Gly Asp Thr His Phe Arg Thr Phe Arg Ser	
45 50 55 60	
cac gcc gat tac cgg cgc atc acg cgc gcc agc gcg ctc ctg gac gcc	241
His Ala Asp Tyr Arg Arg Ile Thr Arg Ala Ser Ala Leu Leu Asp Ala	
65 70 75	
tgc gga ttc tac tgg ggg ccc ctg agc gtg cac ggg gcg cac gag cgg	289
Cys Gly Phe Tyr Trp Gly Pro Leu Ser Val His Gly Ala His Glu Arg	
80 85 90	
ctg cgc gcc gag ccc gtg ggc acc ttc ctg gtg cgc gac agc cgc cag	337
Leu Arg Ala Glu Pro Val Gly Thr Phe Leu Val Arg Asp Ser Arg Gln	
95 100 105	
cgg aac tgc ttt ttc gcc ctt agc gtg aag atg gcc tcc gga ccc acg	385
Arg Asn Cys Phe Phe Ala Leu Ser Val Lys Met Ala Ser Gly Pro Thr	
110 115 120	
agc atc cgc gtg cac ttt cag gcc ggc cgc ttt cac ctg gat ggc agc	433
Ser Ile Arg Val His Phe Gln Ala Gly Arg Phe His Leu Asp Gly Ser	
125 130 135 140	
cgc gag agc ttc gac tgc ctc ttc gag ctg ctg gag cac tac gtg gcg	481
Arg Glu Ser Phe Asp Cys Leu Phe Glu Leu Leu Glu His Tyr Val Ala	
145 150 155	
gcg ccg cgc cgc atg ctg ggg gcc ccg ctg cgc cag cgc cgc gtg cgg	529
Ala Pro Arg Arg Met Leu Gly Ala Pro Leu Arg Gln Arg Arg Val Arg	
160 165 170	
ccg ctg cag gag ctg tgc cgc cag cgc atc gtg gcc acc gtg ggc cgc	577
Pro Leu Gln Glu Leu Cys Arg Gln Arg Ile Val Ala Thr Val Gly Arg	
175 180 185	
gag aac ctg gct cgc atc ccc ctc aac ccc gtc ctc cgc gac tac ctg	625
Glu Asn Leu Ala Arg Ile Pro Leu Asn Pro Val Leu Arg Asp Tyr Leu	
190 195 200	
agc tcc ttc ccc ttc cag att tga ccggcagcgc ccgcogtgca cgcagcatta	679
Ser Ser Phe Pro Phe Gln Ile *	
205 210	
actgggatgc cgtgttattt tgttattact tgcctggaac catgtgggta ccctccccgg	739
cctgggttgg agggagcgga tgggtgtagg ggcgaggcgc ctcccgccct cggctggaga	799
cgaggccgca gaccccttct cactcttga ggggtcctc cccctcctgg tgctccctct	859
gggtccccct ggttggttga gcagcttaac tgtatctgga gccaggacc	908

Figure 6

atg gtc acc cac agc aag ttt ccc gcc gcc ggg atg agc cgc ccc ctg	48
Met Val Thr His Ser Lys Phe Pro Ala Ala Gly Met Ser Arg Pro Leu	
1 5 10 15	
gac acc agc ctg cgc ctc aag acc ttc agc tcc aag agc gag tac cag	96
Asp Thr Ser Leu Arg Leu Lys Thr Phe Ser Ser Lys Ser Glu Tyr Gln	
20 25 30	
ctg gtg gtg aac gca gtg cgc aag ctg cag gag agc ggc ttc tac tgg	144
Leu Val Val Asn Ala Val Arg Lys Leu Gln Glu Ser Gly Phe Tyr Trp	
35 40 45	
agc gca gtg acc ggc ggc gag ggc aac ctg ctg ctc agc gcc gag ccc	192
Ser Ala Val Thr Gly Gly Glu Ala Asn Leu Leu Leu Ser Ala Glu Pro	
50 55 60	
gcc ggc acc ttt ctg atc cgc gac agc tgc gac cag cgc cac ttc ttc	240
Ala Gly Thr Phe Leu Ile Arg Asp Ser Ser Asp Gln Arg His Phe Phe	
65 70 75 80	
acg ctc agc gtc aag acc cag tct ggg acc aag aac ctg cgc atc cag	288
Thr Leu Ser Val Lys Thr Gln Ser Gly Thr Lys Asn Leu Arg Ile Gln	
85 90 95	
tgt gag ggg ggc agc ttc tct ctg cag agc gat ccc cgg agc acg cag	336
Cys Glu Gly Gly Ser Phe Ser Leu Gln Ser Asp Pro Arg Ser Thr Gln	
100 105 110	
ccc gtg ccc cgc ttc gac tgc gtg ctc aag ctg gtg cac cac tac atg	384
Pro Val Pro Arg Phe Asp Cys Val Leu Lys Leu Val His His Tyr Met	
115 120 125	
ccg ccc cct gga gcc ccc tcc ttc ccc tgc cca cct act gaa ccc tcc	432
Pro Pro Pro Gly Ala Pro Ser Phe Pro Ser Pro Pro Thr Glu Pro Ser	
130 135 140	
tcc gag gtg ccc gag cag ccg tct gcc cag cca ctc cct ggg agt ccc	480
Ser Glu Val Pro Glu Gln Pro Ser Ala Gln Pro Leu Pro Gly Ser Pro	
145 150 155 160	
ccc aga aga gcc tat tac atc tac tcc ggg ggc gag aag atc ccc ctg	528
Pro Arg Arg Ala Tyr Tyr Ile Tyr Ser Gly Gly Glu Lys Ile Pro Leu	
165 170 175	
gtg ttg agc cgg ccc ctc tcc tcc aac gtg gcc act ctt cag cat ctc	576
Val Leu Ser Arg Pro Leu Ser Ser Asn Val Ala Thr Leu Gln His Leu	
180 185 190	
tgt cgg aag acc gtc aac ggc cac ctg gac tcc tat gag aaa gtc acc	624
Cys Arg Lys Thr Val Asn Gly His Leu Asp Ser Tyr Glu Lys Val Thr	
195 200 205	
cag ctg ccg ggg ccc att cgg gag ttc ctg gac cag tac gat gcc ccg	672
Gln Leu Pro Gly Pro Ile Arg Glu Phe Leu Asp Gln Tyr Asp Ala Pro	
210 215 220	
ctt taa gggg	682
Leu *	
225	

Figure 7

aattcggcac	gagggggggc	agcagcggac	gocgctaacg	gcctccctcg	gcgctgacag	60
gctggggccg	cgcccggctc	gcttgggtgt	tgcgctcgcc	acttcggctt	ctcggccggt	120
cgggccctc	ggcccgggct	tgcggcgcgc	gtcggggctg	agggctgctg	cggcgcaggg	180
agaggcctgg	tcctcgctgc	cgagggatgt	gagtgggagc	tgagcccaca	ctggagggcc	240
cccaggggcc	cagcctggag	gtcgttcaga	gccgtgcccg	ccccggggct	tgcagacct	300
tgacccgcg	ggtaggagcc	gccctgcgg	gctcgagggc	gcgctctggt	gcgccgatct	360
gtgtagccgg	tttcagaagc	aggcaacagg	aacaagatgt	gaactgtttc	tcttctgcag	420
aaaaagaggc	tcttcctcct	cctcccgcga	cggcaaattgt	tctgaaaaag	actctgc atg	480
					Met	
					1	
gga atg gcc	tgc ctt acg	atg aca gaa	atg gag gga	aca tcc acc	tct	528
Gly Met Ala	Cys Leu Thr	Met Thr Glu	Met Glu Gly	Thr Ser Thr	Ser	
	5		10		15	
tct ata tat	cag aat ggt	gat att tct	gga aat gcc	aat tct atg	aag	576
Ser Ile Tyr	Gln Asn Gly	Asp Ile Ser	Gly Asn Ala	Asn Ser Met	Lys	
	20		25		30	
caa ata gat	cca gtt ctt	cag gtg tat	ctt tac cat	tcc ctt ggg	aaa	624
Gln Ile Asp	Pro Val Leu	Gln Val Tyr	Leu Tyr His	Ser Leu Gly	Lys	
	35		40		45	
tct gag gca	gat tat ctg	acc ttt cca	tct ggg gag	tat gtt gca	gaa	672
Ser Glu Ala	Asp Tyr Leu	Thr Phe Pro	Ser Gly Glu	Tyr Val Ala	Glu	
	50		55		60	65
gaa atc tgt	att gct gct	tct aaa gct	tgt ggt atc	aca cct gtg	tat	720
Glu Ile Cys	Ile Ala Ala	Ser Lys Ala	Cys Gly Ile	Thr Pro Val	Tyr	
	70		75		80	
cat aat atg	ttt gct tta	atg agt gaa	aca gaa agg	atc tgg tat	cca	768
His Asn Met	Phe Ala Leu	Met Ser Glu	Thr Glu Arg	Ile Trp Tyr	Pro	
	85		90		95	
ccc aac cat	gtc ttc cat	ata gat gag	tca acc agg	cat aat gta	ctc	816
Pro Asn His	Val Phe His	Ile Asp Glu	Ser Thr Arg	His Asn Val	Leu	
	100		105		110	
tac aga ata	aga ttt tac	ttt cct cgt	tgg tat tgc	agt ggc agc	aac	864
Tyr Arg Ile	Arg Phe Tyr	Phe Pro Arg	Trp Tyr Cys	Ser Gly Ser	Asn	
	115		120		125	
aga gcc tat	cgg cat gga	ata tct cga	ggt gct gaa	gct cct ctt	ctt	912
Arg Ala Tyr	Arg His Gly	Ile Ser Arg	Gly Ala Glu	Ala Pro Leu	Leu	
	130		135		140	145
gat gac ttt	gtc atg tct	tac ctg ttt	gct cag tgg	cgg cat gat	ttt	960
Asp Asp Phe	Val Met Ser	Tyr Leu Phe	Ala Gln Trp	Arg His Asp	Phe	
	150		155		160	
gtg cac gga	tgg ata aaa	gta cct gtg	act cat gaa	aca cag gaa	gaa	1008
Val His Gly	Trp Ile Lys	Val Pro Val	Thr His Glu	Thr Gln Glu	Glu	
	165		170		175	
tgt ctt ggg	atg gca gtg	tta gat atg	atg aga ata	gcc aaa gaa	aac	1056
Cys Leu Gly	Met Ala Val	Leu Asp Met	Met Arg Ile	Ala Lys Glu	Asn	
	180		185		190	

Figure 8

gat	caa	acc	cca	ctg	gcc	atc	tat	aac	tct	atc	agc	tac	aag	aca	ttc	1104
Asp	Gln	Thr	Pro	Leu	Ala	Ile	Tyr	Asn	Ser	Ile	Ser	Tyr	Lys	Thr	Phe	
	195					200					205					
tta	cca	aaa	tgt	att	cga	gca	aag	atc	caa	gac	tat	cat	att	ttg	aca	1152
Leu	Pro	Lys	Cys	Ile	Arg	Ala	Lys	Ile	Gln	Asp	Tyr	His	Ile	Leu	Thr	
210					215					220					225	
agg	aag	cga	ata	agg	tac	aga	ttt	cgc	aga	ttt	att	cag	caa	ttc	agc	1200
Arg	Lys	Arg	Ile	Arg	Tyr	Arg	Phe	Arg	Arg	Phe	Ile	Gln	Gln	Phe	Ser	
				230					235					240		
caa	tgc	aaa	gcc	act	gcc	aga	aac	ttg	aaa	ctt	aag	tat	ctt	ata	aat	1248
Gln	Cys	Lys	Ala	Thr	Ala	Arg	Asn	Leu	Lys	Leu	Lys	Tyr	Leu	Ile	Asn	
			245					250					255			
ctg	gaa	act	ctg	cag	tct	gcc	ttc	tac	aca	gag	aaa	ttt	gaa	gta	aaa	1296
Leu	Glu	Thr	Leu	Gln	Ser	Ala	Phe	Tyr	Thr	Glu	Lys	Phe	Glu	Val	Lys	
		260					265					270				
gaa	cct	gga	agt	ggt	cct	tca	ggt	gag	gag	att	ttt	gca	acc	att	ata	1344
Glu	Pro	Gly	Ser	Gly	Pro	Ser	Gly	Glu	Glu	Ile	Phe	Ala	Thr	Ile	Ile	
	275					280					285					
ata	act	gga	aac	ggt	gga	att	cag	tgg	tca	aga	ggg	aaa	cat	aaa	gaa	1392
Ile	Thr	Gly	Asn	Gly	Gly	Ile	Gln	Trp	Ser	Arg	Gly	Lys	His	Lys	Glu	
290					295					300					305	
agt	gag	aca	ctg	aca	gaa	cag	gat	tta	cag	tta	tat	tgc	gat	ttt	tct	1440
Ser	Glu	Thr	Leu	Thr	Glu	Gln	Asp	Leu	Gln	Leu	Tyr	Cys	Asp	Phe	Ser	
				310					315					320		
aat	att	att	gat	gtc	agt	att	aag	caa	gca	aac	caa	gag	ggt	tca	aat	1488
Asn	Ile	Ile	Asp	Val	Ser	Ile	Lys	Gln	Ala	Asn	Gln	Glu	Gly	Ser	Asn	
			325					330					335			
gaa	agc	cga	gtt	gta	act	atc	cat	aag	caa	gat	ggt	aaa	aat	ctg	gaa	1536
Glu	Ser	Arg	Val	Val	Thr	Ile	His	Lys	Gln	Asp	Gly	Lys	Asn	Leu	Glu	
		340					345					350				
att	gaa	ctt	agc	tca	tta	agg	gaa	gct	ttg	tct	ttc	gtg	tca	tta	att	1584
Ile	Glu	Leu	Ser	Ser	Leu	Arg	Glu	Ala	Leu	Ser	Phe	Val	Ser	Leu	Ile	
	355					360					365					
gat	gga	tat	tat	aga	tta	act	gca	gat	gca	cat	cat	tac	ctc	tgt	aaa	1632
Asp	Gly	Tyr	Tyr	Arg	Leu	Thr	Ala	Asp	Ala	His	His	Tyr	Leu	Cys	Lys	
370					375					380					385	
gaa	gta	gca	cct	cca	gcc	gtg	ctt	gaa	aat	ata	caa	agc	aac	tgt	cat	1680
Glu	Val	Ala	Pro	Pro	Ala	Val	Leu	Glu	Asn	Ile	Gln	Ser	Asn	Cys	His	
				390					395					400		
ggc	cca	att	tcg	atg	gat	ttt	gcc	att	agt	aaa	ctg	aag	aaa	gca	ggt	1728
Gly	Pro	Ile	Ser	Met	Asp	Phe	Ala	Ile	Ser	Lys	Leu	Lys	Lys	Ala	Gly	
			405				410						415			
aat	cag	act	gga	ctg	tat	gta	ctt	cga	tgc	agt	cct	aag	gac	ttt	aat	1776
Asn	Gln	Thr	Gly	Leu	Tyr	Val	Leu	Arg	Cys	Ser	Pro	Lys	Asp	Phe	Asn	
		420					425					430				

Figure 8 (cont'd)

aaa tat ttt ttg act ttt gct gtc gag cga gaa aat gtc att gaa tat	1824
Lys Tyr Phe Leu Thr Phe Ala Val Glu Arg Glu Asn Val Ile Glu Tyr	
435 440 445	
aaa cac tgt ttg att aca aaa aat gag aat gaa gag tac aac ctc agt	1872
Lys His Cys Leu Ile Thr Lys Asn Glu Asn Glu Glu Tyr Asn Leu Ser	
450 455 460 465	
ggg aca aag aag aac ttc agc agt ctt aaa gat ctt ttg aat tgt tac	1920
Gly Thr Lys Lys Asn Phe Ser Ser Leu Lys Asp Leu Leu Asn Cys Tyr	
470 475 480	
cag atg gaa act gtt cgc tca gac aat ata att ttc cag ttt act aaa	1968
Gln Met Glu Thr Val Arg Ser Asp Asn Ile Ile Phe Gln Phe Thr Lys	
485 490 495	
tgc tgt ccc cca aag cca aaa gat aaa tca aac ctt cta gtc ttc aga	2016
Cys Cys Pro Pro Lys Pro Lys Asp Lys Ser Asn Leu Leu Val Phe Arg	
500 505 510	
acg aat ggt gtt tct gat gta cca acc tca cca aca tta cag agg cct	2064
Thr Asn Gly Val Ser Asp Val Pro Thr Ser Pro Thr Leu Gln Arg Pro	
515 520 525	
act cat atg aac caa atg gtg ttt cac aaa atc aga aat gaa gat ttg	2112
Thr His Met Asn Gln Met Val Phe His Lys Ile Arg Asn Glu Asp Leu	
530 535 540 545	
ata ttt aat gaa agc ctt ggc caa ggc act ttt aca aag att ttt aaa	2160
Ile Phe Asn Glu Ser Leu Gly Gln Gly Thr Phe Thr Lys Ile Phe Lys	
550 555 560	
ggc gta cga aga gaa gta gga gac tac ggt caa ctg cat gaa aca gaa	2208
Gly Val Arg Arg Glu Val Gly Asp Tyr Gly Gln Leu His Glu Thr Glu	
565 570 575	
gtt ctt tta aaa gtt ctg gat aaa gca cac aga aac tat tca gag tct	2256
Val Leu Leu Lys Val Leu Asp Lys Ala His Arg Asn Tyr Ser Glu Ser	
580 585 590	
ttc ttt gaa gca gca agt atg atg agc aag ctt tct cac aag cat ttg	2304
Phe Phe Glu Ala Ala Ser Met Met Ser Lys Leu Ser His Lys His Leu	
595 600 605	
gtt tta aat tat gga gta tgt gtc tgt gga gac gag aat att ctg gtt	2352
Val Leu Asn Tyr Gly Val Cys Val Cys Gly Asp Glu Asn Ile Leu Val	
610 615 620 625	
cag gag ttt gta aaa ttt gga tca cta gat aca tat ctg aaa aag aat	2400
Gln Glu Phe Val Lys Phe Gly Ser Leu Asp Thr Tyr Leu Lys Lys Asn	
630 635 640	
aaa aat tgt ata aat ata tta tgg aaa ctt gaa gtt gct aaa cag ttg	2448
Lys Asn Cys Ile Asn Ile Leu Trp Lys Leu Glu Val Ala Lys Gln Leu	
645 650 655	

Figure 8 (cont'd)

gca tgg gcc atg cat ttt cta gaa gaa aac acc ctt att cat ggg aat Ala Trp Ala Met His Phe Leu Glu Glu Asn Thr Leu Ile His Gly Asn 660 665 670	2496
gta tgt gcc aaa aat att ctg ctt atc aga gaa gaa gac agg aag aca Val Cys Ala Lys Asn Ile Leu Leu Ile Arg Glu Glu Asp Arg Lys Thr 675 680 685	2544
gga aat cct cct ttc atc aaa ctt agt gat cct ggc att agt att aca Gly Asn Pro Pro Phe Ile Lys Leu Ser Asp Pro Gly Ile Ser Ile Thr 690 695 700 705	2592
gtt ttg cca aag gac att ctt cag gag aga ata cca tgg gta cca cct Val Leu Pro Lys Asp Ile Leu Gln Glu Arg Ile Pro Trp Val Pro Pro 710 715 720	2640
gaa tgc att gaa aat cct aaa aat tta aat ttg gca aca gac aaa tgg Glu Cys Ile Glu Asn Pro Lys Asn Leu Asn Leu Ala Thr Asp Lys Trp 725 730 735	2688
agt ttt ggt acc act ttg tgg gaa atc tgc agt gga gga gat aaa cct Ser Phe Gly Thr Thr Leu Trp Glu Ile Cys Ser Gly Gly Asp Lys Pro 740 745 750	2736
cta agt gct ctg gat tct caa aga aag cta caa ttt tat gaa gat agg Leu Ser Ala Leu Asp Ser Gln Arg Lys Leu Gln Phe Tyr Glu Asp Arg 755 760 765	2784
cat cag ctt cct gca cca aag tgg gca gaa tta gca aac ctt ata aat His Gln Leu Pro Ala Pro Lys Trp Ala Glu Leu Ala Asn Leu Ile Asn 770 775 780 785	2832
aat tgt atg gat tat gaa cca gat ttc agg cct tct ttc aga gcc atc Asn Cys Met Asp Tyr Glu Pro Asp Phe Arg Pro Ser Phe Arg Ala Ile 790 795 800	2880
ata cga gat ctt aac agt ttg ttt act cca gat tat gaa cta tta aca Ile Arg Asp Leu Asn Ser Leu Phe Thr Pro Asp Tyr Glu Leu Leu Thr 805 810 815	2928
gaa aat gac atg tta cca aat atg agg ata ggt gcc ctg ggg ttt tct Glu Asn Asp Met Leu Pro Asn Met Arg Ile Gly Ala Leu Gly Phe Ser 820 825 830	2976
ggt gcc ttt gaa gac cgg gat cct aca cag ttt gaa gag aga cat ttg Gly Ala Phe Glu Asp Arg Asp Pro Thr Gln Phe Glu Glu Arg His Leu 835 840 845	3024
aaa ttt cta cag caa ctt ggc aag ggt aat ttt ggg agt gtg gag atg Lys Phe Leu Gln Gln Leu Gly Lys Gly Asn Phe Gly Ser Val Glu Met 850 855 860 865	3072
tgc cgg tat gac cct cta cag gac aac act ggg gag gtg gtc gct gta Cys Arg Tyr Asp Pro Leu Gln Asp Asn Thr Gly Glu Val Val Ala Val 870 875 880	3120
aaa aag ctt cag cat agt act gaa gag cac cta aga gac ttt gaa agg Lys Lys Leu Gln His Ser Thr Glu Glu His Leu Arg Asp Phe Glu Arg 885 890 895	3168

Figure 8 (cont'd)

gaa att gaa atc ctg aaa tcc cta cag cat gac aac att gta aag tac	3216
Glu Ile Glu Ile Leu Lys Ser Leu Gln His Asp Asn Ile Val Lys Tyr	
900 905 910	
aag gga gtg tgc tac agt gct ggt cgg cgt aat cta aaa tta att atg	3264
Lys Gly Val Cys Tyr Ser Ala Gly Arg Arg Asn Leu Lys Leu Ile Met	
915 920 925	
gaa tat tta cca tat gga agt tta cga gac tat ctt caa aaa cat aaa	3312
Glu Tyr Leu Pro Tyr Gly Ser Leu Arg Asp Tyr Leu Gln Lys His Lys	
930 935 940 945	
gaa cgg ata gat cac ata aaa ctt ctg cag tac aca tct cag ata tgc	3360
Glu Arg Ile Asp His Ile Lys Leu Leu Gln Tyr Thr Ser Gln Ile Cys	
950 955 960	
aag ggt atg gag tat ctt ggt aca aaa agg tat atc cac agg gat ctg	3408
Lys Gly Met Glu Tyr Leu Gly Thr Lys Arg Tyr Ile His Arg Asp Leu	
965 970 975	
gca acg aga aat ata ttg gtg gag aac gag aac aga gtt aaa att gga	3456
Ala Thr Arg Asn Ile Leu Val Glu Asn Glu Asn Arg Val Lys Ile Gly	
980 985 990	
gat ttt ggg tta acc aaa gtc ttg cca caa gac aaa gaa tac tat aaa	3504
Asp Phe Gly Leu Thr Lys Val Leu Pro Gln Asp Lys Glu Tyr Tyr Lys	
995 1000 1005	
gta aaa gaa cct ggt gaa agt ccc ata ttc tgg tat gct cca gaa tca	3552
Val Lys Glu Pro Gly Glu Ser Pro Ile Phe Trp Tyr Ala Pro Glu Ser	
1010 1015 1020 1025	
ctg aca gag agc aag ttt tct gtg gcc tca gat gtt tgg agc ttt gga	3600
Leu Thr Glu Ser Lys Phe Ser Val Ala Ser Asp Val Trp Ser Phe Gly	
1030 1035 1040	
gtg gtt ctg tat gaa ctt ttc aca tac att gag aag agt aaa agt cca	3648
Val Val Leu Tyr Glu Leu Phe Thr Tyr Ile Glu Lys Ser Lys Ser Pro	
1045 1050 1055	
cca gcg gaa ttt atg cgt atg att ggc aat gac aaa caa gga cag atg	3696
Pro Ala Glu Phe Met Arg Met Ile Gly Asn Asp Lys Gln Gly Gln Met	
1060 1065 1070	
atc gtg ttc cat ttg ata gaa ctt ttg aag aat aat gga aga tta cca	3744
Ile Val Phe His Leu Ile Glu Leu Leu Lys Asn Asn Gly Arg Leu Pro	
1075 1080 1085	
aga cca gat gga tgc cca gat gag atc tat atg atc atg aca gaa tgc	3792
Arg Pro Asp Gly Cys Pro Asp Glu Ile Tyr Met Ile Met Thr Glu Cys	
1090 1095 1100 1105	
tgg aac aat aat gta aat caa cgc ccc tcc ttt agg gat cta gct ctt	3840
Trp Asn Asn Asn Val Asn Gln Arg Pro Ser Phe Arg Asp Leu Ala Leu	
1110 1115 1120	
cga gtg gat caa ata agg gat aac atg gct gga tga aagaaatgac	3886
Arg Val Asp Gln Ile Arg Asp Asn Met Ala Gly *	
1125 1130	

Figure 8 (cont'd)

cttcattctg	agaccaaagt	agatttacag	aacaaagttt	tatatattcac	attgctgtgg	3946
actattatta	catatatcat	tattatataa	atcatgatgc	tagccagcaa	agatgtgaaa	4006
atatctgctc	aaaactttca	aagtttagta	agtttttctt	catgaggcca	ccagtaaaag	4066
acattaatga	gaattcctta	gcaaggattt	tgtaagaagt	ttcttaaaca	ttgtcagtta	4126
acatcactct	tgtctggcaa	aagaaaaaaa	atagactttt	tcaactcagc	tttttgagac	4186
ctgaaaaaat	tattatgtaa	attttgcaat	gttaaagatg	cacagaatat	gtatgtatag	4246
tttttaccac	agtggatgta	taataccttg	gcatcttggt	tgatgtttta	cacacatgag	4306
ggctggtggt	cattaatact	gttttcta	ttttccatag	ttaatctata	attaattact	4366
tcactataca	aacaaattaa	gatgttcaga	taattgaata	agtacctttg	tgctcctgtt	4426
catttatatc	gctggccagc	attataagca	ggtgtatact	tttagcttgt	agttccatgt	4486
actgtaaata	tttttcacat	aaagggaaca	aatgtctagt	tttatttgta	taggaaattt	4546
ccctgaccct	aaataataca	ttttgaaatg	aaacaagctt	acaaagatat	aatctatttt	4606
attatgggtt	cccttgatc	tatttggtgt	gaatgtgttt	tttaaagga	actatctcca	4666
aatttttcta	agactactat	gaacagtttt	cttttaaaat	tttgagatta	agaatgccag	4726
gaatattgtc	atcctttgag	ctgctgactg	ccaataacat	tcttcgatct	ctgggattta	4786
tgctcatgaa	ctaaatttaa	gcttaagcca	taaaatagat	tagattgttt	tttaaaaatg	4846
gatagctcat	taagaagtgc	agcagggttaa	gaatttttct	ctaaagactg	tatatattgag	4906
gggtttcaga	attttgcatt	gcagtcatag	aagagattta	tttccttttt	agaggggaaa	4966
tgaggtaa	aagtataaaa	gtatgcttgt	taattttatt	caagaatgcc	agtagaaaat	5026
tcataacgtg	tatctttaag	aaaaatgagc	atacatctta	aatcttttca	attaaggtcg	5086
acgcggccgc	ggtcgacgcg	gccgcgaatt	c			5117

Figure 8 (cont'd)

gacgcgggcg	cggaaggagc	gcggccggag	gtcctcagga	agaagccgcg	gggactggct	60
gcgcttgaca	ggctgcactt	ggatggggagc	acctggtgcc	tcgggaactgc	tccgatgccc	120
gggtctgtgc	tgaatgtgta	atatgcggaa	ctatattgaa	acattacaac	catcttttga	180
tggcaacacc	ctgaggacct	cccttttcca	gatggggaaa	ctgaggccca	gaattgctaa	240
gtggcttgct	tgagttgaca	cagggagctc	caggactcac	cctcagctga	gccacctgcc	300
gggagc atg	cct ctg cgc	cac tgg ggg	atg gcc agg	ggc agt aag	ccc	348
Met	Pro	Leu	Arg	His	Trp	Gly
1			5			10
ggt ggg gat	gga gcc cag	ccc atg gct	gcc atg gga	ggc ctg aag	gtg	396
Val Gly Asp	Gly Ala Gln	Pro Met Ala	Ala Ala Met	Gly Gly Leu	Lys Val	
15		20		25		30
ctt ctg cac	tgg gct ggt	cca ggc ggc	ggg gag ccc	tgg gtc act	ttc	444
Leu Leu His	Trp Ala Gly	Pro Gly Gly	Gly Glu Pro	Trp Val Thr	Phe	
	35		40		45	
agt gag tca	tcg ctg aca	gct gag gaa	gtc tgc atc	cac att gca	cat	492
Ser Glu Ser	Ser Leu Thr	Ala Glu Glu	Val Cys Ile	His Ile Ala	His	
	50		55		60	
aaa gtt ggt	atc act cct	cct tgc ttc	aat ctc ttt	gcc ctc ttc	gat	540
Lys Val Gly	Ile Thr Pro	Pro Cys Phe	Asn Leu Phe	Ala Leu Phe	Asp	
	65		70		75	
gct cag gcc	caa gtc tgg	ttg ccc cca	aac cac atc	cta gag atc	ccc	588
Ala Gln Ala	Gln Val Trp	Leu Pro Pro	Asn His Ile	Leu Glu Ile	Pro	
	80		85		90	
aga gat gca	agc ctg atg	cta tat ttc	cgc ata agg	ttt tat ttc	cgg	636
Arg Asp Ala	Ser Leu Met	Leu Tyr Phe	Arg Ile Arg	Phe Tyr Phe	Arg	
	95		100		105	
aac tgg cat	ggc atg aat	cct cgg gaa	ccg gct gtg	tac cgt tgt	ggg	684
Asn Trp His	Gly Met Asn	Pro Arg Glu	Pro Ala Val	Tyr Arg Cys	Gly	
	115		120		125	
ccc cca gga	acc gag gca	tcc tca gat	cag aca gca	cag ggg atg	caa	732
Pro Pro Gly	Thr Glu Ala	Ser Ser Asp	Gln Thr Ala	Gln Gly Met	Gln	
	130		135		140	
ctc ctg gac	cca gcc tca	ttt gag tac	ctc ttt gag	cag ggc aag	cat	780
Leu Leu Asp	Pro Ala Ser	Phe Glu Tyr	Leu Phe Glu	Gln Gly Lys	His	
	145		150		155	
gag ttt gtg	aat gac gtg	gcat tca ctg	tgg gag ctg	tcg acc gag	gag	828
Glu Phe Val	Asn Asp Val	Ala Ser Leu	Trp Glu Leu	Ser Thr Glu	Glu	
	160		165		170	
gag atc cac	cac ttt aag	aat gag agc	ctg ggc atg	gcc ttt ctg	cac	876
Glu Ile His	His Phe Lys	Asn Glu Ser	Leu Gly Met	Ala Phe Leu	His	
	175		180		185	
ctc tgt cac	ctc gct ctc	cgc cat ggc	atc ccc ctg	gag gag gtg	gcc	924
Leu Cys His	Leu Ala Leu	Arg His Gly	Ile Pro Leu	Glu Glu Val	Ala	
	195		200		205	
aag aag acc	agc ttc aag	gac tgc atc	ccg cgc tcc	ttc cgc cgg	cat	972
Lys Lys Thr	Ser Phe Lys	Asp Cys Ile	Pro Arg Ser	Phe Arg Arg	His	
	210		215		220	

Figure 9

atc cgg cag cac agc gcc ctg acc cgg ctg cgc ctt cgg aac gtc ttc Ile Arg Gln His Ser Ala Leu Thr Arg Leu Arg Leu Arg Asn Val Phe 225 230 235	1020
cgc agg ttc ctg cgg gac ttc cag ccg ggc cga ctc tcc cag cag atg Arg Arg Phe Leu Arg Asp Phe Gln Pro Gly Arg Leu Ser Gln Gln Met 240 245 250	1068
gtc atg gtc aaa tac cta gcc aca ctc gag cgg ctg gca ccc cgc ttc Val Met Val Lys Tyr Leu Ala Thr Leu Glu Arg Leu Ala Pro Arg Phe 255 260 265 270	1116
ggc aca gag cgt gtg ccc gtg tgc cac ctg agg ctg ctg gcc cag gcc Gly Thr Glu Arg Val Pro Val Cys His Leu Arg Leu Leu Ala Gln Ala 275 280 285	1164
gag ggg gag ccc tgc tac atc cgg gac agt ggg gtg gcc cct aca gac Glu Gly Glu Pro Cys Tyr Ile Arg Asp Ser Gly Val Ala Pro Thr Asp 290 295 300	1212
cct ggc cct gag tct gct gct ggg ccc cca acc cac gag gtg ctg gtg Pro Gly Pro Glu Ser Ala Ala Gly Pro Pro Thr His Glu Val Leu Val 305 310 315	1260
aca ggc act ggt ggc atc cag tgg tgg cca gta gag gag gag gtg aac Thr Gly Thr Gly Gly Ile Gln Trp Trp Pro Val Glu Glu Glu Val Asn 320 325 330	1308
aag gag gag ggt tct agt ggc agc agt ggc agg aac ccc caa gcc agc Lys Glu Glu Gly Ser Ser Gly Ser Ser Gly Arg Asn Pro Gln Ala Ser 335 340 345 350	1356
ctg ttt ggg aag aag gcc aag gct cac aag gca ttc ggc cag ccg gca Leu Phe Gly Lys Lys Ala Lys Ala His Lys Ala Phe Gly Gln Pro Ala 355 360 365	1404
gac agg ccg cgg gag cca ctg tgg gcc tac ttc tgt gac ttc cgg gac Asp Arg Pro Arg Glu Pro Leu Trp Ala Tyr Phe Cys Asp Phe Arg Asp 370 375 380	1452
atc acc cac gtg gtg ctg aaa gag cac tgt gtc agc atc cac cgg cag Ile Thr His Val Val Leu Lys Glu His Cys Val Ser Ile His Arg Gln 385 390 395	1500
gac aac aag tgc ctg gag ctg agc ttg cct tcc cgg gct gcg gcg ctg Asp Asn Lys Cys Leu Glu Leu Ser Leu Pro Ser Arg Ala Ala Ala Leu 400 405 410	1548
tcc ttc gtg tcg ctg gtg gac ggc tat ttc cgc ctg acg gcc gac tcc Ser Phe Val Ser Leu Val Asp Gly Tyr Phe Arg Leu Thr Ala Asp Ser 415 420 425 430	1596
agc cac tac ctg tgc cac gag gtg gct ccc cca cgg ctg gtg atg agc Ser His Tyr Leu Cys His Glu Val Ala Pro Pro Arg Leu Val Met Ser 435 440 445	1644
atc cgg gat ggg atc cac gga ccc ctg ctg gag cca ttt gtg cag gcc Ile Arg Asp Gly Ile His Gly Pro Leu Leu Glu Pro Phe Val Gln Ala 450 455 460	1692

Figure 9 (cont'd)

aag ctg cgg ccc gag gac ggc ctg tac ctc att cac tgg agc acc agc	1740
Lys Leu Arg Pro Glu Asp Gly Leu Tyr Leu Ile His Trp Ser Thr Ser	
465 470 475	
cac ccc tac cgc ctg atc ctc aca gtg gcc cag cgt agc cag gca cca	1788
His Pro Tyr Arg Leu Ile Leu Thr Val Ala Gln Arg Ser Gln Ala Pro	
480 485 490	
gac ggc atg cag agc ttg cgg ctc cga aag ttc ccc att gag cag cag	1836
Asp Gly Met Gln Ser Leu Arg Leu Arg Lys Phe Pro Ile Glu Gln Gln	
495 500 505 510	
gac ggg gcc ttc gtg ctg gag ggc tgg ggc cgg tcc ttc ccc agc gtt	1884
Asp Gly Ala Phe Val Leu Glu Gly Trp Gly Arg Ser Phe Pro Ser Val	
515 520 525	
cgg gaa ctt ggg gct gcc ttg cag ggc tgc ttg ctg agg gcc ggg gat	1932
Arg Glu Leu Gly Ala Ala Leu Gln Gly Cys Leu Leu Arg Ala Gly Asp	
530 535 540	
gac tgc ttc tct ctg cgt cgc tgt tgc ctg ccc caa cca gga gaa acc	1980
Asp Cys Phe Ser Leu Arg Arg Cys Cys Leu Pro Gln Pro Gly Glu Thr	
545 550 555	
tcc aat ctc atc atc atg cgg ggg gct cgg gcc agc ccc agg aca ctc	2028
Ser Asn Leu Ile Ile Met Arg Gly Ala Arg Ala Ser Pro Arg Thr Leu	
560 565 570	
aac ctc agc cag ctc agc ttc cac cgg gtt gac cag aag gag atc acc	2076
Asn Leu Ser Gln Leu Ser Phe His Arg Val Asp Gln Lys Glu Ile Thr	
575 580 585 590	
cag ctg tcc cac ttg ggc cag ggc aca agg acc aac gtg tat gag ggc	2124
Gln Leu Ser His Leu Gly Gln Gly Thr Arg Thr Asn Val Tyr Glu Gly	
595 600 605	
cgc ctg cga gtg gag ggc agc ggg gac cct gag gag ggc aag atg gat	2172
Arg Leu Arg Val Glu Gly Ser Gly Asp Pro Glu Glu Gly Lys Met Asp	
610 615 620	
gac gag gac ccc ctc gtg cct ggc agg gac cgt ggg cag gag cta cga	2220
Asp Glu Asp Pro Leu Val Pro Gly Arg Asp Arg Gly Gln Glu Leu Arg	
625 630 635	
gtg gtg ctc aaa gtg ctg gac cct agt cac cat gac atc gcc ctg gcc	2268
Val Val Leu Lys Val Leu Asp Pro Ser His His Asp Ile Ala Leu Ala	
640 645 650	
ttc tac gag aca gcc agc ctc atg agc cag gtc tcc cac acg cac ctg	2316
Phe Tyr Glu Thr Ala Ser Leu Met Ser Gln Val Ser His Thr His Leu	
655 660 665 670	
gcc ttc gtg cat ggc gtc tgt gtg cgc ggc cct gaa aat agc atg gtg	2364
Ala Phe Val His Gly Val Cys Val Arg Gly Pro Glu Asn Ser Met Val	
675 680 685	
aca gag tac gtg gag cac gga ccc ctg gat gtg tgg ctg cgg agg gag	2412
Thr Glu Tyr Val Glu His Gly Pro Leu Asp Val Trp Leu Arg Arg Glu	
690 695 700	

Figure 9 (cont'd)

cgg ggc cat gtg ccc atg gct tgg aag atg gtg gtg gcc cag cag ctg	2460
Arg Gly His Val Pro Met Ala Trp Lys Met Val Val Ala Gln Gln Leu	
705 710 715	
gcc agc gcc ctc agc tac ctg gag aac aag aac ctg gtt cat ggt aat	2508
Ala Ser Ala Leu Ser Tyr Leu Glu Asn Lys Asn Leu Val His Gly Asn	
720 725 730	
gtg tgt ggc cgg aac atc ctg ctg gcc cgg ctg ggg ttg gca gag ggc	2556
Val Cys Gly Arg Asn Ile Leu Leu Ala Arg Leu Gly Leu Ala Glu Gly	
735 740 745 750	
acc agc ccc ttc atc aag ctg agt gat cct ggc gtg ggc ctg ggc gcc	2604
Thr Ser Pro Phe Ile Lys Leu Ser Asp Pro Gly Val Gly Leu Gly Ala	
755 760 765	
ctc tcc agg gag gag cgg gtg gag agg atc ccc tgg ctg gcc ccc gaa	2652
Leu Ser Arg Glu Glu Arg Val Glu Arg Ile Pro Trp Leu Ala Pro Glu	
770 775 780	
tgc cta cca ggt ggg gcc aac agc cta agc acc gcc atg gac aag tgg	2700
Cys Leu Pro Gly Gly Ala Asn Ser Leu Ser Thr Ala Met Asp Lys Trp	
785 790 795	
ggg ttt ggc gcc acc ctc ctg gag atc tgc ttt gac gga gag gcc cct	2748
Gly Phe Gly Ala Thr Leu Leu Glu Ile Cys Phe Asp Gly Glu Ala Pro	
800 805 810	
ctg cag agc cgc agt ccc tcc gag aag gag cat ttc tac cag agg cag	2796
Leu Gln Ser Arg Ser Pro Ser Glu Lys Glu His Phe Tyr Gln Arg Gln	
815 820 825 830	
cac cgg ctg ccc gag ccc tcc tgc cca cag ctg gcc aca ctc acc agc	2844
His Arg Leu Pro Glu Pro Ser Cys Pro Gln Leu Ala Thr Leu Thr Ser	
835 840 845	
cag tgt ctg acc tat gag cca acc cag agg cca tca ttc cgc acc atc	2892
Gln Cys Leu Thr Tyr Glu Pro Thr Gln Arg Pro Ser Phe Arg Thr Ile	
850 855 860	
ctg cgt gac ctc acc cgc gtg cag ccc cac aat ctt gct gac gtc ttg	2940
Leu Arg Asp Leu Thr Arg Val Gln Pro His Asn Leu Ala Asp Val Leu	
865 870 875	
act gtg aac cgg gac tca ccg gcc gtc gga cct act act ttc cac aag	2988
Thr Val Asn Arg Asp Ser Pro Ala Val Gly Pro Thr Thr Phe His Lys	
880 885 890	
cgc tat ttg aaa aag atc cga gat ctg ggc gag ggt cac ttc ggc aag	3036
Arg Tyr Leu Lys Lys Ile Arg Asp Leu Gly Glu Gly His Phe Gly Lys	
895 900 905 910	
gtc agc ttg tac tgc tac gat ccg acc aac gac ggc act ggc gag atg	3084
Val Ser Leu Tyr Cys Tyr Asp Pro Thr Asn Asp Gly Thr Gly Glu Met	
915 920 925	
gtg gcg gtg aaa gcc ctc aag gca gac tgc ggc ccc cag cac cgc tgc	3132
Val Ala Val Lys Ala Leu Lys Ala Asp Cys Gly Pro Gln His Arg Ser	
930 935 940	

Figure 9 (cont'd)

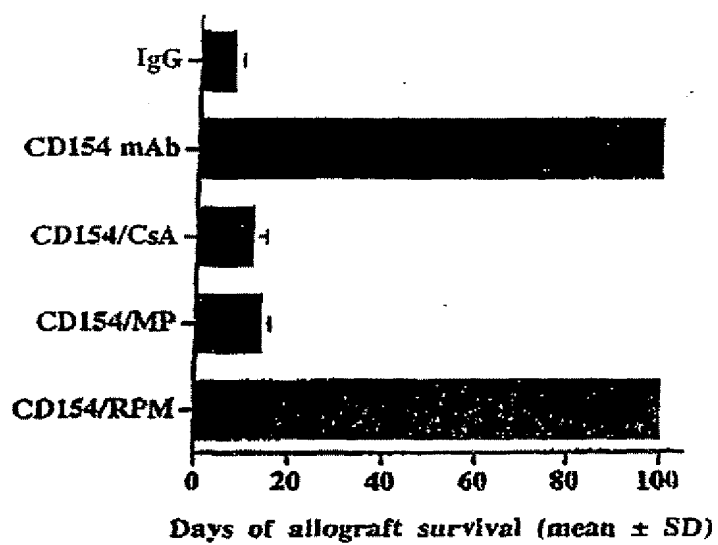
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cag ctg gtc atg gag tac gtg ccc ctg ggc agc ctc cga gac tac ctg Gln Leu Val Met Glu Tyr Val Pro Leu Gly Ser Leu Arg Asp Tyr Leu 975 980 985 990	3276
ccc cgg cac agc atc ggg ctg gcc cag ctg ctg ctc ttc gcc cag cag Pro Arg His Ser Ile Gly Leu Ala Gln Leu Leu Leu Phe Ala Gln Gln 995 1000 1005	3324
atc tgc gag ggc atg gcc tat ctg cac gcg cac gac tac atc cac cga Ile Cys Glu Gly Met Ala Tyr Leu His Ala His Asp Tyr Ile His Arg 1010 1015 1020	3372
gac cta gcc gcg cgc aac gtg ctg ctg gac aac gac agg ctg gtc aag Asp Leu Ala Ala Arg Asn Val Leu Leu Asp Asn Asp Arg Leu Val Lys 1025 1030 1035	3420
atc ggg gac ttt ggc cta gcc aag gcc gtg ccc gaa ggc cac gag tac Ile Gly Asp Phe Gly Leu Ala Lys Ala Val Pro Glu Gly His Glu Tyr 1040 1045 1050	3468
tac cgc gtg cgc gag gat ggg gac agc ccc gtg ttc tgg tat gcc cca Tyr Arg Val Arg Glu Asp Gly Asp Ser Pro Val Phe Trp Tyr Ala Pro 1055 1060 1065 1070	3516
gag tgc ctg aag gag tat aag ttc tac tat gcg tca gat gtc tgg tcc Glu Cys Leu Lys Glu Tyr Lys Phe Tyr Tyr Ala Ser Asp Val Trp Ser 1075 1080 1085	3564
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atg aca gtt ctg aga ctc act gag ttg ctg gaa cga ggg gag agg ctg Met Thr Val Leu Arg Leu Thr Glu Leu Leu Glu Arg Gly Glu Arg Leu 1120 1125 1130	3708
cca cgg ccc gac aaa tgt ccc tgt gag gtc tat cat ctc atg aag aac Pro Arg Pro Asp Lys Cys Pro Cys Glu Val Tyr His Leu Met Lys Asn 1135 1140 1145 1150	3756
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Figure 9 (cont'd)

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ggggatgtca gcctcaccca caccgtgtgc cttactcctg tctagagacc ccacctctgt      4020
gaacttattt ttctttcttg gccgtgagcc taaccatgat cttgaggagc ccaacatttg      4080
taggggcact aatccagccc ttaaataccc cagcttccaa acttgaggcc caccatctcc      4140
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Figure 9 (cont'd)

**Figure 10**

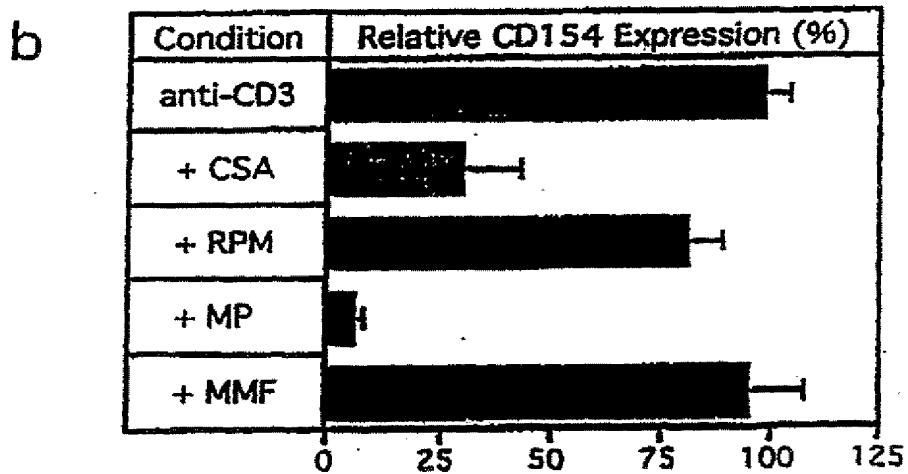
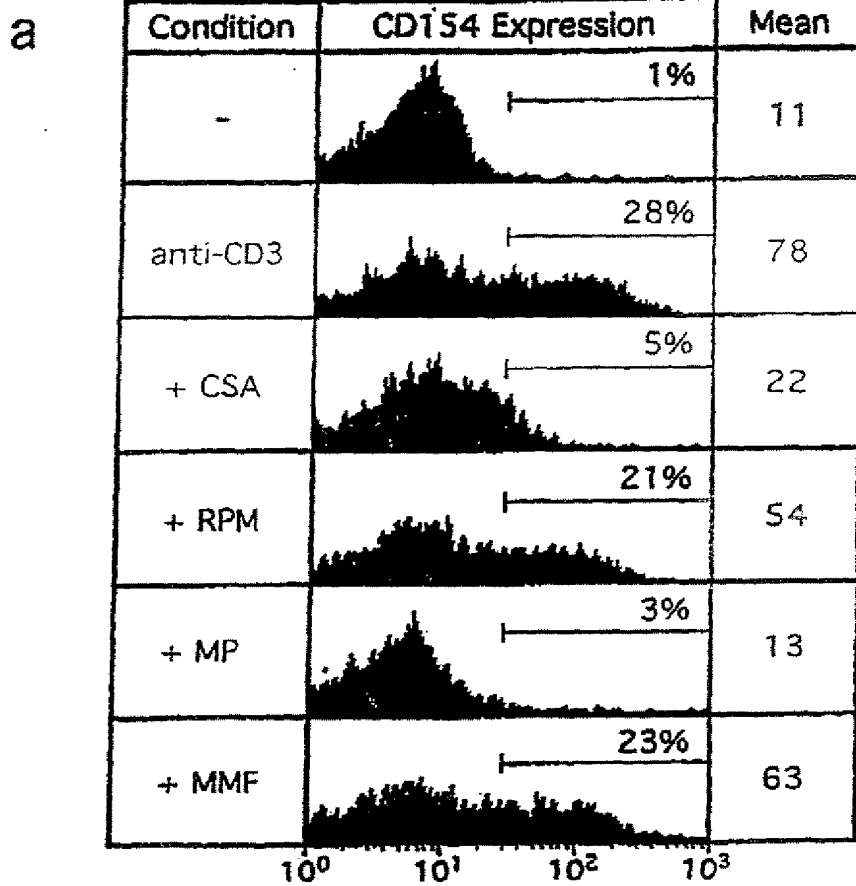
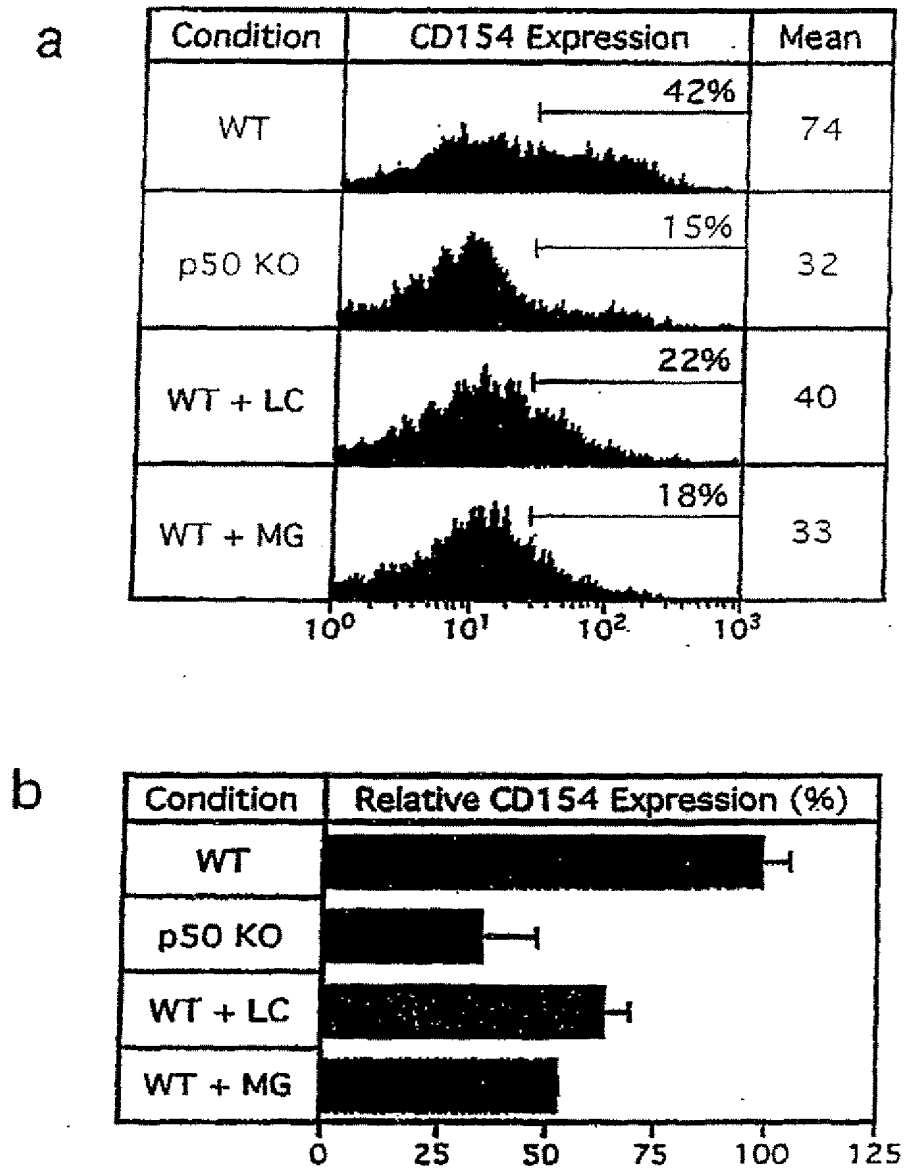
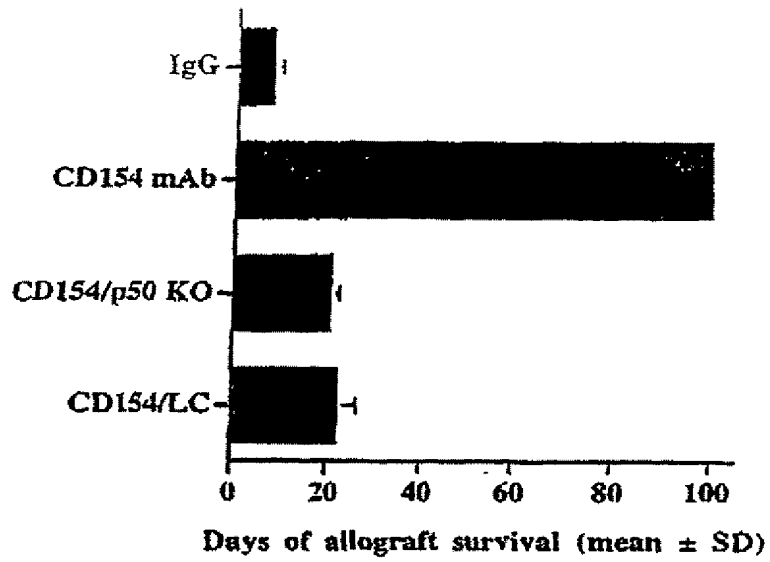


Figure 11

**Figure 12**

**Figure 13**

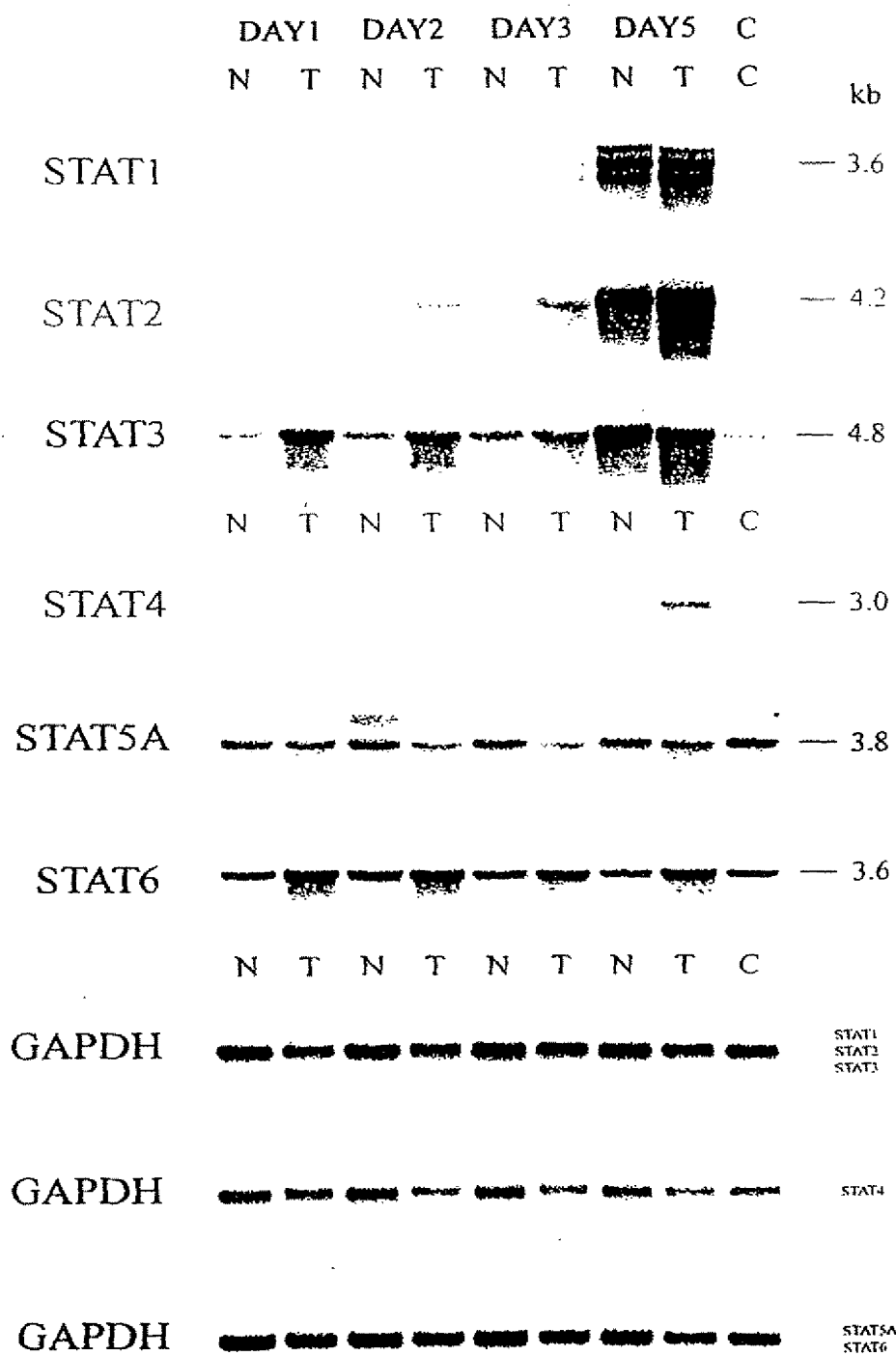
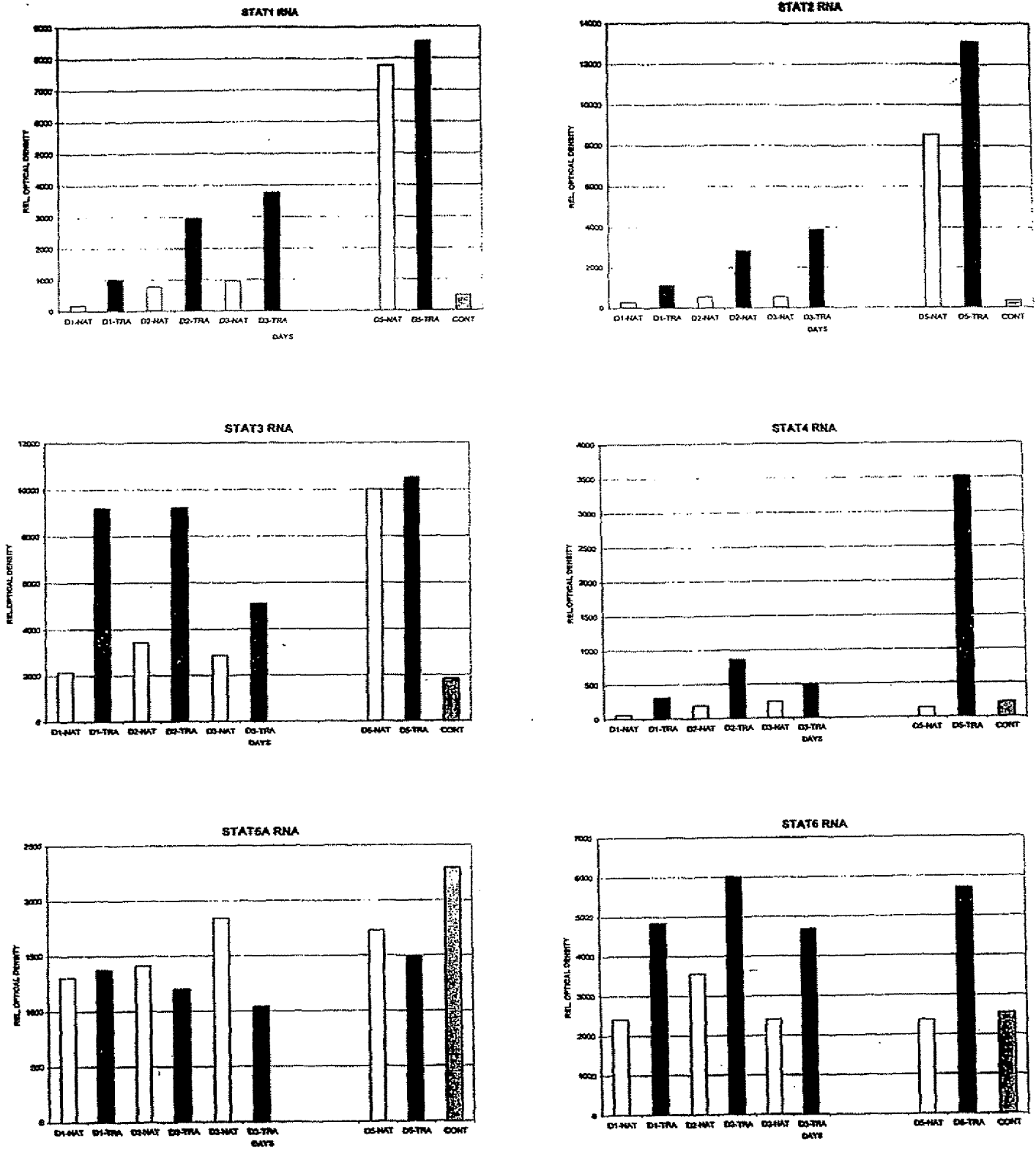


Figure 14A



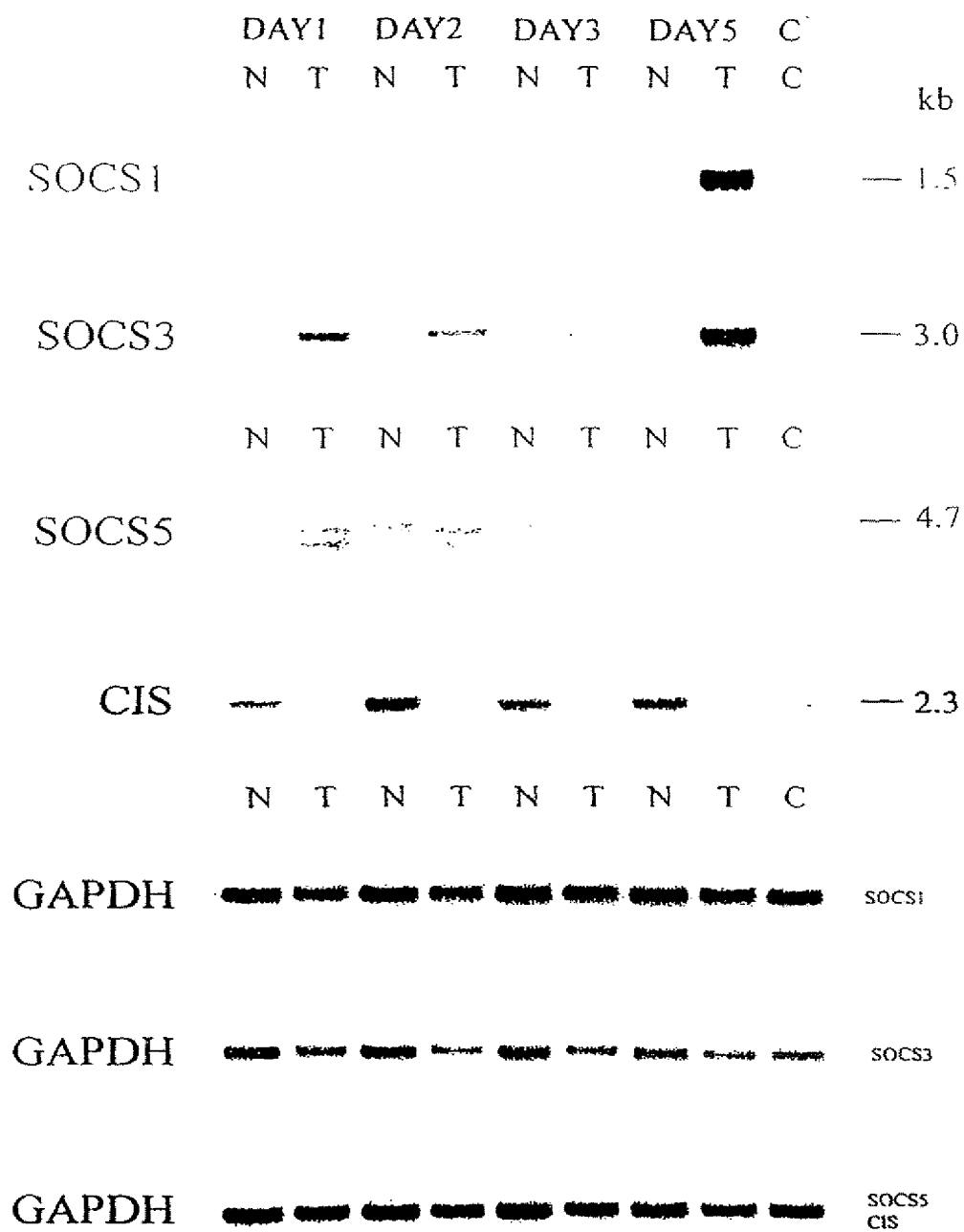
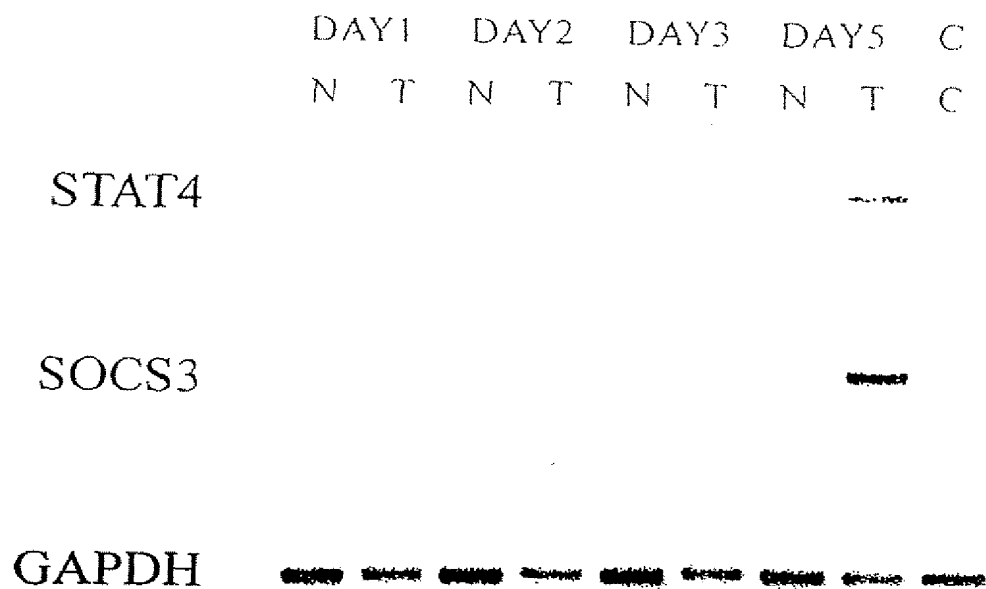


Figure 15

**Figure 16**

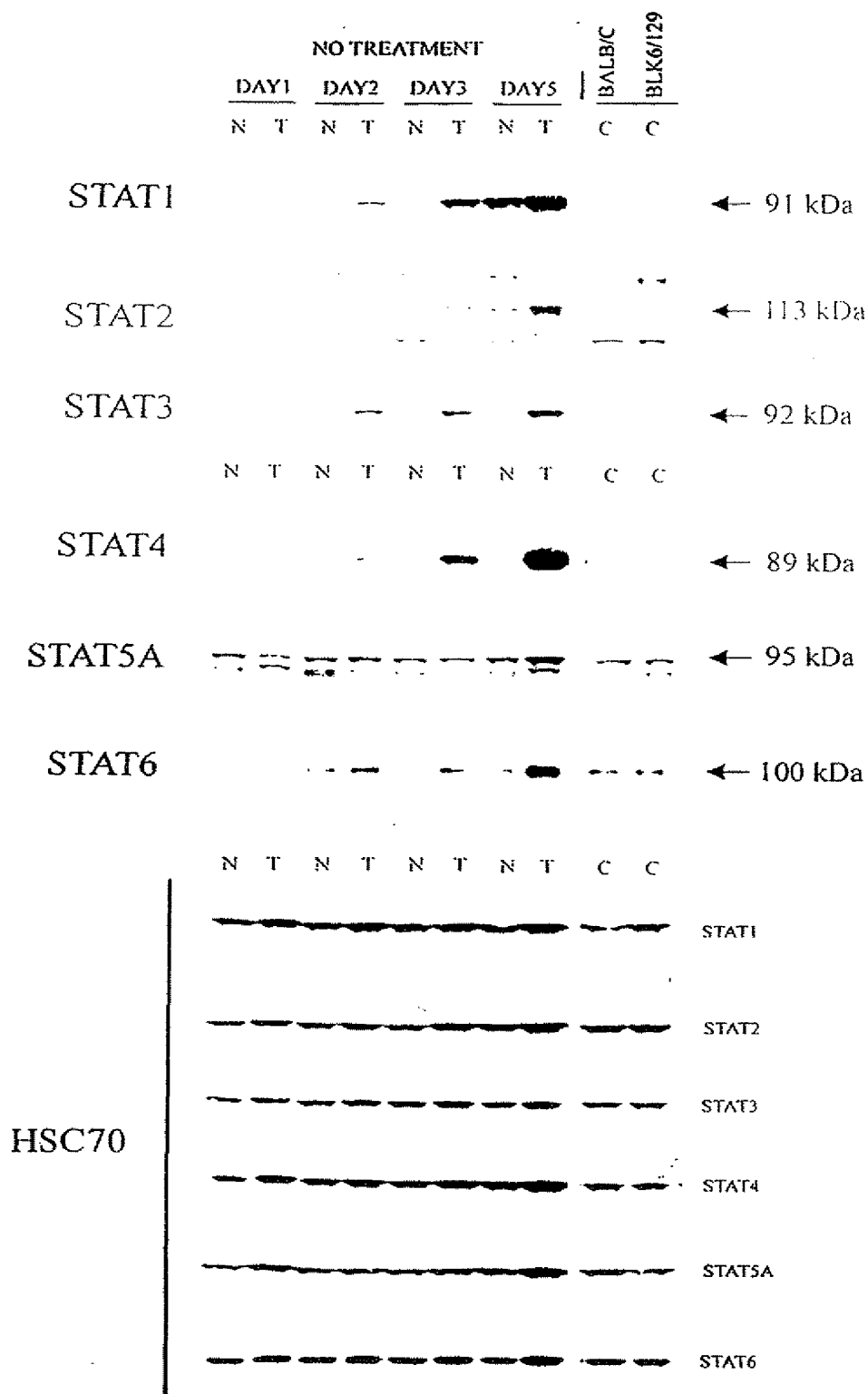


Figure 17

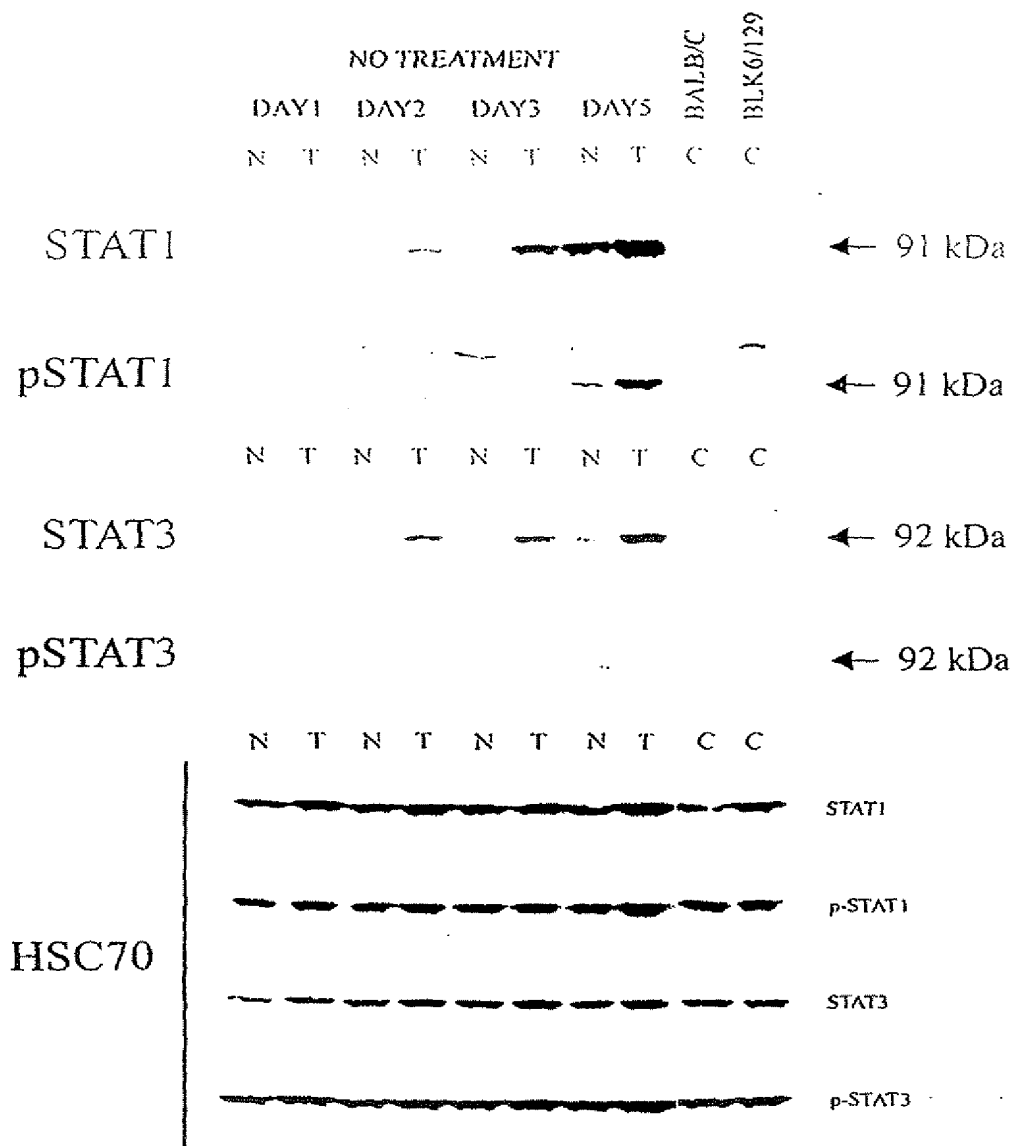


Figure 18

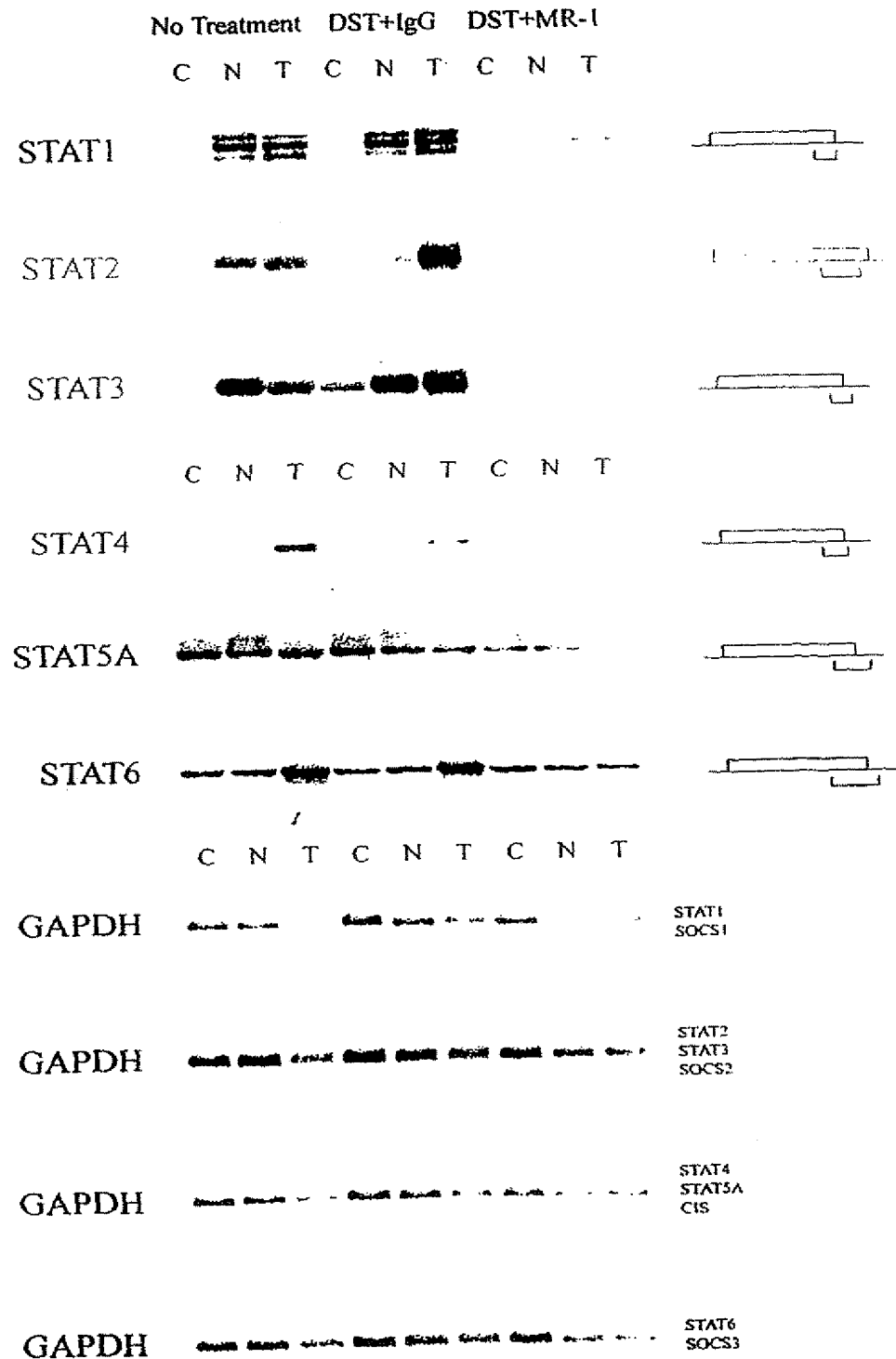
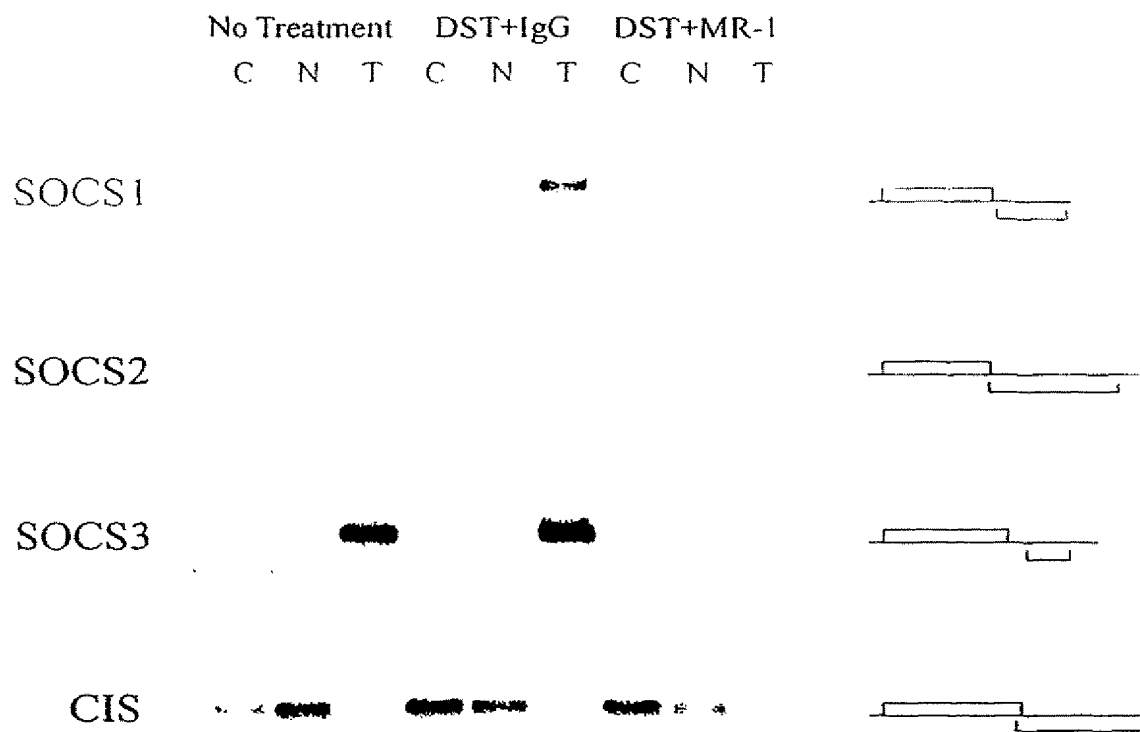
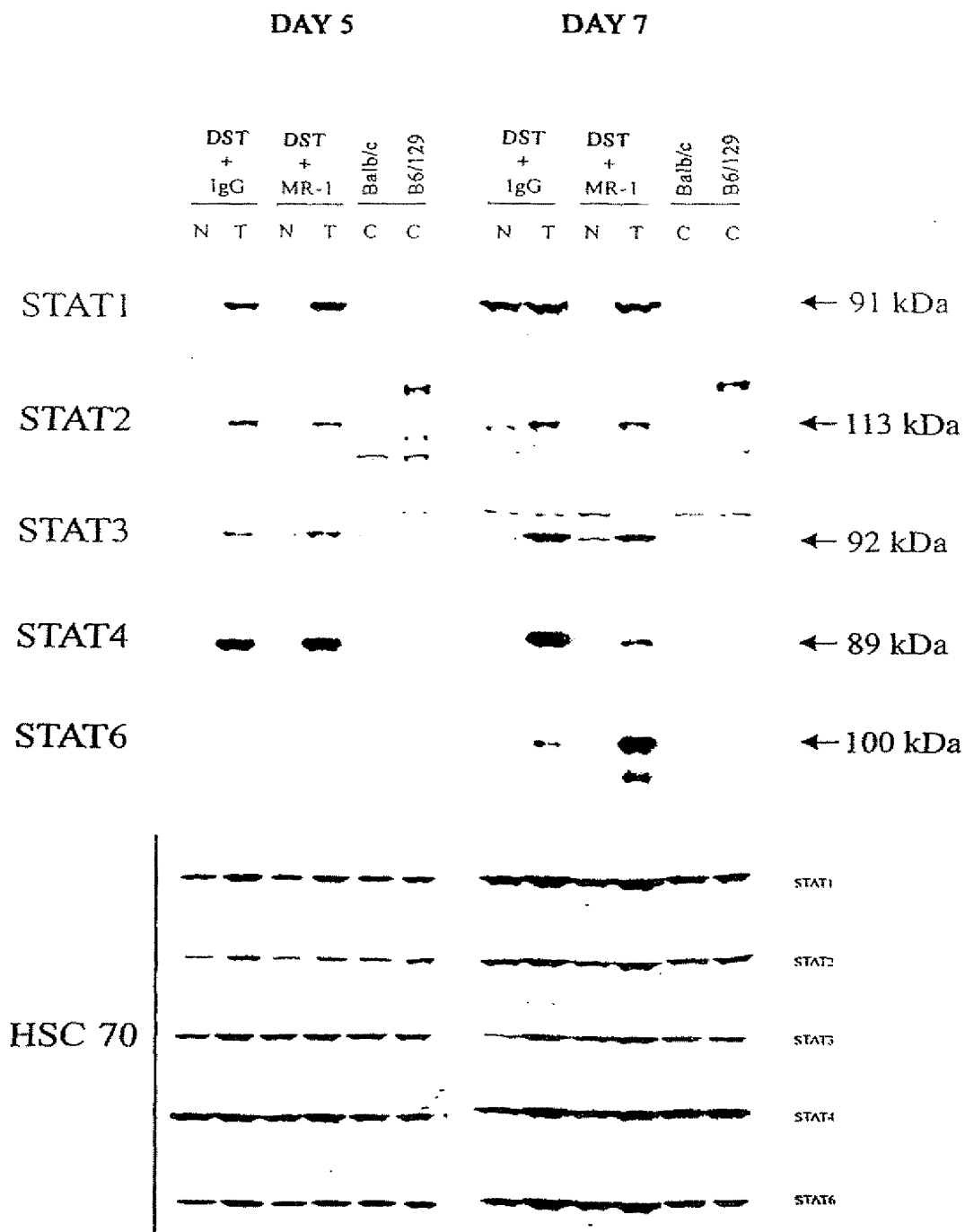


Figure 19

**Figure 20**

**Figure 21**

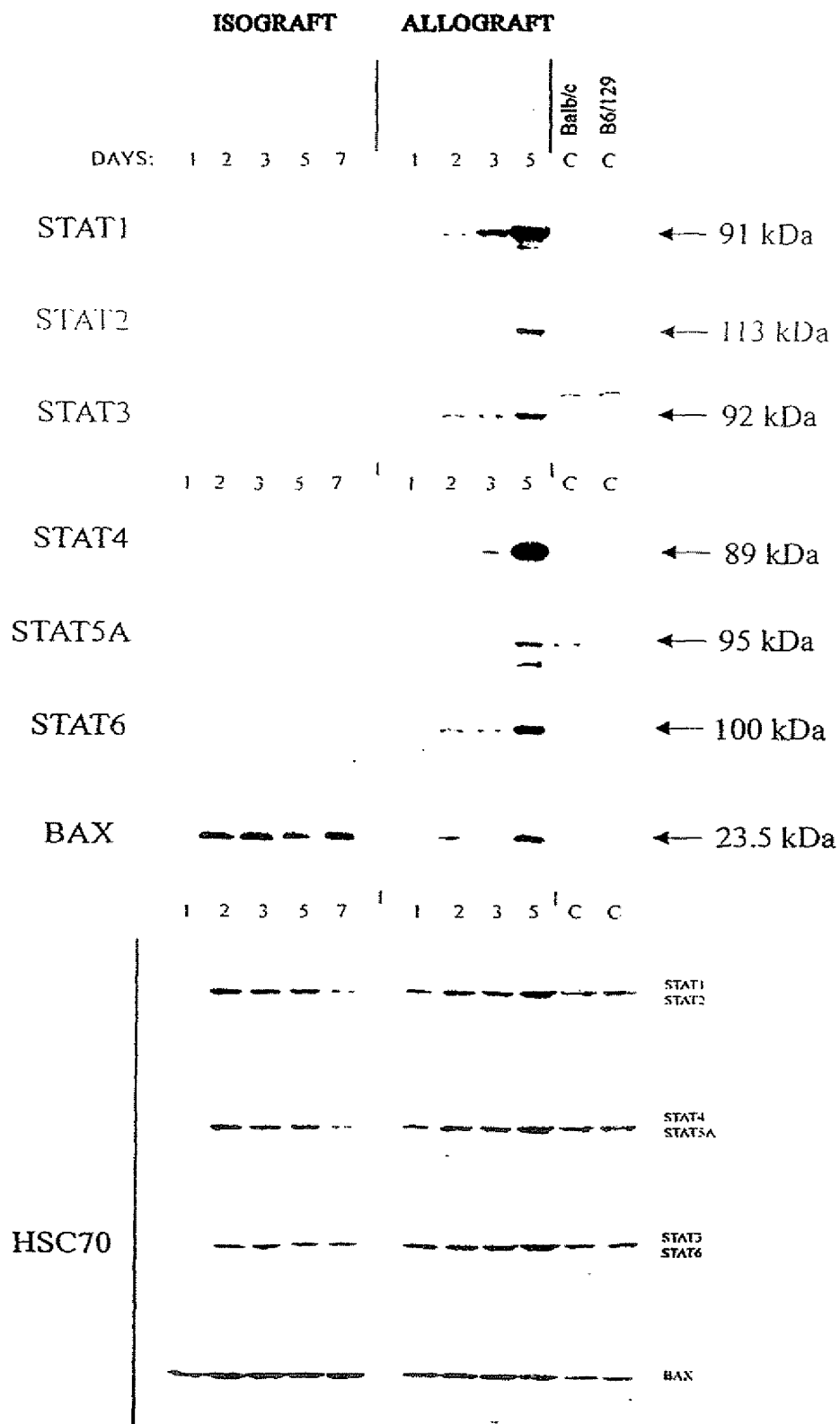


Figure 22

SEQUENCE LISTING

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ttggcaccta acgtgctgtg cgtagctgct cctttgggtg aatccccagg cccttggttg	180
ggcacaaggt ggcagg atg tct cag tgg tac gaa ctt cag cag ctt gac tca	232
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1 5 10	
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Lys Phe Leu Glu Gln Val His Gln Leu Tyr Asp Asp Ser Phe Pro Met	
15 20 25	
gaa atc aga cag tac ctg gca cag tgg tta gaa aag caa gac tgg gag	328
Glu Ile Arg Gln Tyr Leu Ala Gln Trp Leu Glu Lys Gln Asp Trp Glu	
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cac gct gcc aat gat gtt tca ttt gcc acc atc cgt ttt cat gac ctc	376
His Ala Ala Asn Asp Val Ser Phe Ala Thr Ile Arg Phe His Asp Leu	
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Leu Ser Gln Leu Asp Asp Gln Tyr Ser Arg Phe Ser Leu Glu Asn Asn	
65 70 75	
ttc ttg cta cag cat aac ata agg aaa agc aag cgt aat ctt cag gat	472
Phe Leu Leu Gln His Asn Ile Arg Lys Ser Lys Arg Asn Leu Gln Asp	
80 85 90	

aat ttt cag gaa gac cca atc cag atg tct atg atc att tac agc tgt	520
Asn Phe Gln Glu Asp Pro Ile Gln Met Ser Met Ile Ile Tyr Ser Cys	
95 100 105	
ctg aag gaa gaa agg aaa att ctg gaa aac gcc cag aga ttt aat cag	568
Leu Lys Glu Glu Arg Lys Ile Leu Glu Asn Ala Gln Arg Phe Asn Gln	
110 115 120	
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Ala Gln Ser Gly Asn Ile Gln Ser Thr Val Met Leu Asp Lys Gln Lys	
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Glu Leu Asp Ser Lys Val Arg Asn Val Lys Asp Lys Val Met Cys Ile	
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gag cat gaa atc aag agc ctg gaa gat tta caa gat gaa tat gac ttc	712
Glu His Glu Ile Lys Ser Leu Glu Asp Leu Gln Asp Glu Tyr Asp Phe	
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Leu Asn Val Thr Glu Leu Thr Gln Asn Ala Leu Ile Asn Asp Glu Leu	
225 230 235	
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Val Glu Trp Lys Arg Arg Gln Gln Ser Ala Cys Ile Gly Gly Pro Pro	
240 245 250	
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Asn Ala Cys Leu Asp Gln Leu Gln Asn Trp Phe Thr Ile Val Ala Glu	
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Ser Leu Gln Gln Val Arg Gln Gln Leu Lys Lys Leu Glu Glu Leu Glu	
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cag aaa tac acc tac gaa cat gac cct atc aca aaa aac aaa caa gtg	1096
Gln Lys Tyr Thr Tyr Glu His Asp Pro Ile Thr Lys Asn Lys Gln Val	
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Phe	Val	Val	Glu	Arg	Gln	Pro	Cys	Met	Pro	Thr	His	Pro	Gln	Arg	Pro	
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ctg	gtc	ttg	aag	aca	ggg	gtc	cag	ttc	act	gtg	aag	ttg	aga	ctg	ttg	1240
Leu	Val	Leu	Lys	Thr	Gly	Val	Gln	Phe	Thr	Val	Lys	Leu	Arg	Leu	Leu	
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gtg	aaa	ttg	caa	gag	ctg	aat	tat	aat	ttg	aaa	gtc	aaa	gtc	tta	ttt	1288
Val	Lys	Leu	Gln	Glu	Leu	Asn	Tyr	Asn	Leu	Lys	Val	Lys	Val	Leu	Phe	
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Asp	Lys	Asp	Val	Asn	Glu	Arg	Asn	Thr	Val	Lys	Gly	Phe	Arg	Lys	Phe	
	365				370					375					380	
aac	att	ttg	ggc	acg	cac	aca	aaa	gtg	atg	aac	atg	gag	gag	tcc	acc	1384
Asn	Ile	Leu	Gly	Thr	His	Thr	Lys	Val	Met	Asn	Met	Glu	Glu	Ser	Thr	
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aat	ggc	agt	ctg	gcg	gct	gaa	ttt	cgg	cac	ctg	caa	ttg	aaa	gaa	cag	1432
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Lys	Asn	Ala	Gly	Thr	Arg	Thr	Asn	Glu	Gly	Pro	Leu	Ile	Val	Thr	Glu	
		415					420					425				
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Glu	Leu	His	Ser	Leu	Ser	Phe	Glu	Thr	Gln	Leu	Cys	Gln	Pro	Gly	Leu	
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gta	att	gac	ctc	gag	acg	acc	tct	ctg	ccc	gtt	gtg	gtg	atc	tcc	aac	1576
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Val	Ser	Gln	Leu	Pro	Ser	Gly	Trp	Ala	Ser	Ile	Leu	Trp	Tyr	Asn	Met	
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ctg	gtg	gcg	gaa	ccc	agg	aat	ctg	tcc	ttc	ttc	ctg	act	cca	cca	tgt	1672
Leu	Val	Ala	Glu	Pro	Arg	Asn	Leu	Ser	Phe	Phe	Leu	Thr	Pro	Pro	Cys	
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Ala	Arg	Trp	Ala	Gln	Leu	Ser	Glu	Val	Leu	Ser	Trp	Gln	Phe	Ser	Ser	
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gtc	acc	aaa	aga	ggt	ctc	aat	gtg	gac	cag	ctg	aac	atg	ttg	gga	gag	1768
Val	Thr	Lys	Arg	Gly	Leu	Asn	Val	Asp	Gln	Leu	Asn	Met	Leu	Gly	Glu	
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tgg att gaa agc atc cta gaa ctc att aaa aaa cac ctg ctc cct ctc	1912
Trp Ile Glu Ser Ile Leu Glu Leu Ile Lys Lys His Leu Leu Pro Leu	
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Trp Asn Asp Gly Cys Ile Met Gly Phe Ile Ser Lys Glu Arg Glu Arg	
575 580 585	
gcc ctg ttg aag gac cag cag ccg ggg acc ttc ctg ctg cgg ttc agt	2008
Ala Leu Leu Lys Asp Gln Gln Pro Gly Thr Phe Leu Leu Arg Phe Ser	
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Glu Ser Ser Arg Glu Gly Ala Ile Thr Phe Thr Trp Val Glu Arg Ser	
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Gln Asn Gly Gly Glu Pro Asp Phe His Ala Val Glu Pro Tyr Thr Lys	
625 630 635	
aaa gaa ctt tct gct gtt act ttc cct gac atc att cgc aat tac aaa	2152
Lys Glu Leu Ser Ala Val Thr Phe Pro Asp Ile Ile Arg Asn Tyr Lys	
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Pro Asn Ile Asp Lys Asp His Ala Phe Gly Lys Tyr Tyr Ser Arg Pro	
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Lys Glu Ala Pro Glu Pro Met Glu Leu Asp Gly Pro Lys Gly Thr Gly	
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Tyr Ile Lys Thr Glu Leu Ile Ser Val Ser Glu Val His Pro Ser Arg	
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Thr Val *	
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35      40      45
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His Asn Ile Arg Lys Ser Lys Arg Asn Leu Gln Asp Asn Phe Gln Glu
85      90      95
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Arg Lys Ile Leu Glu Asn Ala Gln Arg Phe Asn Gln Ala Gln Ser Gly
115     120     125
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130     135     140
Lys Val Arg Asn Val Lys Asp Lys Val Met Cys Ile Glu His Glu Ile
145     150     155     160
Lys Ser Leu Glu Asp Leu Gln Asp Glu Tyr Asp Phe Lys Cys Lys Thr
165     170     175
Leu Gln Asn Arg Glu His Glu Thr Asn Gly Val Ala Lys Ser Asp Gln
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Lys Gln Glu Gln Leu Leu Leu Lys Lys Met Tyr Leu Met Leu Asp Asn
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Arg	Arg	Gln	Gln	Ser	Ala	Cys	Ile	Gly	Gly	Pro	Pro	Asn	Ala	Cys	Leu	245	250	255
Asp	Gln	Leu	Gln	Asn	Trp	Phe	Thr	Ile	Val	Ala	Glu	Ser	Leu	Gln	Gln	260	265	270
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Arg	Gln	Pro	Cys	Met	Pro	Thr	His	Pro	Gln	Arg	Pro	Leu	Val	Leu	Lys	325	330	335
Thr	Gly	Val	Gln	Phe	Thr	Val	Lys	Leu	Arg	Leu	Leu	Val	Lys	Leu	Gln	340	345	350
Glu	Leu	Asn	Tyr	Asn	Leu	Lys	Val	Lys	Val	Leu	Phe	Asp	Lys	Asp	Val	355	360	365
Asn	Glu	Arg	Asn	Thr	Val	Lys	Gly	Phe	Arg	Lys	Phe	Asn	Ile	Leu	Gly	370	375	380
Thr	His	Thr	Lys	Val	Met	Asn	Met	Glu	Glu	Ser	Thr	Asn	Gly	Ser	Leu	385	390	395
Ala	Ala	Glu	Phe	Arg	His	Leu	Gln	Leu	Lys	Glu	Gln	Lys	Asn	Ala	Gly	405	410	415
Thr	Arg	Thr	Asn	Glu	Gly	Pro	Leu	Ile	Val	Thr	Glu	Glu	Leu	His	Ser	420	425	430
Leu	Ser	Phe	Glu	Thr	Gln	Leu	Cys	Gln	Pro	Gly	Leu	Val	Ile	Asp	Leu	435	440	445
Glu	Thr	Thr	Ser	Leu	Pro	Val	Val	Val	Ile	Ser	Asn	Val	Ser	Gln	Leu	450	455	460
Pro	Ser	Gly	Trp	Ala	Ser	Ile	Leu	Trp	Tyr	Asn	Met	Leu	Val	Ala	Glu	465	470	475
Pro	Arg	Asn	Leu	Ser	Phe	Phe	Leu	Thr	Pro	Pro	Cys	Ala	Arg	Trp	Ala	485	490	495
Gln	Leu	Ser	Glu	Val	Leu	Ser	Trp	Gln	Phe	Ser	Ser	Val	Thr	Lys	Arg	500	505	510
Gly	Leu	Asn	Val	Asp	Gln	Leu	Asn	Met	Leu	Gly	Glu	Lys	Leu	Leu	Gly	515	520	525
Pro	Asn	Ala	Ser	Pro	Asp	Gly	Leu	Ile	Pro	Trp	Thr	Arg	Phe	Cys	Lys	530	535	540
Glu	Asn	Ile	Asn	Asp	Lys	Asn	Phe	Pro	Phe	Trp	Leu	Trp	Ile	Glu	Ser	545	550	555
Ile	Leu	Glu	Leu	Ile	Lys	Lys	His	Leu	Leu	Pro	Leu	Trp	Asn	Asp	Gly	565	570	575
Cys	Ile	Met	Gly	Phe	Ile	Ser	Lys	Glu	Arg	Glu	Arg	Ala	Leu	Leu	Lys	580	585	590
Asp	Gln	Gln	Pro	Gly	Thr	Phe	Leu	Leu	Arg	Phe	Ser	Glu	Ser	Ser	Arg	595	600	605
Glu	Gly	Ala	Ile	Thr	Phe	Thr	Trp	Val	Glu	Arg	Ser	Gln	Asn	Gly	Gly	610	615	620
Glu	Pro	Asp	Phe	His	Ala	Val	Glu	Pro	Tyr	Thr	Lys	Lys	Glu	Leu	Ser	625	630	635
Ala	Val	Thr	Phe	Pro	Asp	Ile	Ile	Arg	Asn	Tyr	Lys	Val	Met	Ala	Ala	645	650	655
Glu	Asn	Ile	Pro	Glu	Asn	Pro	Leu	Lys	Tyr	Leu	Tyr	Pro	Asn	Ile	Asp	660	665	670

Lys Asp His Ala Phe Gly Lys Tyr Tyr Ser Arg Pro Lys Glu Ala Pro
 675 680 685
 Glu Pro Met Glu Leu Asp Gly Pro Lys Gly Thr Gly Tyr Ile Lys Thr
 690 695 700
 Glu Leu Ile Ser Val Ser Glu Val His Pro Ser Arg Leu Gln Thr Thr
 705 710 715 720
 Asp Asn Leu Leu Pro Met Ser Pro Glu Glu Phe Asp Glu Val Ser Arg
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 Ala Gln Trp Glu Met Leu Gln Asn Leu Asp Ser Pro Phe Gln Asp Gln
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 ctg cac cag ctt tac tcg cac agc ctc ctg cct gtg gac att cga cag 156
 Leu His Gln Leu Tyr Ser His Ser Leu Leu Pro Val Asp Ile Arg Gln
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 tac ttg gct gtc tgg att gaa gac cag aac tgg cag gaa gct gca ctt 204
 Tyr Leu Ala Val Trp Ile Glu Asp Gln Asn Trp Gln Glu Ala Ala Leu
 35 40 45

 ggg agt gat gat tcc aag gct acc atg cta ttc ttc cac ttc ttg gat 252
 Gly Ser Asp Asp Ser Lys Ala Thr Met Leu Phe Phe His Phe Leu Asp
 50 55 60 65

 cag ctg aac tat gag tgt ggc cgt tgc agc cag gac cca gag tcc ttg 300
 Gln Leu Asn Tyr Glu Cys Gly Arg Cys Ser Gln Asp Pro Glu Ser Leu
 70 75 80

 ttg ctg cag cac aat ttg cgg aaa ttc tgc cgg gac att cag ccc ttt 348
 Leu Leu Gln His Asn Leu Arg Lys Phe Cys Arg Asp Ile Gln Pro Phe
 85 90 95

 tcc cag gat cct acc cag ttg gct gag atg atc ttt aac ctc ctt ctg 396
 Ser Gln Asp Pro Thr Gln Leu Ala Glu Met Ile Phe Asn Leu Leu Leu
 100 105 110

 gaa gaa aaa aga att ttg atc cag gct cag agg gcc caa ttg gaa caa 444
 Glu Glu Lys Arg Ile Leu Ile Gln Ala Gln Arg Ala Gln Leu Glu Gln
 115 120 125

gga gag cca gtt ctc gaa aca cct gtg gag agc cag caa cat gag att	492
Gly Glu Pro Val Leu Glu Thr Pro Val Glu Ser Gln Gln His Glu Ile	
130 135 140 145	
gaa tcc cgg atc ctg gat tta agg gct atg atg gag aag ctg gta aaa	540
Glu Ser Arg Ile Leu Asp Leu Arg Ala Met Met Glu Lys Leu Val Lys	
150 155 160	
tcc atc agc caa ctg aaa gac cag cag gat gtc ttc tgc ttc cga tat	588
Ser Ile Ser Gln Leu Lys Asp Gln Gln Asp Val Phe Cys Phe Arg Tyr	
165 170 175	
aag atc cag gcc aaa ggg aag aca ccc tct ctg gac ccc cat cag acc	636
Lys Ile Gln Ala Lys Gly Lys Thr Pro Ser Leu Asp Pro His Gln Thr	
180 185 190	
aaa gag cag aag att ctg cag gaa act ctc aat gaa ctg gac aaa agg	684
Lys Glu Gln Lys Ile Leu Gln Glu Thr Leu Asn Glu Leu Asp Lys Arg	
195 200 205	
aga aag gag gtg ctg gat gcc tcc aaa gca ctg cta ggc cga tta act	732
Arg Lys Glu Val Leu Asp Ala Ser Lys Ala Leu Leu Gly Arg Leu Thr	
210 215 220 225	
acc cta atc gag cta ctg ctg cca aag ttg gag gag tgg aag gcc cag	780
Thr Leu Ile Glu Leu Leu Leu Pro Lys Leu Glu Glu Trp Lys Ala Gln	
230 235 240	
cag caa aaa gcc tgc atc aga gct ccc att gac cac ggg ttg gaa cag	828
Gln Gln Lys Ala Cys Ile Arg Ala Pro Ile Asp His Gly Leu Glu Gln	
245 250 255	
ctg gag aca tgg ttc aca gct gga gca aag ctg ttg ttt cac ctg agg	876
Leu Glu Thr Trp Phe Thr Ala Gly Ala Lys Leu Leu Phe His Leu Arg	
260 265 270	
cag ctg ctg aag gag ctg aag gga ctg agt tgc ctg gtt agc tat cag	924
Gln Leu Leu Lys Glu Leu Lys Gly Leu Ser Cys Leu Val Ser Tyr Gln	
275 280 285	
gat gac cct ctg acc aaa ggg gtg gac cta cgc aac gcc cag gtc aca	972
Asp Asp Pro Leu Thr Lys Gly Val Asp Leu Arg Asn Ala Gln Val Thr	
290 295 300 305	
gag ttg cta cag cgt ctg ctc cac aga gcc ttt gtg gta gaa acc cag	1020
Glu Leu Leu Gln Arg Leu Leu His Arg Ala Phe Val Val Glu Thr Gln	
310 315 320	
ccc tgc atg ccc caa act ccc cat cga ccc ctc atc ctc aag act ggc	1068
Pro Cys Met Pro Gln Thr Pro His Arg Pro Leu Ile Leu Lys Thr Gly	
325 330 335	
agc aag ttc acc gtc cga aca agg ctg ctg gtg aga ctc cag gaa ggc	1116
Ser Lys Phe Thr Val Arg Thr Arg Leu Leu Val Arg Leu Gln Glu Gly	
340 345 350	

aat gag tca ctg act gtg gaa gtc tcc att gac agg aat cct cct caa Asn Glu Ser Leu Thr Val Glu Val Ser Ile Asp Arg Asn Pro Pro Gln 355 360 365	1164
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ttg acc ccc gag aag ggg cag agt cag ggt ttg att tgg gac ttt ggt Leu Thr Pro Glu Lys Gly Gln Ser Gln Gly Leu Ile Trp Asp Phe Gly 390 395 400	1260
tac ctg act ctg gtg gag caa cgt tca ggt ggt tca gga aag ggc agc Tyr Leu Thr Leu Val Glu Gln Arg Ser Gly Gly Ser Gly Lys Gly Ser 405 410 415	1308
aat aag ggg cca cta ggt gtg aca gag gaa ctg cac atc atc agc ttc Asn Lys Gly Pro Leu Gly Val Thr Glu Glu Leu His Ile Ile Ser Phe 420 425 430	1356
acg gtc aaa tat acc tac cag ggt ctg aag cag gag ctg aaa acg gac Thr Val Lys Tyr Thr Tyr Gln Gly Leu Lys Gln Glu Leu Lys Thr Asp 435 440 445	1404
acc ctc cct gtg gtg att att tcc aac atg aac cag ctc tca att gcc Thr Leu Pro Val Val Ile Ile Ser Asn Met Asn Gln Leu Ser Ile Ala 450 455 460 465	1452
tgg gct tca gtt ctc tgg ttc aat ttg ctc agc cca aac ctt cag aac Trp Ala Ser Val Leu Trp Phe Asn Leu Leu Ser Pro Asn Leu Gln Asn 470 475 480	1500
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Met Ser Gly Thr Phe Leu Leu Arg Phe Ser Glu Ser Ser Glu Gly Gly	
595 600 605	
att acc tgc tcc tgg gtg gag cac cag gat gat gac aag gtg ctc atc	1932
Ile Thr Cys Ser Trp Val Glu His Gln Asp Asp Asp Lys Val Leu Ile	
610 615 620 625	
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Tyr Ser Val Gln Pro Tyr Thr Lys Glu Val Leu Gln Ser Leu Pro Leu	
630 635 640	
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Thr Glu Ile Ile Arg His Tyr Gln Leu Leu Thr Glu Glu Asn Ile Pro	
645 650 655	
gaa aac cca ctg cgc ttc ctc tat ccc cga atc ccc cgg gat gaa gct	2076
Glu Asn Pro Leu Arg Phe Leu Tyr Pro Arg Ile Pro Arg Asp Glu Ala	
660 665 670	
ttt ggg tgc tac tac cag gag aaa gtt aat ctc cag gaa cgg agg aaa	2124
Phe Gly Cys Tyr Tyr Gln Glu Lys Val Asn Leu Gln Glu Arg Arg Lys	
675 680 685	
tac ctg aaa cac agg ctc att gtg gtc tct aat aga cag gtg gat gaa	2172
Tyr Leu Lys His Arg Leu Ile Val Val Ser Asn Arg Gln Val Asp Glu	
690 695 700 705	
ctg caa caa ccg ctg gag ctt aag cca gag cca gag ctg gag tca tta	2220
Leu Gln Gln Pro Leu Glu Leu Lys Pro Glu Pro Glu Leu Glu Ser Leu	
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gag ctg gaa cta ggg ctg gtg cca gag cca gag ctc agc ctg gac tta	2268
Glu Leu Glu Leu Gly Leu Val Pro Glu Pro Glu Leu Ser Leu Asp Leu	
725 730 735	
gag cca ctg ctg aag gca ggg ctg gat ctg ggg cca gag cta gag tct	2316
Glu Pro Leu Leu Lys Ala Gly Leu Asp Leu Gly Pro Glu Leu Glu Ser	
740 745 750	
gtg ctg gag tcc act ctg gag cct gtg ata gag ccc acc cta tgc atg	2364
Val Leu Glu Ser Thr Leu Glu Pro Val Ile Glu Pro Thr Leu Cys Met	
755 760 765	
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Val Ser Gln Thr Val Pro Glu Pro Asp Gln Gly Pro Val Ser Gln Pro	
770 775 780 785	
gtg cca gag cca gat ttg ccc tgt gat ctg aga cat ttg aac act gag	2460
Val Pro Glu Pro Asp Leu Pro Cys Asp Leu Arg His Leu Asn Thr Glu	
790 795 800	

cca atg gaa atc ttc aga aac tgt gta aag att gaa gaa atc atg ccg 2508
 Pro Met Glu Ile Phe Arg Asn Cys Val Lys Ile Glu Glu Ile Met Pro
 805 810 815

aat ggt gac cca ctg ttg gct ggc cag aac acc gtg gat gag gtt tac 2556
 Asn Gly Asp Pro Leu Leu Ala Gly Gln Asn Thr Val Asp Glu Val Tyr
 820 825 830

gtc tcc cgc ccc agc cac ttc tac act gat gga ccc ttg atg cct tct 2604
 Val Ser Arg Pro Ser His Phe Tyr Thr Asp Gly Pro Leu Met Pro Ser
 835 840 845

gac ttc tag gaaccacatt tcctctgttc ttttcatatc tctttgcctt 2653
 Asp Phe *
 850

tcctactcct catagcatga tattgtttctc caaggatggg aatcaggcat gtgtcccttc 2713
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<212> PRT

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 Gln Tyr Leu Ala Val Trp Ile Glu Asp Gln Asn Trp Gln Glu Ala Ala
 35 40 45
 Leu Gly Ser Asp Asp Ser Lys Ala Thr Met Leu Phe Phe His Phe Leu
 50 55 60
 Asp Gln Leu Asn Tyr Glu Cys Gly Arg Cys Ser Gln Asp Pro Glu Ser
 65 70 75 80
 Leu Leu Leu Gln His Asn Leu Arg Lys Phe Cys Arg Asp Ile Gln Pro
 85 90 95
 Phe Ser Gln Asp Pro Thr Gln Leu Ala Glu Met Ile Phe Asn Leu Leu
 100 105 110
 Leu Glu Glu Lys Arg Ile Leu Ile Gln Ala Gln Arg Ala Gln Leu Glu
 115 120 125
 Gln Gly Glu Pro Val Leu Glu Thr Pro Val Glu Ser Gln Gln His Glu
 130 135 140
 Ile Glu Ser Arg Ile Leu Asp Leu Arg Ala Met Met Glu Lys Leu Val
 145 150 155 160
 Lys Ser Ile Ser Gln Leu Lys Asp Gln Gln Asp Val Phe Cys Phe Arg
 165 170 175
 Tyr Lys Ile Gln Ala Lys Gly Lys Thr Pro Ser Leu Asp Pro His Gln
 180 185 190

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Arg Arg Lys Glu Val Leu Asp Ala Ser Lys Ala Leu Leu Gly Arg Leu
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Thr Thr Leu Ile Glu Leu Leu Leu Pro Lys Leu Glu Glu Trp Lys Ala
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Gln Gln Gln Lys Ala Cys Ile Arg Ala Pro Ile Asp His Gly Leu Glu
      245                250                255
Gln Leu Glu Thr Trp Phe Thr Ala Gly Ala Lys Leu Leu Phe His Leu
      260                265                270
Arg Gln Leu Leu Lys Glu Leu Lys Gly Leu Ser Cys Leu Val Ser Tyr
      275                280                285
Gln Asp Asp Pro Leu Thr Lys Gly Val Asp Leu Arg Asn Ala Gln Val
      290                295                300
Thr Glu Leu Leu Gln Arg Leu Leu His Arg Ala Phe Val Val Glu Thr
305      310                315                320
Gln Pro Cys Met Pro Gln Thr Pro His Arg Pro Leu Ile Leu Lys Thr
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Gly Ser Lys Phe Thr Val Arg Thr Arg Leu Leu Val Arg Leu Gln Glu
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Gly Asn Glu Ser Leu Thr Val Glu Val Ser Ile Asp Arg Asn Pro Pro
      355                360                365
Gln Leu Gln Gly Phe Arg Lys Phe Asn Ile Leu Thr Ser Asn Gln Lys
      370                375                380
Thr Leu Thr Pro Glu Lys Gly Gln Ser Gln Gly Leu Ile Trp Asp Phe
385      390                395                400
Gly Tyr Leu Thr Leu Val Glu Gln Arg Ser Gly Gly Ser Gly Lys Gly
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Ser Asn Lys Gly Pro Leu Gly Val Thr Glu Glu Leu His Ile Ile Ser
      420                425                430
Phe Thr Val Lys Tyr Thr Tyr Gln Gly Leu Lys Gln Glu Leu Lys Thr
      435                440                445
Asp Thr Leu Pro Val Val Ile Ile Ser Asn Met Asn Gln Leu Ser Ile
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Ala Trp Ala Ser Val Leu Trp Phe Asn Leu Leu Ser Pro Asn Leu Gln
465      470                475                480
Asn Gln Gln Phe Phe Ser Asn Pro Pro Lys Ala Pro Trp Ser Leu Leu
      485                490                495
Gly Pro Ala Leu Ser Trp Gln Phe Ser Ser Tyr Val Gly Arg Gly Leu
      500                505                510
Asn Ser Asp Gln Leu Ser Met Leu Arg Asn Lys Leu Phe Gly Gln Asn
      515                520                525
Cys Arg Thr Glu Asp Pro Leu Leu Ser Trp Ala Asp Phe Thr Lys Arg
      530                535                540
Glu Ser Pro Pro Gly Lys Leu Pro Phe Trp Thr Trp Leu Asp Lys Ile
545      550                555                560
Leu Glu Leu Val His Asp His Leu Lys Asp Leu Trp Asn Asp Gly Arg
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Ile Met Gly Phe Val Ser Arg Ser Gln Glu Arg Arg Leu Leu Lys Lys
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Thr Met Ser Gly Thr Phe Leu Leu Arg Phe Ser Glu Ser Ser Glu Gly
      595                600                605
Gly Ile Thr Cys Ser Trp Val Glu His Gln Asp Asp Asp Lys Val Leu
      610                615                620
Ile Tyr Ser Val Gln Pro Tyr Thr Lys Glu Val Leu Gln Ser Leu Pro
625      630                635                640
Leu Thr Glu Ile Ile Arg His Tyr Gln Leu Leu Thr Glu Glu Asn Ile
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Lys Tyr Leu Lys His Arg Leu Ile Val Val Ser Asn Arg Gln Val Asp
      690      695      700
Glu Leu Gln Gln Pro Leu Glu Leu Lys Pro Glu Pro Glu Leu Glu Ser
705      710      715      720
Leu Glu Leu Glu Leu Gly Leu Val Pro Glu Pro Glu Leu Ser Leu Asp
      725      730      735
Leu Glu Pro Leu Leu Lys Ala Gly Leu Asp Leu Gly Pro Glu Leu Glu
      740      745      750
Ser Val Leu Glu Ser Thr Leu Glu Pro Val Ile Glu Pro Thr Leu Cys
      755      760      765
Met Val Ser Gln Thr Val Pro Glu Pro Asp Gln Gly Pro Val Ser Gln
      770      775      780
Pro Val Pro Glu Pro Asp Leu Pro Cys Asp Leu Arg His Leu Asn Thr
785      790      795      800
Glu Pro Met Glu Ile Phe Arg Asn Cys Val Lys Ile Glu Glu Ile Met
      805      810      815
Pro Asn Gly Asp Pro Leu Leu Ala Gly Gln Asn Thr Val Asp Glu Val
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Tyr Val Ser Arg Pro Ser His Phe Tyr Thr Asp Gly Pro Leu Met Pro
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Ser Asp Phe
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ccccctcggc tcggagaggg ccttcggcct gagggagcct cgccgcccgt ccccggcaca      180
cgcgcagccc cggcctctcg gcctctgccg gagaaacagg atg gcc caa tgg aat      235
                               Met Ala Gln Trp Asn
                               1           5

cag cta cag cag ctt gac aca cgg tac ctg gag cag ctc cat cag ctc      283
Gln Leu Gln Gln Leu Asp Thr Arg Tyr Leu Glu Gln Leu His Gln Leu
      10      15      20

tac agt gac agc ttc cca atg gag ctg cgg cag ttt ctg gcc cct tgg      331
Tyr Ser Asp Ser Phe Pro Met Glu Leu Arg Gln Phe Leu Ala Pro Trp
      25      30      35

att gag agt caa gat tgg gca tat gcg gcc agc aaa gaa tca cat gcc      379
Ile Glu Ser Gln Asp Trp Ala Tyr Ala Ala Ser Lys Glu Ser His Ala
      40      45      50

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Thr	Leu	Val	Phe	His	Asn	Leu	Leu	Gly	Glu	Ile	Asp	Gln	Gln	Tyr	Ser	
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cgc	ttc	ctg	caa	gag	tgc	aat	gtt	ctc	tat	cag	cac	aat	cta	cga	aga	475
Arg	Phe	Leu	Gln	Glu	Ser	Asn	Val	Leu	Tyr	Gln	His	Asn	Leu	Arg	Arg	
	70				75				80					85		
atc	aag	cag	ttt	ctt	cag	agc	agg	tat	ctt	gag	aag	cca	atg	gag	att	523
Ile	Lys	Gln	Phe	Leu	Gln	Ser	Arg	Tyr	Leu	Glu	Lys	Pro	Met	Glu	Ile	
				90					95					100		
gcc	cgg	att	gtg	gcc	cgg	tgc	ctg	tgg	gaa	gaa	tca	cgc	ctt	cta	cag	571
Ala	Arg	Ile	Val	Ala	Arg	Cys	Leu	Trp	Glu	Glu	Ser	Arg	Leu	Leu	Gln	
			105					110					115			
act	gca	gcc	act	gcg	gcc	cag	caa	ggg	ggc	cag	gcc	aac	cac	ccc	aca	619
Thr	Ala	Ala	Thr	Ala	Ala	Gln	Gln	Gly	Gly	Gln	Ala	Asn	His	Pro	Thr	
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gca	gcc	gtg	gtg	acg	gag	aag	cag	cag	atg	ctg	gag	cag	cac	ctt	cag	667
Ala	Ala	Val	Val	Thr	Glu	Lys	Gln	Gln	Met	Leu	Glu	Gln	His	Leu	Gln	
		135				140					145					
gat	gtc	cgg	aag	aga	gtg	cag	gat	cta	gaa	cag	aaa	atg	aaa	gtg	gta	715
Asp	Val	Arg	Lys	Arg	Val	Gln	Asp	Leu	Glu	Gln	Lys	Met	Lys	Val	Val	
	150					155				160				165		
gag	aat	ctc	cag	gat	gac	ttt	gat	ttc	aac	tat	aaa	acc	ctc	aag	agt	763
Glu	Asn	Leu	Gln	Asp	Asp	Phe	Asp	Phe	Asn	Tyr	Lys	Thr	Leu	Lys	Ser	
				170					175					180		
caa	gga	gac	atg	caa	gat	ctg	aat	gga	aac	aac	cag	tca	gtg	acc	agg	811
Gln	Gly	Asp	Met	Gln	Asp	Leu	Asn	Gly	Asn	Asn	Gln	Ser	Val	Thr	Arg	
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Gln	Lys	Met	Gln	Gln	Leu	Glu	Gln	Met	Leu	Thr	Ala	Leu	Asp	Gln	Met	
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Arg	Arg	Ser	Ile	Val	Ser	Glu	Leu	Ala	Gly	Leu	Leu	Ser	Ala	Met	Glu	
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Tyr	Val	Gln	Lys	Thr	Leu	Thr	Asp	Glu	Glu	Leu	Ala	Asp	Trp	Lys	Arg	
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Arg	Gln	Gln	Ile	Ala	Cys	Ile	Gly	Gly	Pro	Pro	Asn	Ile	Cys	Leu	Asp	
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Arg	Leu	Glu	Asn	Trp	Ile	Thr	Ser	Leu	Ala	Glu	Ser	Gln	Leu	Gln	Thr	
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cgt	caa	caa	att	aag	aaa	ctg	gag	gag	ttg	cac	caa	aaa	gtt	tcc	tac	1099
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Lys	Gly	Asp	Pro	Ile	Val	Gln	His	Arg	Pro	Met	Leu	Glu	Glu	Arg	Ile	
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Val	Glu	Leu	Phe	Arg	Asn	Leu	Met	Lys	Ser	Ala	Phe	Val	Val	Glu	Arg	
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Gln	Pro	Cys	Met	Pro	Met	His	Pro	Asp	Arg	Pro	Leu	Val	Ile	Lys	Thr	
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ggc	gtc	cag	ttc	act	act	aaa	gtc	agg	ttg	ctg	gtc	aag	ttc	cct	gag	1291
Gly	Val	Gln	Phe	Thr	Thr	Lys	Val	Arg	Leu	Leu	Val	Lys	Phe	Pro	Glu	
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Asp	Val	Ala	Ala	Leu	Arg	Gly	Ser	Arg	Lys	Phe	Asn	Ile	Leu	Gly	Thr	
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Asn	Thr	Lys	Val	Met	Asn	Met	Glu	Glu	Ser	Asn	Asn	Gly	Ser	Leu	Ser	
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Ala	Glu	Phe	Lys	His	Leu	Thr	Leu	Arg	Glu	Gln	Arg	Cys	Gly	Asn	Gly	
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ggc	cga	gcc	aat	tgt	gat	gct	tcc	ctg	att	gtg	act	gag	gag	ctg	cac	1531
Gly	Arg	Ala	Asn	Cys	Asp	Ala	Ser	Leu	Ile	Val	Thr	Glu	Glu	Leu	His	
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cta	gag	acc	cac	tcc	ttg	tca	gtt	gtg	gtg	atc	tcc	aac	atc	tgt	cag	1627
Leu	Glu	Thr	His	Ser	Leu	Ser	Val	Val	Val	Ile	Ser	Asn	Ile	Cys	Gln	
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Met	Pro	Asn	Ala	Trp	Ala	Ser	Ile	Leu	Trp	Tyr	Asn	Met	Leu	Thr	Asn	
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Asn	Pro	Lys	Asn	Val	Asn	Phe	Phe	Thr	Lys	Pro	Pro	Ile	Gly	Thr	Trp	
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Asp	Gln	Val	Ala	Glu	Val	Leu	Ser	Trp	Gln	Phe	Ser	Ser	Thr	Thr	Lys	
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Arg	Gly	Leu	Ser	Ile	Glu	Gln	Leu	Thr	Thr	Leu	Ala	Glu	Lys	Leu	Leu	
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ggg	cct	ggt	gtg	aac	tac	tca	ggg	tgt	cag	atc	aca	tgg	gct	aac	ttc	1867
Gly	Pro	Gly	Val	Asn	Tyr	Ser	Gly	Cys	Gln	Ile	Thr	Trp	Ala	Asn	Phe	
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tgc	aaa	g	aac	atg	gct	ggc	aag	ggc	ttc	tcc	tac	tgg	gtc	tgg	cta	1915
Cys	Lys	G	Asn	Met	Ala	Gly	Lys	Gly	Phe	Ser	Tyr	Trp	Val	Trp	Leu	
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Asp	Asn	Ile	Ile	Asp	Leu	Val	Lys	Lys	Tyr	Ile	Leu	Ala	Leu	Trp	Asn	
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gaa	ggg	tac	atc	atg	ggt	ttc	atc	agc	aag	gag	cgg	gag	cgg	gcc	atc	2011
Glu	Gly	Tyr	Ile	Met	Gly	Phe	Ile	Ser	Lys	Glu	Arg	Glu	Arg	Ala	Ile	
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Leu	Ser	Thr	Lys	Pro	Pro	Gly	Thr	Phe	Leu	Leu	Arg	Phe	Ser	Glu	Ser	
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agc	aaa	gaa	gga	ggc	gtc	act	ttc	act	tgg	gtg	gag	aag	gac	atc	agc	2107
Ser	Lys	Glu	Gly	Gly	Val	Thr	Phe	Thr	Trp	Val	Glu	Lys	Asp	Ile	Ser	
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Asn	Asn	Met	Ser	Phe	Ala	Glu	Ile	Ile	Met	Gly	Tyr	Lys	Ile	Met	Asp	
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Ala	Thr	Asn	Ile	Leu	Leu	Ser	Pro	Leu	Val	Tyr	Leu	Tyr	Pro	Asp	Ile	
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ccc	aag	gag	gag	gca	ttc	ggg	aag	tat	tgt	cgg	cca	gag	agc	cag	gag	2299
Pro	Lys	Glu	Glu	Ala	Phe	Gly	Lys	Tyr	Cys	Arg	Pro	Glu	Ser	Gln	Glu	
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His	Pro	Glu	Ala	Asp	Pro	Gly	Ser	Ala	Ala	Pro	Tyr	Leu	Lys	Thr	Lys	
	695					700					705					
ttt	atc	tgt	gtg	aca	cca	acg	acc	tgc	agc	aat	acc	att	gac	ctg	ccg	2395
Phe	Ile	Cys	Val	Thr	Pro	Thr	Thr	Cys	Ser	Asn	Thr	Ile	Asp	Leu	Pro	
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gaa ggt gct gaa ccc tca gca gga ggg cag ttt gag tcc ctc acc ttt      2491
Glu Gly Ala Glu Pro Ser Ala Gly Gly Gln Phe Glu Ser Leu Thr Phe
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gac atg gag ttg acc tcg gag tgc gct acc tcc ccc atg tga      2533
Asp Met Glu Leu Thr Ser Glu Cys Ala Thr Ser Pro Met *
              760                      765                      770

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Phe Leu Ala Pro Trp Ile Glu Ser Gln Asp Trp Ala Tyr Ala Ala Ser
 35          40          45
Lys Glu Ser His Ala Thr Leu Val Phe His Asn Leu Leu Gly Glu Ile
 50          55          60
Asp Gln Gln Tyr Ser Arg Phe Leu Gln Glu Ser Asn Val Leu Tyr Gln
 65          70          75          80
His Asn Leu Arg Arg Ile Lys Gln Phe Leu Gln Ser Arg Tyr Leu Glu
 85          90          95
Lys Pro Met Glu Ile Ala Arg Ile Val Ala Arg Cys Leu Trp Glu Glu
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Ser Arg Leu Leu Gln Thr Ala Ala Thr Ala Ala Gln Gln Gly Gly Gln
115          120          125
Ala Asn His Pro Thr Ala Ala Val Val Thr Glu Lys Gln Gln Met Leu
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Glu Gln His Leu Gln Asp Val Arg Lys Arg Val Gln Asp Leu Glu Gln
145          150          155          160
Lys Met Lys Val Val Glu Asn Leu Gln Asp Asp Phe Asp Phe Asn Tyr
165          170          175
Lys Thr Leu Lys Ser Gln Gly Asp Met Gln Asp Leu Asn Gly Asn Asn
180          185          190
Gln Ser Val Thr Arg Gln Lys Met Gln Gln Leu Glu Gln Met Leu Thr
195          200          205
Ala Leu Asp Gln Met Arg Arg Ser Ile Val Ser Glu Leu Ala Gly Leu
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Leu Ser Ala Met Glu Tyr Val Gln Lys Thr Leu Thr Asp Glu Glu Leu
225          230          235          240
Ala Asp Trp Lys Arg Arg Gln Gln Ile Ala Cys Ile Gly Gly Pro Pro
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Asn Ile Cys Leu Asp Arg Leu Glu Asn Trp Ile Thr Ser Leu Ala Glu
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Gln Lys Val Ser Tyr Lys Gly Asp Pro Ile Val Gln His Arg Pro Met
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Leu Glu Glu Arg Ile Val Glu Leu Phe Arg Asn Leu Met Lys Ser Ala
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Phe Val Val Glu Arg Gln Pro Cys Met Pro Met His Pro Asp Arg Pro
    325                                330                                335
Leu Val Ile Lys Thr Gly Val Gln Phe Thr Thr Lys Val Arg Leu Leu
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Val Lys Phe Pro Glu Leu Asn Tyr Gln Leu Lys Ile Lys Val Cys Ile
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Asp Lys Asp Ser Gly Asp Val Ala Ala Leu Arg Gly Ser Arg Lys Phe
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Asn Ile Leu Gly Thr Asn Thr Lys Val Met Asn Met Glu Glu Ser Asn
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Arg Cys Gly Asn Gly Gly Arg Ala Asn Cys Asp Ala Ser Leu Ile Val
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Thr Glu Glu Leu His Leu Ile Thr Phe Glu Thr Glu Val Tyr His Gln
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Gly Leu Lys Ile Asp Leu Glu Thr His Ser Leu Ser Val Val Val Ile
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Ser Asn Ile Cys Gln Met Pro Asn Ala Trp Ala Ser Ile Leu Trp Tyr
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Asn Met Leu Thr Asn Asn Pro Lys Asn Val Asn Phe Phe Thr Lys Pro
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Pro Ile Gly Thr Trp Asp Gln Val Ala Glu Val Leu Ser Trp Gln Phe
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Ser Ser Thr Thr Lys Arg Gly Leu Ser Ile Glu Gln Leu Thr Thr Leu
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Ala Glu Lys Leu Leu Gly Pro Gly Val Asn Tyr Ser Gly Cys Gln Ile
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Thr Trp Ala Asn Phe Cys Lys Glu Asn Met Ala Gly Lys Gly Phe Ser
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Tyr Trp Val Trp Leu Asp Asn Ile Ile Asp Leu Val Lys Lys Tyr Ile
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Leu Ala Leu Trp Asn Glu Gly Tyr Ile Met Gly Phe Ile Ser Lys Glu
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Arg Glu Arg Ala Ile Leu Ser Thr Lys Pro Pro Gly Thr Phe Leu Leu
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Arg Phe Ser Glu Ser Ser Lys Glu Gly Gly Val Thr Phe Thr Trp Val
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Glu Lys Asp Ile Ser Gly Lys Thr Gln Ile Gln Ser Val Glu Pro Tyr
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Thr Lys Gln Gln Leu Asn Asn Met Ser Phe Ala Glu Ile Ile Met Gly
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Tyr Lys Ile Met Asp Ala Thr Asn Ile Leu Leu Ser Pro Leu Val Tyr
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Leu Tyr Pro Asp Ile Pro Lys Glu Glu Ala Phe Gly Lys Tyr Cys Arg
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Pro Glu Ser Gln Glu His Pro Glu Ala Asp Pro Gly Ser Ala Ala Pro
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Tyr Leu Lys Thr Lys Phe Ile Cys Val Thr Pro Thr Thr Cys Ser Asn
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Thr Ile Asp Leu Pro Met Ser Pro Arg Ala Leu Asp Ser Leu Met Gln
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 Pro Met
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 Met Ser Gln Trp Asn Gln Val Gln Gln Leu
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 Glu Ile Lys Phe Leu Glu Gln Val Asp Gln Phe Tyr Asp Asp Asn Phe
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 Pro Met Glu Ile Arg His Leu Leu Ala Gln Trp Ile Glu Asn Gln Asp
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 Trp Glu Ala Ala Ser Asn Asn Glu Thr Met Ala Thr Ile Leu Leu Gln
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 Asn Leu Leu Ile Gln Leu Asp Glu Gln Leu Gly Arg Val Ser Lys Glu
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 Lys Asn Leu Leu Leu Ile His Asn Leu Lys Arg Ile Arg Lys Val Leu
 75 80 85 90
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 Gln Gly Lys Phe His Gly Asn Pro Met His Val Ala Val Val Ile Ser
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 aac tgt tta agg gaa gag agg aga ata ttg gct gca gcc aac atg cct 447
 Asn Cys Leu Arg Glu Glu Arg Arg Ile Leu Ala Ala Ala Asn Met Pro
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 Val Gln Gly Pro Leu Glu Lys Ser Leu Gln Ser Ser Ser Val Ser Glu
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Lys Asp Lys Ala Phe Gly Lys His Tyr Ser Ser Gln Pro Cys Glu Val
670 675 680

tca aga cca aca gaa agg ggt gac aaa ggt tat gtt cct tct gtt ttt 2175
Ser Arg Pro Thr Glu Arg Gly Asp Lys Gly Tyr Val Pro Ser Val Phe
685 690 695

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Ile Pro Ile Ser Thr Ile Arg Ser Asp Ser Thr Glu Pro His Ser Pro
700 705 710

tca gac ctt ctt ccc atg tct cca agt gtg tat gcg gtg ttg aga gaa 2271
Ser Asp Leu Leu Pro Met Ser Pro Ser Val Tyr Ala Val Leu Arg Glu
715 720 725 730

aac ctg agt ccc aca aca att gaa act gca atg aag tct cct tat tct 2319
Asn Leu Ser Pro Thr Thr Ile Glu Thr Ala Met Lys Ser Pro Tyr Ser
735 740 745

gct gaa tga caggataaac tctgacgcac caagaaagga agcaaatagaa 2369
Ala Glu *

aaagttaaaa gactgttctt tgcccaataa ccacatttta tttcttcagc tttgtaaata 2428
ccaggttcta ggaaatgttt gacatctgaa gctctcttca cactcccggtg gcactcctca 2488
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<212> PRT

<213> Homo sapiens

<400> 8

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Leu Leu Ala Gln Trp Ile Glu Asn Gln Asp Trp Glu Ala Ala Ser Asn
35 40 45

Asn	Glu	Thr	Met	Ala	Thr	Ile	Leu	Leu	Gln	Asn	Leu	Leu	Ile	Gln	Leu
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Asp	Glu	Gln	Leu	Gly	Arg	Val	Ser	Lys	Glu	Lys	Asn	Leu	Leu	Leu	Ile
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His	Asn	Leu	Lys	Arg	Ile	Arg	Lys	Val	Leu	Gln	Gly	Lys	Phe	His	Gly
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Asn	Pro	Met	His	Val	Ala	Val	Val	Ile	Ser	Asn	Cys	Leu	Arg	Glu	Glu
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Arg	Arg	Ile	Leu	Ala	Ala	Ala	Asn	Met	Pro	Val	Gln	Gly	Pro	Leu	Glu
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Thr	Lys	Tyr	Leu	Glu	Asp	Leu	Gln	Asp	Glu	Phe	Asp	Tyr	Arg	Tyr	Lys
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Gln	Glu	Val	Leu	Thr	Leu	Gln	Glu	Met	Leu	Asn	Ser	Leu	Asp	Phe	Lys
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Arg	Lys	Glu	Ala	Leu	Ser	Lys	Met	Thr	Gln	Ile	Ile	His	Glu	Thr	Asp
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Gln	Leu	Gln	Asn	Cys	Phe	Thr	Leu	Leu	Ala	Glu	Ser	Leu	Phe	Gln	Leu
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Thr	Phe	Leu	Ile	Tyr	Asn	Leu	Phe	Lys	Asn	Ser	Phe	Val	Val	Glu	Arg
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Gln	Pro	Cys	Met	Pro	Thr	His	Pro	Gln	Arg	Pro	Leu	Val	Leu	Lys	Thr
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Thr	Leu	Ser	Asn	Arg	Arg	Phe	Val	Leu	Cys	Gly	Thr	Asn	Val	Lys	Ala
	370					375					380				
Met	Ser	Ile	Glu	Glu	Ser	Ser	Asn	Gly	Ser	Leu	Ser	Val	Glu	Phe	Arg
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His	Leu	Gln	Pro	Lys	Glu	Met	Lys	Ser	Ser	Ala	Gly	Gly	Lys	Gly	Asn
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Glu	Gly	Cys	His	Met	Val	Thr	Glu	Glu	Leu	His	Ser	Ile	Thr	Phe	Glu
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Thr	Gln	Ile	Cys	Leu	Tyr	Gly	Leu	Thr	Ile	Asp	Leu	Glu	Thr	Ser	Ser
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Leu	Pro	Val	Val	Met	Ile	Ser	Asn	Val	Ser	Gln	Leu	Pro	Asn	Ala	Trp
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Val	Phe	Phe	Asn	Asn	Pro	Pro	Pro	Ala	Thr	Leu	Ser	Gln	Leu	Leu	Glu
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Val	Met	Ser	Trp	Gln	Phe	Ser	Ser	Tyr	Val	Gly	Arg	Gly	Leu	Asn	Ser
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Gln Pro Trp Glu Phe Leu Val Gly Ser Asp Ala Phe Cys Cys Asn Leu
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gag agc ata tat cag agg gac ccc ctg aag ctg gtg gcc act ttc aga Glu Ser Ile Tyr Gln Arg Asp Pro Leu Lys Leu Val Ala Thr Phe Arg 85 90 95 100	465
caa ata ctt caa gga gag aaa aaa gct gtt atg gaa cag ttc cgc cac Gln Ile Leu Gln Gly Glu Lys Lys Ala Val Met Glu Gln Phe Arg His 105 110 115	513
ttg cca atg cct ttc cac tgg aag cag gaa gaa ctc aag ttt aag aca Leu Pro Met Pro Phe His Trp Lys Gln Glu Glu Leu Lys Phe Lys Thr 120 125 130	561
ggc ttg cgg agg ctg cag cac cga gta ggg gag atc cac ctt ctc cga Gly Leu Arg Arg Leu Gln His Arg Val Gly Glu Ile His Leu Leu Arg 135 140 145	609
gaa gcc ctg cag aag ggg gct gag gct ggc caa gtg tct ctg cac agc Glu Ala Leu Gln Lys Gly Ala Glu Ala Gly Gln Val Ser Leu His Ser 150 155 160	657
ttg ata gaa act cct gct aat ggg act ggg cca agt gag gcc ctg gcc Leu Ile Glu Thr Pro Ala Asn Gly Thr Gly Pro Ser Glu Ala Leu Ala 165 170 175 180	705
atg cta ctg cag gag acc act gga gag cta gag gca gcc aaa gcc cta Met Leu Leu Gln Glu Thr Thr Gly Glu Leu Glu Ala Ala Lys Ala Leu 185 190 195	753
gtg ctg aag agg atc cag att tgg aaa cgg cag cag cag ctg gca ggg Val Leu Lys Arg Ile Gln Ile Trp Lys Arg Gln Gln Gln Leu Ala Gly 200 205 210	801
aat ggc gca ccg ttt gag gag agc ctg gcc cca ctc cag gag agg tgt Asn Gly Ala Pro Phe Glu Glu Ser Leu Ala Pro Leu Gln Glu Arg Cys 215 220 225	849
gaa agc ctg gtg gac att tat tcc cag cta cag cag gag gta ggg gcg Glu Ser Leu Val Asp Ile Tyr Ser Gln Leu Gln Gln Glu Val Gly Ala 230 235 240	897
gct ggt ggg gag ctt gag ccc aag acc cgg gca tcg ctg act ggc cgg Ala Gly Gly Glu Leu Glu Pro Lys Thr Arg Ala Ser Leu Thr Gly Arg 245 250 255 260	945
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aag cag ccc ccc cag gta ctg aag act cag acc aag ttc cag gct gga Lys Gln Pro Pro Gln Val Leu Lys Thr Gln Thr Lys Phe Gln Ala Gly 280 285 290	1041
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cgg aag ggc act gag tct gtc aca gag gag aag tgc gct gtg ctc ttc Arg Lys Gly Thr Glu Ser Val Thr Glu Glu Lys Cys Ala Val Leu Phe 375 380 385	1329
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gcc ctg tct ctg ccc ctg gtg gtc atc gtc cat ggc aac caa gac aac Ala Leu Ser Leu Pro Leu Val Val Ile Val His Gly Asn Gln Asp Asn 405 410 415 420	1425
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ctg ctc cca gag cac ttc ctc ttc ctg gcc cag aag atc ttc aat gac Leu Leu Pro Glu His Phe Leu Phe Leu Ala Gln Lys Ile Phe Asn Asp 470 475 480	1617
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Trp Phe Asp Gly Val Leu Asp Leu Thr Lys Arg Cys Leu Arg Ser Tyr	
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Trp Ser Asp Arg Leu Ile Ile Gly Phe Ile Ser Lys Gln Tyr Val Thr	
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Ser Leu Leu Leu Asn Glu Pro Asp Gly Thr Phe Leu Leu Arg Phe Ser	
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Asp Ser Glu Ile Gly Gly Ile Thr Ile Ala His Val Ile Arg Gly Gln	
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Asp Gly Ser Pro Gln Ile Glu Asn Ile Gln Pro Phe Ser Ala Lys Asp	
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Lys Asn Leu Tyr Pro Lys Lys Pro Lys Asp Glu Ala Phe Arg Ser His	
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Tyr Lys Pro Glu Gln Met Gly Lys Asp Gly Arg Gly Tyr Val Pro Ala	
630 635 640	
acc atc aag atg acc gtg gaa agg gac caa cca ctt cct acc cca gag	2145
Thr Ile Lys Met Thr Val Glu Arg Asp Gln Pro Leu Pro Thr Pro Glu	
645 650 655 660	
ctc cag atg cct acc atg gtg cct tct tat gac ctt gga atg gcc cct	2193
Leu Gln Met Pro Thr Met Val Pro Ser Tyr Asp Leu Gly Met Ala Pro	
665 670 675	
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Asp Ser Ser Met Ser Met Gln Leu Gly Pro Asp Met Val Pro Gln Val	
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Tyr Pro Pro His Ser His Ser Ile Pro Pro Tyr Gln Gly Leu Ser Pro	
695 700 705	
gaa gaa tca gtc aac gtg ttg tca gcc ttc cag gag cct cac ctg cag	2337
Glu Glu Ser Val Asn Val Leu Ser Ala Phe Gln Glu Pro His Leu Gln	
710 715 720	

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atg ccc ccc agc ctg ggc cag atg agc ctg ccc ttt gac cag cct cac      2385
Met Pro Pro Ser Leu Gly Gln Met Ser Leu Pro Phe Asp Gln Pro His
725                      730                      735                      740

ccc cag ggc ctg ctg ccg tgc cag cct cag gag cat gct gtg tcc agc      2433
Pro Gln Gly Leu Leu Pro Cys Gln Pro Gln Glu His Ala Val Ser Ser
745                      750                      755

cct gac ccc ctg ctc tgc tca gat gtg acc atg gtg gaa gac agc tgc      2481
Pro Asp Pro Leu Leu Cys Ser Asp Val Thr Met Val Glu Asp Ser Cys
760                      765                      770

ctg agc cag cca gtg aca ggc ttt cct cag ggc act tgg att ggt gaa      2529
Leu Ser Gln Pro Val Thr Ala Phe Pro Gln Gly Thr Trp Ile Gly Glu
775                      780                      785

gac ata ttc cct cct ctg ctg cct ccc act gaa cag gac ctc act aag      2577
Asp Ile Phe Pro Pro Leu Leu Pro Pro Thr Glu Gln Asp Leu Thr Lys
790                      795                      800

ctt ctc ctg gag ggg caa ggg gag tgc ggg gga ggg tcc ttg ggg gca      2625
Leu Leu Leu Glu Gly Gln Gly Glu Ser Gly Gly Gly Ser Leu Gly Ala
805                      810                      815

cag ccc ctc ctg cag ccc tcc cac tat ggg caa tct ggg atc tca atg      2673
Gln Pro Leu Leu Gln Pro Ser His Tyr Gly Gln Ser Gly Ile Ser Met
825                      830                      835

tcc cac atg gac cta agg gcc aac ccc agt tgg tga tcccagctgg      2719
Ser His Met Asp Leu Arg Ala Asn Pro Ser Trp *
840                      845

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Trp Leu Glu Ser Gln Pro Trp Glu Phe Leu Val Gly Ser Asp Ala Phe
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Cys Cys Asn Leu Ala Ser Ala Leu Leu Ser Asp Thr Val Gln His Leu
50          55          60
Gln Ala Ser Val Gly Glu Gln Gly Glu Gly Ser Thr Ile Leu Gln His
65          70          75          80
Ile Ser Thr Leu Glu Ser Ile Tyr Gln Arg Asp Pro Leu Lys Leu Val
85          90          95

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Lys	Phe	Lys	Thr	Gly	Leu	Arg	Arg	Leu	Gln	His	Arg	Val	Gly	Glu	Ile
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His	Leu	Leu	Arg	Glu	Ala	Leu	Gln	Lys	Gly	Ala	Glu	Ala	Gly	Gln	Val
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Ser	Leu	His	Ser	Leu	Ile	Glu	Thr	Pro	Ala	Asn	Gly	Thr	Gly	Pro	Ser
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Glu	Ala	Leu	Ala	Met	Leu	Leu	Gln	Glu	Thr	Thr	Gly	Glu	Leu	Glu	Ala
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Ala	Lys	Ala	Leu	Val	Leu	Lys	Arg	Ile	Gln	Ile	Trp	Lys	Arg	Gln	Gln
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Gln	Leu	Ala	Gly	Asn	Gly	Ala	Pro	Phe	Glu	Glu	Ser	Leu	Ala	Pro	Leu
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Gln	Glu	Arg	Cys	Glu	Ser	Leu	Val	Asp	Ile	Tyr	Ser	Gln	Leu	Gln	Gln
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Glu	Val	Gly	Ala	Ala	Gly	Gly	Glu	Leu	Glu	Pro	Lys	Thr	Arg	Ala	Ser
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Leu	Thr	Gly	Arg	Leu	Asp	Glu	Val	Leu	Arg	Thr	Leu	Val	Thr	Ser	Cys
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Phe	Leu	Val	Glu	Lys	Gln	Pro	Pro	Gln	Val	Leu	Lys	Thr	Gln	Thr	Lys
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Pro	Ala	Lys	Pro	Pro	Leu	Val	Arg	Ala	Asp	Met	Val	Thr	Glu	Lys	Gln
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Ala	Arg	Glu	Leu	Ser	Val	Pro	Gln	Gly	Pro	Gly	Ala	Gly	Ala	Glu	Ser
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Thr	Gly	Glu	Ile	Ile	Asn	Asn	Thr	Val	Pro	Leu	Glu	Asn	Ser	Ile	Pro
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Gly	Asn	Cys	Cys	Ser	Ala	Leu	Phe	Lys	Asn	Leu	Leu	Leu	Lys	Lys	Ile
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Lys	Arg	Cys	Glu	Arg	Lys	Gly	Thr	Glu	Ser	Val	Thr	Glu	Glu	Lys	Cys
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Ala	Val	Leu	Phe	Ser	Ala	Ser	Phe	Thr	Leu	Gly	Pro	Gly	Lys	Leu	Pro
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Ile	Gln	Leu	Gln	Ala	Leu	Ser	Leu	Pro	Leu	Val	Val	Ile	Val	His	Gly
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Asn	Gln	Asp	Asn	Asn	Ala	Lys	Ala	Thr	Ile	Leu	Trp	Asp	Asn	Ala	Phe
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Ser	Glu	Met	Asp	Arg	Val	Pro	Phe	Val	Val	Ala	Glu	Arg	Val	Pro	Trp
		435					440					445			
Glu	Lys	Met</													

Leu Arg Phe Ser Asp Ser Glu Ile Gly Gly Ile Thr Ile Ala His Val
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 Ile Arg Gly Gln Asp Gly Ser Pro Gln Ile Glu Asn Ile Gln Pro Phe
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 Ser Ala Lys Asp Leu Ser Ile Arg Ser Leu Gly Asp Arg Ile Arg Asp
 595 600 605
 Leu Ala Gln Leu Lys Asn Leu Tyr Pro Lys Lys Pro Lys Asp Glu Ala
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 Phe Arg Ser His Tyr Lys Pro Glu Gln Met Gly Lys Asp Gly Arg Gly
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 645 650 655
 Pro Thr Pro Glu Leu Gln Met Pro Thr Met Val Pro Ser Tyr Asp Leu
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 Gly Met Ala Pro Asp Ser Ser Met Ser Met Gln Leu Gly Pro Asp Met
 675 680 685
 Val Pro Gln Val Tyr Pro Pro His Ser His Ser Ile Pro Pro Tyr Gln
 690 695 700
 Gly Leu Ser Pro Glu Glu Ser Val Asn Val Leu Ser Ala Phe Gln Glu
 705 710 715 720
 Pro His Leu Gln Met Pro Pro Ser Leu Gly Gln Met Ser Leu Pro Phe
 725 730 735
 Asp Gln Pro His Pro Gln Gly Leu Leu Pro Cys Gln Pro Gln Glu His
 740 745 750
 Ala Val Ser Ser Pro Asp Pro Leu Leu Cys Ser Asp Val Thr Met Val
 755 760 765
 Glu Asp Ser Cys Leu Ser Gln Pro Val Thr Ala Phe Pro Gln Gly Thr
 770 775 780
 Trp Ile Gly Glu Asp Ile Phe Pro Pro Leu Leu Pro Pro Thr Glu Gln
 785 790 795 800
 Asp Leu Thr Lys Leu Leu Leu Glu Gly Gln Gly Glu Ser Gly Gly Gly
 805 810 815
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 Val Ser Thr Ala Ala Glu Pro Arg Arg Arg Pro Glu Pro Ser Ser Ser
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 tcc tcc tcc tcg ccc gcg gcc ccc gcg cgc ccg cgg ccg tgc ccc gcg 145
 Ser Ser Ser Pro Ala Ala Pro Ala Arg Pro Arg Pro Cys Pro Ala
 30 35 40

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His Ala Asp Tyr Arg Arg Ile Thr Arg Ala Ser Ala Leu Leu Asp Ala
                      65                      70                      75

tgc gga ttc tac tgg ggg ccc ctg agc gtg cac ggg gcg cac gag cgg      289
Cys Gly Phe Tyr Trp Gly Pro Leu Ser Val His Gly Ala His Glu Arg
                      80                      85                      90

ctg cgc gcc gag ccc gtg ggc acc ttc ctg gtg cgc gac agc cgc cag      337
Leu Arg Ala Glu Pro Val Gly Thr Phe Leu Val Arg Asp Ser Arg Gln
                      95                      100                      105

cgg aac tgc ttt ttc gcc ctt agc gtg aag atg gcc tcg gga ccc acg      385
Arg Asn Cys Phe Phe Ala Leu Ser Val Lys Met Ala Ser Gly Pro Thr
  110                      115                      120

agc atc cgc gtg cac ttt cag gcc ggc cgc ttt cac ctg gat ggc agc      433
Ser Ile Arg Val His Phe Gln Ala Gly Arg Phe His Leu Asp Gly Ser
  125                      130                      135                      140

cgc gag agc ttc gac tgc ctc ttc gag ctg ctg gag cac tac gtg gcg      481
Arg Glu Ser Phe Asp Cys Leu Phe Glu Leu Leu Glu His Tyr Val Ala
                      145                      150                      155

gcg ccg cgc cgc atg ctg ggg gcc ccg ctg cgc cag cgc cgc gtg cgg      529
Ala Pro Arg Arg Met Leu Gly Ala Pro Leu Arg Gln Arg Arg Val Arg
                      160                      165                      170

ccg ctg cag gag ctg tgc cgc cag cgc atc gtg gcc acc gtg ggc cgc      577
Pro Leu Gln Glu Leu Cys Arg Gln Arg Ile Val Ala Thr Val Gly Arg
                      175                      180                      185

gag aac ctg gct cgc atc ccc ctc aac ccc gtc ctc cgc gac tac ctg      625
Glu Asn Leu Ala Arg Ile Pro Leu Asn Pro Val Leu Arg Asp Tyr Leu
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agc tcc ttc ccc ttc cag att tga ccggcagcgc ccgccgtgca cgcagcatta      679
Ser Ser Phe Pro Phe Gln Ile *
  205                      210

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cgaggccgca gaccccttct cacctcttga gggggtcctc cccctcctgg tgctccctct      859
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 Arg Arg Ile Thr Arg Ala Ser Ala Leu Leu Asp Ala Cys Gly Phe Tyr
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 Trp Gly Pro Leu Ser Val His Gly Ala His Glu Arg Leu Arg Ala Glu
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 Pro Val Gly Thr Phe Leu Val Arg Asp Ser Arg Gln Arg Asn Cys Phe
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 Phe Ala Leu Ser Val Lys Met Ala Ser Gly Pro Thr Ser Ile Arg Val
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 His Phe Gln Ala Gly Arg Phe His Leu Asp Gly Ser Arg Glu Ser Phe
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 Asp Thr Ser Leu Arg Leu Lys Thr Phe Ser Ser Lys Ser Glu Tyr Gln
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 ctg gtg gtg aac gca gtg cgc aag ctg cag gag agc ggc ttc tac tgg 144
 Leu Val Val Asn Ala Val Arg Lys Leu Gln Glu Ser Gly Phe Tyr Trp
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 Ser Ala Val Thr Gly Gly Glu Ala Asn Leu Leu Leu Ser Ala Glu Pro
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 gcc ggc acc ttt ctg atc cgc gac agc tcg gac cag cgc cac ttc ttc 240
 Ala Gly Thr Phe Leu Ile Arg Asp Ser Ser Asp Gln Arg His Phe Phe
 65 70 75 80

acg ctc agc gtc aag acc cag tct ggg acc aag aac ctg cgc atc cag 288
 Thr Leu Ser Val Lys Thr Gln Ser Gly Thr Lys Asn Leu Arg Ile Gln
 85 90 95
 tgt gag ggg ggc agc ttc tct ctg cag agc gat ccc cgg agc acg cag 336
 Cys Glu Gly Gly Ser Phe Ser Leu Gln Ser Asp Pro Arg Ser Thr Gln
 100 105 110
 ccc gtg ccc cgc ttc gac tgc gtg ctc aag ctg gtg cac cac tac atg 384
 Pro Val Pro Arg Phe Asp Cys Val Leu Lys Leu Val His His Tyr Met
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 Pro Pro Pro Gly Ala Pro Ser Phe Pro Ser Pro Pro Thr Glu Pro Ser
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Ser Ile Tyr Gln Asn Gly Asp Ile Ser Gly Asn Ala Asn Ser Met Lys															
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Gln Ile Asp Pro Val Leu Gln Val Tyr Leu Tyr His Ser Leu Gly Lys															
	35	40	45												
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Ser Glu Ala Asp Tyr Leu Thr Phe Pro Ser Gly Glu Tyr Val Ala Glu															
	50	55	60				65								

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Pro Asn His Val Phe His Ile Asp Glu Ser Thr Arg His Asn Val Leu	
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Tyr Arg Ile Arg Phe Tyr Phe Pro Arg Trp Tyr Cys Ser Gly Ser Asn	
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Arg Ala Tyr Arg His Gly Ile Ser Arg Gly Ala Glu Ala Pro Leu Leu	
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Val His Gly Trp Ile Lys Val Pro Val Thr His Glu Thr Gln Glu Glu	
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Arg Lys Arg Ile Arg Tyr Arg Phe Arg Arg Phe Ile Gln Gln Phe Ser	
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275 280 285	

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Phe	Phe	Glu	Ala	Ala	Ser	Met	Met	Ser	Lys	Leu	Ser	His	Lys	His	Leu	
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Gln	Glu	Phe	Val	Lys	Phe	Gly	Ser	Leu	Asp	Thr	Tyr	Leu	Lys	Lys	Asn	
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Lys	Asn	Cys	Ile	Asn	Ile	Leu	Trp	Lys	Leu	Glu	Val	Ala	Lys	Gln	Leu	
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Ala	Trp	Ala	Met	His	Phe	Leu	Glu	Glu	Asn	Thr	Leu	Ile	His	Gly	Asn	
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Val	Cys	Ala	Lys	Asn	Ile	Leu	Leu	Ile	Arg	Glu	Glu	Asp	Arg	Lys	Thr	
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755 760 765	
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His Gln Leu Pro Ala Pro Lys Trp Ala Glu Leu Ala Asn Leu Ile Asn	
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<212> PRT

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Lys Gln Ile Asp Pro Val Leu Gln Val Tyr Leu Tyr His Ser Leu Gly
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Lys Ser Glu Ala Asp Tyr Leu Thr Phe Pro Ser Gly Glu Tyr Val Ala
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Glu Glu Ile Cys Ile Ala Ala Ser Lys Ala Cys Gly Ile Thr Pro Val
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Tyr His Asn Met Phe Ala Leu Met Ser Glu Thr Glu Arg Ile Trp Tyr
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Pro Pro Asn His Val Phe His Ile Asp Glu Ser Thr Arg His Asn Val
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Phe Val His Gly Trp Ile Lys Val Pro Val Thr His Glu Thr Gln Glu
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Glu Cys Leu Gly Met Ala Val Leu Asp Met Met Arg Ile Ala Lys Glu
180     185     190
Asn Asp Gln Thr Pro Leu Ala Ile Tyr Asn Ser Ile Ser Tyr Lys Thr
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Phe Leu Pro Lys Cys Ile Arg Ala Lys Ile Gln Asp Tyr His Ile Leu
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Thr Arg Lys Arg Ile Arg Tyr Arg Phe Arg Arg Phe Ile Gln Gln Phe
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Ser Gln Cys Lys Ala Thr Ala Arg Asn Leu Lys Leu Lys Tyr Leu Ile
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Lys Glu Pro Gly Ser Gly Pro Ser Gly Glu Glu Ile Phe Ala Thr Ile
275     280     285
Ile Ile Thr Gly Asn Gly Gly Ile Gln Trp Ser Arg Gly Lys His Lys
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Glu Ser Glu Thr Leu Thr Glu Gln Asp Leu Gln Leu Tyr Cys Asp Phe
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Ser Asn Ile Ile Asp Val Ser Ile Lys Gln Ala Asn Gln Glu Gly Ser
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Asn Glu Ser Arg Val Val Thr Ile His Lys Gln Asp Gly Lys Asn Leu
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Glu	Ile	Glu	Leu	Ser	Ser	Leu	Arg	Glu	Ala	Leu	Ser	Phe	Val	Ser	Leu	355	360	365
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Lys	Glu	Val	Ala	Pro	Pro	Ala	Val	Leu	Glu	Asn	Ile	Gln	Ser	Asn	Cys	385	390	395
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Gly	Asn	Gln	Thr	Gly	Leu	Tyr	Val	Leu	Arg	Cys	Ser	Pro	Lys	Asp	Phe	420	425	430
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Tyr	Gln	Met	Glu	Thr	Val	Arg	Ser	Asp	Asn	Ile	Ile	Phe	Gln	Phe	Thr	485	490	495
Lys	Cys	Cys	Pro	Pro	Lys	Pro	Lys	Asp	Lys	Ser	Asn	Leu	Leu	Val	Phe	500	505	510
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Pro	Thr	His	Met	Asn	Gln	Met	Val	Phe	His	Lys	Ile	Arg	Asn	Glu	Asp	530	535	540
Leu	Ile	Phe	Asn	Glu	Ser	Leu	Gly	Gln	Gly	Thr	Phe	Thr	Lys	Ile	Phe	545	550	555
Lys	Gly	Val	Arg	Arg	Glu	Val	Gly	Asp	Tyr	Gly	Gln	Leu	His	Glu	Thr	565	570	575
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Ser	Phe	Phe	Glu	Ala	Ala	Ser	Met	Met	Ser	Lys	Leu	Ser	His	Lys	His	595	600	605
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Asn	Val	Cys	Ala	Lys	Asn	Ile	Leu	Leu	Ile	Arg	Glu	Glu	Asp	Arg	Lys	675	680	685
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gggagc atg cct ctg cgc cac tgg ggg atg gcc agg ggc agt aag ccc	348
Met Pro Leu Arg His Trp Gly Met Ala Arg Gly Ser Lys Pro	
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Val Gly Asp Gly Ala Gln Pro Met Ala Ala Met Gly Gly Leu Lys Val	
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Leu Leu His Trp Ala Gly Pro Gly Gly Gly Glu Pro Trp Val Thr Phe	
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Ser Glu Ser Ser Leu Thr Ala Glu Glu Val Cys Ile His Ile Ala His	
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Lys Val Gly Ile Thr Pro Pro Cys Phe Asn Leu Phe Ala Leu Phe Asp	
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Ala Gln Ala Gln Val Trp Leu Pro Pro Asn His Ile Leu Glu Ile Pro	
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Arg Asp Ala Ser Leu Met Leu Tyr Phe Arg Ile Arg Phe Tyr Phe Arg	
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 Gly Ile Thr Pro Pro Cys Phe Asn Leu Phe Ala Leu Phe Asp Ala Gln
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 Ala Gln Val Trp Leu Pro Pro Asn His Ile Leu Glu Ile Pro Arg Asp
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 Val Asn Asp Val Ala Ser Leu Trp Glu Leu Ser Thr Glu Glu Glu Ile
 165 170 175
 His His Phe Lys Asn Glu Ser Leu Gly Met Ala Phe Leu His Leu Cys
 180 185 190
 His Leu Ala Leu Arg His Gly Ile Pro Leu Glu Glu Val Ala Lys Lys
 195 200 205

Thr	Ser	Phe	Lys	Asp	Cys	Ile	Pro	Arg	Ser	Phe	Arg	Arg	His	Ile	Arg
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Gln	His	Ser	Ala	Leu	Thr	Arg	Leu	Arg	Leu	Arg	Asn	Val	Phe	Arg	Arg
225					230					235					240
Phe	Leu	Arg	Asp	Phe	Gln	Pro	Gly	Arg	Leu	Ser	Gln	Gln	Met	Val	Met
			245						250					255	
Val	Lys	Tyr	Leu	Ala	Thr	Leu	Glu	Arg	Leu	Ala	Pro	Arg	Phe	Gly	Thr
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Glu	Arg	Val	Pro	Val	Cys	His	Leu	Arg	Leu	Leu	Ala	Gln	Ala	Glu	Gly
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Glu	Pro	Cys	Tyr	Ile	Arg	Asp	Ser	Gly	Val	Ala	Pro	Thr	Asp	Pro	Gly
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Pro	Glu	Ser	Ala	Ala	Gly	Pro	Pro	Thr	His	Glu	Val	Leu	Val	Thr	Gly
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Thr	Gly	Gly	Ile	Gln	Trp	Trp	Pro	Val	Glu	Glu	Glu	Val	Asn	Lys	Glu
			325						330					335	
Glu	Gly	Ser	Ser	Gly	Ser	Ser	Gly	Arg	Asn	Pro	Gln	Ala	Ser	Leu	Phe
		340					345					350			
Gly	Lys	Lys	Ala	Lys	Ala	His	Lys	Ala	Phe	Gly	Gln	Pro	Ala	Asp	Arg
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Pro	Arg	Glu	Pro	Leu	Trp	Ala	Tyr	Phe	Cys	Asp	Phe	Arg	Asp	Ile	Thr
	370					375					380				
His	Val	Val	Leu	Lys	Glu	His	Cys	Val	Ser	Ile	His	Arg	Gln	Asp	Asn
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Lys	Cys	Leu	Glu	Leu	Ser	Leu	Pro	Ser	Arg	Ala	Ala	Ala	Leu	Ser	Phe
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		420						425					430		
Tyr	Leu	Cys	His	Glu	Val	Ala	Pro	Arg	Leu	Val	Met	Ser	Ile	Arg	
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Asp	Gly	Ile	His	Gly	Pro	Leu	Leu	Glu	Pro	Phe	Val	Gln	Ala	Lys	Leu
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Arg	Pro	Glu	Asp	Gly	Leu	Tyr	Leu	Ile	His	Trp	Ser	Thr	Ser	His	Pro
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Tyr	Arg	Leu	Ile	Leu	Thr	Val	Ala	Gln	Arg	Ser	Gln	Ala	Pro	Asp	Gly
			485						490					495	
Met	Gln	Ser	Leu	Arg	Leu	Arg	Lys	Phe	Pro	Ile	Glu	Gln	Gln	Asp	Gly
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Ala	Phe	Val	Leu	Glu	Gly	Trp	Gly	Arg	Ser	Phe	Pro	Ser	Val	Arg	Glu
	515						520					525			
Leu	Gly	Ala	Ala	Leu	Gln	Gly	Cys	Leu	Leu	Arg	Ala	Gly	Asp	Asp	Cys
	530					535					540				
Phe	Ser	Leu	Arg	Arg	Cys	Cys	Leu	Pro	Gln	Pro	Gly	Glu	Thr	Ser	Asn
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Leu	Ile	Ile	Met	Arg	Gly	Ala	Arg	Ala	Ser	Pro	Arg	Thr	Leu	Asn	Leu
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Ser	Gln	Leu	Ser	Phe	His	Arg	Val	Asp	Gln	Lys	Glu	Ile	Thr	Gln	Leu
		580						585					590		
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Arg	Val	Glu	Gly	Ser	Gly	Asp	Pro	Glu	Glu	Gly	Lys	Met	Asp	Asp	Glu
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Leu	Lys	Val	Leu	Asp	Pro	Ser	His	His	Asp	Ile	Ala	Leu	Ala	Phe	Tyr
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Glu	Thr	Ala	Ser	Leu	Met	Ser	Gln	Val	Ser	His	Thr	His	Leu	Ala	Phe
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Val	His	Gly	Val	Cys	Val	Arg	Gly	Pro	Glu	Asn	Ser	Met	Val	Thr	Glu
		675					680					685			
Tyr	Val	Glu	His	Gly	Pro	Leu	Asp	Val	Trp	Leu	Arg	Arg	Glu	Arg	Gly
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His	Val	Pro	Met	Ala	Trp	Lys	Met	Val	Val	Ala	Gln	Gln	Leu	Ala	Ser
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Ala	Leu	Ser	Tyr	Leu	Glu	Asn	Lys	Asn	Leu	Val	His	Gly	Asn	Val	Cys
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			740					745					750		
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	755					760						765			
Arg	Glu	Glu	Arg	Val	Glu	Arg	Ile	Pro	Trp	Leu	Ala	Pro	Glu	Cys	Leu
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	850					855						860			
Asp	Leu	Thr	Arg	Val	Gln	Pro	His	Asn	Leu	Ala	Asp	Val	Leu	Thr	Val
865					870					875					880
Asn	Arg	Asp	Ser	Pro	Ala	Val	Gly	Pro	Thr	Thr	Phe	His	Lys	Arg	Tyr
				885					890					895	
Leu	Lys	Lys	Ile	Arg	Asp	Leu	Gly	Glu	Gly	His	Phe	Gly	Lys	Val	Ser
			900					905					910		
Leu	Tyr	Cys	Tyr	Asp	Pro	Thr	Asn	Asp	Gly	Thr	Gly	Glu	Met	Val	Ala
	915						920					925			
Val	Lys	Ala	Leu	Lys	Ala	Asp	Cys	Gly	Pro	Gln	His	Arg	Ser	Gly	Trp
	930					935						940			
Lys	Gln	Glu	Ile	Asp	Ile	Leu	Arg	Thr	Leu	Tyr	His	Glu	His	Ile	Ile
945					950					955					960
Lys	Tyr	Lys	Gly	Cys	Cys	Glu	Asp	Gln	Gly	Glu	Lys	Ser	Leu	Gln	Leu
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Val	Met	Glu	Tyr	Val	Pro	Leu	Gly	Ser	Leu	Arg	Asp	Tyr	Leu	Pro	Arg
			980					985					990		
His	Ser	Ile	Gly	Leu	Ala	Gln	Leu	Leu	Leu	Phe	Ala	Gln	Gln	Ile	Cys
	995						1000					1005			
Glu	Gly	Met	Ala	Tyr	Leu	His	Ala	His	Asp	Tyr	Ile	His	Arg	Asp	Leu
	1010					1015					1020				
Ala	Ala														

Pro Asp Lys Cys Pro Cys Glu Val Tyr His Leu Met Lys Asn Cys Trp
 1140 1145 1150
 Glu Thr Glu Ala Ser Phe Arg Pro Thr Phe Glu Asn Leu Ile Pro Ile
 1155 1160 1165
 Leu Lys Thr Val His Glu Lys Tyr Gln Gly Gln Ala Pro Ser Val Phe
 1170 1175 1180
 Ser Val Cys
 1185

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<220>
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<220>
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<400> 44
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<210> 46
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<400> 46
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<210> 47
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<400> 49

ctgtgccgca gcattaagtg 20

<210> 50
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 <220>
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 ggggaagtatt attccagacc aaaggaagca ccagaaccga tggagcttga cgaccctaag 180
 cgaactggat acatcaagac tgagttgatt tctgtgtctg aagtccaccc ttctagactt 240
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 ggggtgttact accaggaaaa agttaatttg gaagaacagg aggaatattt gaaacataaa 180
 ctcatgtgta tctctaacag acagggtggac gagctgcagc agcctctgga gctcaaacag 240
 gattcagagt ccttagaagt gaatgcagag ctcttggttag cacacgacca ggagttgccca 300
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 ctgcagcagc catcagagtc agatttgctt gagg 694

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<220>
 <223> probe

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 tgccgatgtc cccccgact ttagattcat tgatgcagtt tggaaataac ggtgaagggtg 180
 ctgagccctc agcaggaggg cagtttgagt cgctcacgtt tgacatggat ctgacctcgg 240
 agtgtgttac ctcccccatg tgaggagctg aaaccagaag ctgcagagac gtgacttgag 300
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 tttgtgggtc cag 373

<210> 62
 <211> 442
 <212> DNA
 <213> Artificial Sequence

<220>
 <223> probe

<400> 62
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 aagtacctct accctgacat tcccaaagac aaagcctttg gcaaactacta cagctcccag 180
 ccgtgcgaag totcaagacc aaccgaacgg ggagacaagg gttacgtccc ctctgttttt 240
 atccccattt caacaatccg aagcgattcc acggagccac aatctccttc agaccttctc 300
 cccatgtctc caagtgcata tgetgtgctg agagaaaacc tgagcccaac gacaattgaa 360
 actgcaatga attccccata ttctgctgaa tgacgggtgca aacggacact ttaaagaagg 420
 aagcagatga aactggagag tg 442

<210> 63

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<210> 64
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 <212> DNA
 <213> Artificial Sequence

<220>
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<400> 64
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 gcttacagtc tcaactcatt tcgtcgtaaa tgctctcgta acgacattga ttcttctacc 240
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<220>
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 cagatgectg ccattggtgcc accttatgat cttggaatgg ccctgatgc ttccatgcaa 180
 ctcagctcag atatggggta tcctccacag tccatccact catttcagag cctagaagag 240
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 cagataacca tgccctttga ccagcctcac cccaggggtc tgctgcagtg ccagtccag 360
 gaacatgctg tgtccagccc tgaacccatg ctttggtcag atgtgactat ggtagaggac 420
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<211> 381

<212> DNA

<213> Artificial Sequence

<220>

<223> probe

<400> 66

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gatgcctccc	acttctggct	ggagacctca	tcccacctct	caggggtggg	ggtgctcccc	180
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gaattccact	cctacctctc	catgtttaca	tattcccagt	atctttgcac	aaaccagggg	300
tcggggaggg	tctctggctt	cattttttctg	ctgtgcagaa	tatcctattt	tatattttta	360
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<211> 266

<212> DNA

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<400> 67

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(74) Agents: **CORUZZI, Laura, A.** et al.; Pennie & Edmonds LLP, 1155 Avenue of the Americas, New York, NY 10036 (US).

(81) Designated States (*national*): AE, AG, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, BZ, CA, CH, CN, CO, CR, CU, CZ, DE, DK, DM, DZ, EE, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MA, MD, MG, MK, MN, MW, MX, MZ, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, TZ, UA, UG, UZ, VN, YU, ZA, ZW.

(84) Designated States (*regional*): ARIPO patent (GH, GM, KE, LS, MW, MZ, SD, SL, SZ, TZ, UG, ZW), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE, TR), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, GW, ML, MR, NE, SN, TD, TG).

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For two-letter codes and other abbreviations, refer to the "Guidance Notes on Codes and Abbreviations" appearing at the beginning of each regular issue of the PCT Gazette.

(54) Title: ROLES OF JAK/STAT FAMILY MEMBERS IN TOLERANCE INDUCTION

(57) Abstract: The present invention relates to methods and compositions for reducing immune rejection, for example, transplant- or autoimmune disorder-related immune rejection. The present invention also relates to methods and compositions for monitoring transplant acceptance and for monitoring an autoimmune disorder in a subject mammal. The present invention still further relates to methods for identifying compounds that can reduce immune rejection. The present invention is based, in part, on the discovery, demonstrated herein, that immune rejection can be monitored by determining the amount of particular members of the Jak/Stat signal transduction pathway present within an affected tissue (that is, a transplant cell, tissue, organ, or organ system, or a cell, tissue, organ, or organ system that is, or is suspected of, being affected by an autoimmune disorder). The present invention is further based, in part, on the discovery, demonstrated herein, that immune rejection can be reduced and tolerance can be induced by modulating the amount of these particular members of the Jak/Stat signal transduction pathway present, expressed or active within an affected tissue. In particular, the results presented herein demonstrate that immune rejection can be monitored by determining the amount of Stat1 mRNA or protein, Stat2 mRNA or protein, Stat3 mRNA or protein, Stat4 mRNA or protein, Stat6 mRNA or protein, SOCS1 mRNA or protein, or SOCS3 mRNA or protein present, e.g., present in an affected tissue.

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INTERNATIONAL SEARCH REPORT

In **onal Application No**
PCT/US 01/12131

A. CLASSIFICATION OF SUBJECT MATTER
IPC 7 C12Q1/68 G01N33/68

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC 7 C12Q

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

EPO-Internal, WPI Data, PAJ, SEQUENCE SEARCH, MEDLINE, BIOSIS

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☒ Further documents are listed in the continuation of box C.

☒ Patent family members are listed in annex.

* Special categories of cited documents:

A document defining the general state of the art which is not considered to be of particular relevance

E earlier document but published on or after the international filing date

L document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)

O document referring to an oral disclosure, use, exhibition or other means

P document published prior to the international filing date but later than the priority date claimed

T later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention

X document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone

Y document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art.

Z document member of the same patent family

Date of the actual completion of the international search

14 October 2002

Date of mailing of the international search report

25/10/2002

Name and mailing address of the ISA

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Hagenmaier, S

INTERNATIONAL SEARCH REPORT

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INTERNATIONAL SEARCH REPORT

Initial Application No
PCT/US 01/12131

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT		
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Y	<p>KOGLIN J ET AL: "Attenuated cardiac allograft vasculopathy in mice with targeted deletion of the transcription factor STAT4." CIRCULATION. UNITED STATES 7 MAR 2000, vol. 101, no. 9, 7 March 2000 (2000-03-07), pages 1034-1039, XP002216628 ISSN: 1524-4539 the whole document</p> <p>---</p>	1-56
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Form PCT/ISA/210 (continuation of second sheet) (July 1992)

INTERNATIONAL SEARCH REPORT

In International Application No.

PCT/US 01/12131

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

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International Application No.
PCT/US 01/12131

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INTERNATIONAL SEARCH REPORT

Int. Patent Application No

PCT/US 01/12131

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
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A	WO 98 07436 A (VASOGEN INC) 26 February 1998 (1998-02-26) page 9 -page 10 ----	
P,Y	ZHOU P ET AL: "Role of STAT4 and STAT6 signaling in allograft rejection and CTLA4-Ig-mediated tolerance." JOURNAL OF IMMUNOLOGY (BALTIMORE, MD.: 1950) UNITED STATES 15 NOV 2000, vol. 165, no. 10, 15 November 2000 (2000-11-15), pages 5580-5587, XP002216635 ISSN: 0022-1767 the whole document ----	1-56
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Form PCT/ISA/210 (continuation of second sheet) (July 1992)

INTERNATIONAL SEARCH REPORT

national application No.
PCT/US 01/12131

Box I Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)

This International Search Report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:

1. ☐ Claims Nos.:
because they relate to subject matter not required to be searched by this Authority, namely:
2. ☒ Claims Nos.: 57-67
because they relate to parts of the International Application that do not comply with the prescribed requirements to such an extent that no meaningful International Search can be carried out, specifically:
see FURTHER INFORMATION sheet PCT/ISA/210
3. ☐ Claims Nos.:
because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).

Box II Observations where unity of invention is lacking (Continuation of item 2 of first sheet)

This International Searching Authority found multiple inventions in this international application, as follows:

see additional sheet

1. ☐ As all required additional search fees were timely paid by the applicant, this International Search Report covers all searchable claims.
2. ☒ As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.
3. ☐ As only some of the required additional search fees were timely paid by the applicant, this International Search Report covers only those claims for which fees were paid, specifically claims Nos.:
4. ☐ No required additional search fees were timely paid by the applicant. Consequently, this International Search Report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:

Remark on Protest

- ☐ The additional search fees were accompanied by the applicant's protest.
- ☐ No protest accompanied the payment of additional search fees.

FURTHER INFORMATION CONTINUED FROM PCT/ISA/ 210

This International Searching Authority found multiple (groups of) inventions in this international application, as follows:

1. Claims: 1-56 (all partially)

A method for monitoring acceptance of a transplant in a subject mammal that has undergone a transplant, comprising: determining the amount of Stat4 mRNA or Stat4 protein, present in a transplant sample from the subject, a method for monitoring an autoimmune disorder in a subject mammal, comprising : determining the amount of Stat4 mRNA or Stat4 protein, present in a sample mammal from a subject being treated for or suspected of exhibiting the disorder, wherein the sample is obtained from a tissue affected by the disorder as well as a method for identifying a compound to be tested for an ability to reduce immune rejection, comprising determining the amount of Stat4 mRNA or Stat4 protein in T-cell samples.

2. Claims: 1-56 (all partially)

A method for monitoring acceptance of a transplant in a subject mammal that has undergone a transplant, comprising: determining the amount of Stat6 mRNA or Stat6 protein present in a transplant sample from the subject, a method for monitoring an autoimmune disorder in a subject mammal, comprising : determining the amount of Stat6 mRNA or Stat6 protein, present in a sample mammal from a subject being treated for or suspected of exhibiting the disorder, wherein the sample is obtained from a tissue affected by the disorder as well as a method for identifying a compound to be tested for an ability to reduce immune rejection, comprising determining the amount of Stat6 mRNA or Stat6 protein in T-cell samples.

3. Claims: 1-20,23-36, 39-46, 49-54 (all partially)

A method for monitoring acceptance of a transplant in a subject mammal that has undergone a transplant, comprising: determining the amount of SOCS1 mRNA or SOCS1 protein present in a transplant sample from the subject, a method for monitoring an autoimmune disorder in a subject mammal, comprising : determining the amount of SOCS1 mRNA or SOCS1 protein, present in a sample mammal from a subject being treated for or suspected of exhibiting the disorder, wherein the sample is obtained from a tissue affected by the disorder as well as a method for identifying a compound to be tested for an ability to reduce immune rejection, comprising determining the amount of SOCS1 mRNA or SOCS1 protein in T-cell samples.

4. Claims: 1-20,23-36, 39-46, 49-54 (all partially)

FURTHER INFORMATION CONTINUED FROM PCT/SA/ 210

A method for monitoring acceptance of a transplant in a subject mammal that has undergone a transplant, comprising: determining the amount of SOCS3 mRNA or SOCS3 protein, present in a transplant sample from the subject, a method for monitoring an autoimmune disorder in a subject mammal, comprising determining the amount SOCS3 mRNA or SOCS3 protein, present in a sample mammal from a subject being treated for or suspected of exhibiting the disorder, wherein the sample is obtained from a tissue affected by the disorder as well as a method for identifying a compound to be tested for an ability to reduce immune rejection, comprising determining the amount of SOCS3 mRNA or SOCS3 protein in T-cell samples.

FURTHER INFORMATION CONTINUED FROM PCT/SA/ 210

Continuation of Box I.2

Claims Nos.: 57-67

Although claims 57-67 are directed to a method of treatment of the human/animal body, a search could have been carried out based on the compound and its alleged effect. Nevertheless, present claims 57-67 only relate to a compound defined by reference to a desirable characteristic or property, namely a compound able to reduce the level of STAT4 mRNA and/or maintaining or increasing the level of STAT6 mRNA. The claims cover all compounds having this characteristic or property, whereas the application does not provide any support within the meaning of Article 6 PCT and/or disclosure within the meaning of Article 5 PCT for such compounds. In the present case, the claims so lack support, and the application so lacks disclosure, that a meaningful search over the whole of the claimed scope is impossible. Independent of the above reasoning, the claims also lack clarity (Article 6 PCT). An attempt is made to define the compound by reference to a result to be achieved. Again, this lack of clarity in the present case is such as to render a meaningful search over the whole of the claimed scope impossible. Consequently, no search has been carried out for these claims.

The applicant's attention is drawn to the fact that claims, or parts of claims, relating to inventions in respect of which no international search report has been established need not be the subject of an international preliminary examination (Rule 66.1(e) PCT). The applicant is advised that the EPO policy when acting as an International Preliminary Examining Authority is normally not to carry out a preliminary examination on matter which has not been searched. This is the case irrespective of whether or not the claims are amended following receipt of the search report or during any Chapter II procedure.

INTERNATIONAL SEARCH REPORT

Information on patent family members

In national Application No

PCT/US 01/12131

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PCT/US 01/12131

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